

Factor IX19. GlycoPEGylation of Factor IX produced in CHO cells

This example sets forth the preparation of asialoFactor IX and its sialylation with CMP-sialic acid-PEG.

- 5 **Desialylation of rFactor IX.** A recombinant form of Coagulation Factor IX (rFactor IX) was made in CHO cells. 6000 IU of rFactor IX were dissolved in a total of 12 mL USP H₂O. This solution was transferred to a Centricon Plus 20, PL-10 centrifugal filter with another 6 mL USP H₂O. The solution was concentrated to 2 mL and then diluted with 15 mL 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 5 mM CaCl₂, 0.05% NaN₃ and then reconcentrated.
- 10 The dilution/concentration was repeated 4 times to effectively change the buffer to a final volume of 3.0 mL. Of this solution, 2.9 mL (about 29 mg of rFactor IX) was transferred to a small plastic tube and to it was added 530 mU α 2-3,6,8-Neuraminidase– agarose conjugate (*Vibrio cholerae*, Calbiochem, 450 μ L). The reaction mixture was rotated gently for 26.5 hours at 32 °C. The mixture was centrifuged 2 minutes at 10,000 rpm and the supernatant
- 15 was collected. The agarose beads (containing neuraminidase) were washed 6 times with 0.5 mL 50 mM Tris-HCl pH 7.12, 1 M NaCl, 0.05% NaN₃. The pooled washings and supernatants were centrifuged again for 2 minutes at 10,000 rpm to remove any residual agarose resin. The pooled, desialylated protein solution was diluted to 19 mL with the same buffer and concentrated down to ~ 2 mL in a Centricon Plus 20 PL-10 centrifugal filter. The
- 20 solution was twice diluted with 15 mL of 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 0.05% NaN₃ and reconcentrated to 2 mL. The final desialylated rFactor IX solution was diluted to 3 mL final volume (~10 mg/mL) with the Tris Buffer. Native and desialylated rFactor IX samples were analyzed by IEF-Electrophoresis. Isoelectric Focusing Gels (pH 3-7) were run using 1.5 μ L (15 μ g) samples first diluted with 10 μ L Tris buffer and mixed with 12 μ L
- 25 sample loading buffer. Gels were loaded, run and fixed using standard procedures. Gels were stained with Colloidal Blue Stain (Figure 154), showing a band for desialylated Factor IX.

- Preparation of PEG (1 kDa and 10 kDa)-SA-Factor IX.** Desialylated rFactor-IX (29 mg, 3 mL) was divided into two 1.5 mL (14.5 mg) samples in two 15 mL centrifuge
- 30 tubes. Each solution was diluted with 12.67 mL 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 0.05% NaN₃ and either CMP-SA-PEG-1k or 10k (7.25 μ mol) was added. The tubes were

inverted gently to mix and 2.9 U ST3Gal3 (326 μ L) was added (total volume 14.5 mL). The tubes were inverted again and rotated gently for 65 hours at 32 °C. The reactions were stopped by freezing at -20 °C. 10 μ g samples of the reactions were analyzed by SDS-PAGE. The PEGylated proteins were purified on a Toso Haas Biosep G3000SW (21.5 x 30 cm, 13 μ m) HPLC column with Dulbecco's Phosphate Buffered Saline, pH 7.1 (Gibco), 6 mL/min. The reaction and purification were monitored using SDS Page and IEF gels. Novex Tris-Glycine 4-20% 1 mm gels were loaded with 10 μ L (10 μ g) of samples after dilution with 2 μ L of 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.05% NaN₃ buffer and mixing with 12 μ L sample loading buffer and 1 μ L 0.5 M DTT and heated for 6 minutes at 85 °C. Gels were stained with Colloidal Blue Stain (Figure 155) showing a band for PEG (1 kDa and 10 kDa)-SA-Factor IX.

20. Direct Sialyl-GlycoPEGylation of Factor IX

This example sets forth the preparation of sialyl-PEGylation of Factor IX without prior sialidase treatment.

Sialyl-PEGylation of Factor-IX with CMP-SA-PEG-(10 kDa). Factor IX (1100 IU), which was expressed in CHO cells and was fully sialylated, was dissolved in 5 mL of 20 mM histidine, 520 mM glycine, 2% sucrose, 0.05% NaN₃ and 0.01% polysorbate 80, pH 5.0. The CMP-SA-PEG-(10 kDa) (27 mg, 2.5 μ mol) was then dissolved in the solution and 1 U of ST3Gal3 was added. The reaction was complete after gently mixing for 28 hours at 32°C. The reaction was analyzed by SDS-PAGE as described by Invitrogen. The product protein was purified on an Amersham Superdex 200 (10 x 300 mm, 13 μ m) HPLC column with phosphate buffered saline, pH 7.0 (PBS), 1 mL/min. R_t = 9.5 min.

Sialyl-PEGylation of Factor-IX with CMP-SA-PEG-(20 kDa). Factor IX (1100 IU), which was expressed in CHO cells and was fully sialylated, was dissolved in 5 mL of 20 mM histidine, 520 mM glycine, 2% sucrose, 0.05% NaN₃ and 0.01% polysorbate 80, pH 5.0. The CMP-SA-PEG-(20 kDa) (50 mg, 2.3 μ mol) was then dissolved in the solution and CST-II was added. The reaction mixture was complete after gently mixing for 42 hours at 32°C. The reaction was analyzed by SDS-PAGE as described by Invitrogen.

The product protein was purified on an Amersham Superdex 200 (10 x 300 mm, 13 μ m) HPLC column with phosphate buffered saline, pH 7.0 (Fisher), 1 mL/min. R_t = 8.6 min.

21. Sialic Acid Capping of GlycoPEGylated Factor IX

5 This examples sets forth the procedure for sialic acid capping of sialyl-glycoPEGylated peptides. Here, Factor-IX is the exemplary peptide.

Sialic acid capping of N-linked and O-linked Glycans of Factor-IX-SA-PEG (10 kDa). Purified r-Factor-IX-PEG (10 kDa) (2.4 mg) was concentrated in a Centricon[®] Plus 20 PL-10 (Millipore Corp., Bedford, MA) centrifugal filter and the buffer was changed to 50
10 mM Tris-HCl pH 7.2, 0.15 M NaCl, 0.05% NaN₃ to a final volume of 1.85 mL. The protein solution was diluted with 372 μ L of the same Tris buffer and 7.4 mg CMP-SA (12 μ mol) was added as a solid. The solution was inverted gently to mix and 0.1 U ST3Gal1 and 0.1 U ST3Gal3 were added. The reaction mixture was rotated gently for 42 hours at 32 °C.

A 10 μ g sample of the reaction was analyzed by SDS-PAGE. Novex Tris-Glycine 4-
15 12% 1 mm gels were performed and stained using Colloidal Blue as described by Invitrogen. Briefly, samples, 10 μ L (10 μ g), were mixed with 12 μ L sample loading buffer and 1 μ L 0.5 M DTT and heated for 6 minutes at 85 °C (Figure 156, lane 4).

Factor VIIa

20 22. GlycoPEGylation of Recombinant Factor VIIa produced in BHK cells

This example sets forth the PEGylation of recombinant Factor VIIa made in BHK cells.

Preparation of Asialo-Factor VIIa. Recombinant Factor VIIa was produced in BHK cells (baby hamster kidney cells). Factor VIIa (14.2 mg) was dissolved at 1 mg/ml in
25 buffer solution (pH 7.4, 0.05 M Tris, 0.15 M NaCl, 0.001 M CaCl₂, 0.05% NaN₃) and was incubated with 300 mU/mL sialidase (*Vibrio cholera*)-agarose conjugate for 3 days at 32 °C. To monitor the reaction a small aliquot of the reaction was diluted with the appropriate buffer and an IEF gel performed according to Invitrogen procedures (Figure 157). The mixture was centrifuged at 3,500 rpm and the supernatant was collected. The resin was washed three
30 times (3 \times 2 mL) with the above buffer solution (pH 7.4, 0.05 M Tris, 0.15 M NaCl, 0.05% NaN₃) and the combined washes were concentrated in a Centricon-Plus-20. The remaining

solution was buffer exchanged with 0.05 M Tris (pH 7.4), 0.15 M NaCl, 0.05% NaN₃ to a final volume of 14.4 mL.

Preparation of Factor VIIa-SA-PEG (1 kDa and 10 kDa). The desialylation of Factor VIIa solution was split into two equal 7.2 ml samples. To each sample was added either CMP-SA-5-PEG(1 kDa) (7.4 mg) or CMP-SA-5-PEG(10 kDa) (7.4 mg). ST3Gal3 (1.58U) was added to both tubes and the reaction mixtures were incubated at 32°C for 96 hrs. The reaction was monitored by SDS-PAGE gel using reagents and conditions described by Invitrogen. When the reaction was complete, the reaction mixture was purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The combined fractions containing the product were concentrated at 4°C in Centricon-Plus-20 centrifugal filters (Millipore, Bedford, MA) and the concentrated solution reformulated to yield 1.97 mg (bicinchoninic acid protein assay, BCA assay, Sigma-Aldrich, St. Louis MO) of Factor VIIa-PEG. The product of the reaction was analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples were dialyzed against water and analyzed by MALDI-TOF. Figure 158 shows the MALDI results for native Factor VIIa. Figure 159 contains the MALDI results for Factor VIIa PEGylated with 1 kDa PEG where peak of Factor VIIa PEGylated with 1KDa PEG is evident. Figure 160 contains the MALDI results for Factor VIIa PEGylated with 10 kDa PEG where a peak for Factor VIIa PEGylated with 10 kDa PEG is evident. Figure 161 depicts the SDS-PAGE analysis of all of the reaction products, where a band for Factor VIIa-SA-PEG (10 kDa) is evident.

Follicle Stimulating Hormone (FSH)

23. GlycoPEGylation of human pituitary-derived FSH

This example illustrates the assembly of a conjugate of the invention. Follicle Stimulating Hormone (FSH) is desialylated and then conjugated with CMP-(sialic acid)-PEG.

Desialylation of Follicle Stimulating Hormone. Follicle Stimulating Hormone (FSH) (Human Pituitary, Calbiochem Cat No. 869001), 1 mg, was dissolved in 500 µL 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 5 mM CaCl₂. This solution, 375 µL, was transferred to a small plastic tube and to it was added 263 mU Neuraminidase II (*Vibrio cholerae*). The reaction mixture was shaken gently for 15 hours at 32 °C. The reaction mixture was added to

N-(*p*-aminophenyl)oxamic acid-agarose conjugate, 600 μ L, pre-equilibrated with 50 mM Tris-HCl pH 7.4, 150 mM NaCl and 0.05% NaN₃ and gently rotated 6.5 hours at 4 °C. The suspension was centrifuged for 2 minutes at 14,000 rpm and the supernatant was collected. The beads were washed 5 times with 0.5 mL of the buffer and all supernatants were pooled.

5 The enzyme solution was dialyzed (7000 MWCO) for 15 hours at 4 °C with 2 L of a solution containing 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃, and then twice for 4 hours at 4 °C into 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃. The solution was concentrated to 2 μ g/ μ L by Speed Vac and stored at -20 °C. Reaction samples were analyzed by IEF gels (pH 3-7) (Invitrogen) (Figure 162).

10 **Preparation of human pituitary-derived SA-FSH and PEG-SA-Follicle Stimulating Hormone.** Desialylated FSH (100 μ g, 50 μ L) and CMP-sialic acid or CMP-SA-PEG (1 kDa or 10 kDa) (0.05 μ mol) were dissolved in 13.5 μ L H₂O (adjusted to pH 8 with NaOH) in 0.5 mL plastic tubes. The tubes were vortexed briefly and 40 mU ST3Gal3 (36.5 μ L) was added (total volume 100 μ L). The tubes were vortexed again and shaken gently for

15 24 hours at 32 °C. The reactions were stopped by freezing at -80 °C. Reaction samples of 15 μ g were analyzed by SDS-PAGE (Figure 163), IEF gels (Figure 164) and MALDI-TOF. Native FSH was also analyzed by SDS-PAGE (Figure 165)

Analysis of SDS PAGE and IEF Gels of Reaction Products. Novex Tris-Glycine 8-16% 1 mm gels for SDS PAGE analysis were purchased from Invitrogen. 7.5 μ L (15 μ g) of FSH reaction samples were diluted with 5 μ L of 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.05% NaN₃ buffer, mixed with 15 μ L sample loading buffer and 1 μ L 9 M μ -mercaptoethanol and heated for 6 minutes at 85 °C. Gels were run as directed by Invitrogen and stained with Colloidal Blue Stain (Invitrogen).

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FSH samples (15 μ g) were diluted with 5 μ L Tris buffer and mixed with 15 μ L sample loading buffer (Figure 162). The samples were then applied to Isoelectric Focusing Gels (pH 3-7) (Invitrogen) (Figure 165). Gels were run and fixed as directed by Invitrogen and then stained with Colloidal Blue Stain.

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24. GlycoPEGylation of recombinant FSH produced recombinantly in CHO cells

This example illustrates the assembly of a conjugate of the invention. Desialylated FSH was conjugated with CMP-(sialic acid)-PEG.

5 **Preparation of recombinant Asialo-Follicle Stimulation Hormone.** Recombinant Follicle Stimulation Hormone (rFSH) produced from CHO was used in these studies. The 7,500 IU of rFSH was dissolved in 8 mL of water. The FSH solution was dialyzed in 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 5 mM CaCl₂ and concentrated to 500 µL in a Centricon Plus 20 centrifugal filter. A portion of this solution (400 µL) (~ 0.8 mg FSH) was transferred to a
10 small plastic tube and to it was added 275 mU Neuraminidase II (*Vibrio cholerae*). The reaction mixture was mixed for 16 hours at 32 °C. The reaction mixture was added to prewashed N-(*p*-aminophenyl)oxamic acid-agarose conjugate (800 µL) and gently rotated for 24 hours at 4 °C. The mixture was centrifuged at 10,000 rpm and the supernatant was collected. The beads were washed 3 times with 0.6 mL Tris-EDTA buffer, once with 0.4 mL
15 Tris-EDTA buffer and once with 0.2 mL of the Tris-EDTA buffer and all supernatants were pooled. The supernatant was dialyzed at 4 °C against 2 L of 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃ and then twice more against 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃. The dialyzed solution was then concentrated to 420 µL in a Centricon Plus 20 centrifugal filter and stored at -20 °C.

20 Native and desialylated rFSH samples were analyzed by SDS-PAGE and IEF (Figure 166). Novex Tris-Glycine 8-16% 1 mm gels were purchased from Invitrogen. Samples (7.5 µL, 15 µg) samples were diluted with 5 µL of 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.05% NaN₃ buffer, mixed with 15 µL sample loading buffer and 1 µL 9 M β-mercaptoethanol and heated for 6 minutes at 85 °C. Gels were run as directed by Invitrogen
25 and stained with Colloidal Blue Stain (Invitrogen). Isoelectric Focusing Gels (pH 3-7) were purchased from Invitrogen. Samples (7.5 µL, 15 µg) were diluted with 5 µL Tris buffer and mixed with 15 µL sample loading buffer. Gels were loaded, run and fixed as directed by Invitrogen. Gels were stained with Colloidal Blue Stain. Samples of native and desialylated FSH were also dialyzed against water and analyzed by MALDI-TOF.

30 **Sialyl-PEGylation of recombinant Follicle Stimulation Hormone.** Desialylated FSH (100 µg, 54 µL) and CMP-SA-PEG (1 kDa or 10 kDa) (0.05 µmol) were dissolved in 28

μL 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2 in 0.5 mL plastic tubes. The tubes were vortexed briefly and 20 mU of ST3Gal3 was added (total volume 100 μL). The tubes were vortexed again, mixed gently for 24 hours at 32 °C and the reactions stopped by freezing at -80 °C. Samples of this reaction were analyzed as described above by SDS-PAGE gels (Figure 167), IEF gels (Figure 168) and MALDI-TOF MS.

MALDI was also performed on the PEGylated rFSH. During ionization, SA-PEG is eliminated from the N-glycan structure of the glycoprotein. Native FSH gave a peak at 13928; AS-rFSH (13282); resialylated r-FSH (13332); PEG1000-rFSH (13515; 14960 (1); 16455 (2); 17796 (3); 19321 (4)); and PEG 10000 (23560 (1); 34790 (2); 45670 (3); and 56760 (4)).

25. Pharmacokinetic Study of GlycoPEGylated FSH

This example sets forth the *in vivo* testing of the pharmacokinetic properties glycoPEGylated Follicle Stimulating Hormone (FSH) prepared according to the methods of the invention as compared to non-PEGylated FSH.

FSH, FSH-SA-PEG (1 kDa) and FSH-SA-PEG (10 kDa) were radioiodinated using standard conditions (Amersham Biosciences, Arlington Heights, IL) and formulated in phosphate buffered saline containing 0.1% BSA. After dilution in phosphate buffer to the appropriate concentration, each of the test FSH proteins (0.4 μg, each) was injected intravenously into female Sprague Dawley rats (250-300 g body weight) and blood drawn at time points from 0 to 80 hours. Radioactivity in blood samples was analyzed using a gamma counter and the pharmacokinetics analyzed using standard methods (Figure 169). FSH was cleared from the blood much more quickly than FSH-PEG(1 kDa), which in turn was clear somewhat more quickly than FSH-PEG(10 kDa).

26. Sertoli Cell Bioassay for *In Vitro* Activity of GlycoPEGylated FSH

This example sets forth a bioassay for follicle stimulating hormone (FSH) activity based on cultured Sertoli cells. This assay is useful to determine the bioactivity of FSH after glycan remodeling, including glycoconjugation.

This bioassay is based on the dose-response relationship that exists between the amount of estradiol produced when FSH, but not lutenizing hormone (LH), is added to

cultured Sertoli cells obtained from immature old rats. Exogenous testosterone is converted to 17 β -estradiol in the presence of FSH.

Seven to 10 days old Sprague-Dawley rats were used to obtain Sertoli cells. After sacrifice, testes were decapsulated and tissue was dispersed by incubation in collagenase (1 mg/ml), trypsin (1mg/ml), hyaluronidase (1 mg/ml) and DNases (5 μ g/ml) for 5 to 10 min. The tubule fragments settled to the bottom of the flask and were washed in PBS (1x). The tubule fragments were reincubated for 20 min with a media containing the same enzymes: collagenase (1 mg/ml), trypsin (1mg/ml), hyaluronidase (1 mg/ml) and DNases (5 μ g/ml).

The tubule fragments were homogenized and plated into a 24 well plate in a serum free media. 5 x 10⁵ cells were dispersed per well. After 48h incubation at 37° C and 5% CO₂, fresh media was added to the cells. Composition of the serum free media: DMEM (1 vol), Ham's F10 nutrient mixture (1 vol), insulin 1 μ g/ml, Transferrin 5 μ g/ml, EGF 10 ng/ml, T4 20 pg/ml, Hydrocortisone 10⁻⁸ M, Retinoic acid 10⁻⁶ M.

The stimulation experiment consists of a 24 hour incubation with standard FSH or samples at 37°C and 5% CO₂. The mean intra-assay coefficient of variation is 9% and the mean inter-assay coefficient of variation is 11%.

The 17B-estradiol Elisa Kit DE2000 (R&D Systems, Minneapolis, MN) was used to quantify the level of estradiol after incubation with FSH, FSH-SA-PEG (1 kDa) and FSH-SA-PEG (10 kDa).

The procedure was as follows: 100 μ l of Estradiol Standard (provided with kit and prepared as per instructions with kit) or sample was pipetted into wells of 17B-estradiol Elisa plate(s); 50 μ l of 17B-estradiol Conjugate (provided with kit, prepared as per instructions with kit) was added to each well; 50 μ l of 17B-estradiol antibody solution (provided with kit and prepared as per instructions with kit) was added to each well; plates were incubated for 2 hour at room temperature at 200 rpm; the liquid was aspirated from each well; the wells were washed 4 times using the washing solution; all the liquid was removed from the wells; 200 μ l of pNPP Substrate (provided with kit and prepared as per instructions with kit) was added to all wells and incubated for 45 min; 50 μ l of Stop solution (provided with kit and prepared as per instructions with kit) was added and the plates were read it at 405 nm (Figure 170).

While FSH-PEG(10 kDa) exhibited a modest stimulation of Sertoli cells, at 1 μ g/ml, FSH-PEG(1 kDa) stimulated Sertoli cells up to 50% more than unPEGylated FSH.

27. Steelman-Pohley Bioassay of *In Vivo* Activity of GlycoPEGylated FSH

In this example, the Steelman-Pohley bioassay (Steelman and Pohley, 1953, Endocrinology 53:604-615) was used to determine the *in vivo* activity of glycoPEGylated FSH. The Steelman-Pohley assay uses the change in ovary weight of a rat to measure the *in vivo* activity of FSH that is coinjected with human chorionic gonadotropin.

The Steelman-Pohley bioassay was performed according to the protocol described in Christin-Maitre et al. (2000, Methods 21:51-57). Seventy female Sprague-Dawley Rats (Charles River Laboratories, Wilmington, MA), aged 21 to 22 days, were housed in the testing facility for at least 5 days before the beginning the assay procedure. Throughout the procedure, the animal room was climate controlled at 18 to 26°C, 30 to 70% relative humidity, and 12 hr. artificial light/12 hr. dark. All animals were fed Certified Rodent Chow (Harlan Teklad, Madison WI) or the equivalent, and water, both *ad libitum*. Animal procedures were performed at Calvert Preclinical Services, Inc. (Olyphant, PA).

Recombinant FSH was expressed in CHO cells, purified by standard techniques and glycoPEGylated with PEG (1 kDa). The rats were divided into seven test groups, with ten animals per group. On days -1 and 0, animals of all groups were subcutaneously injected with 20 I.U. of human chorionic gonadotropin (HCG) in 0.5 ml of 0.9 % NaCl. On days 1, 2 and 3, the control animals were subcutaneously injected with a dose of 0.5 ml containing 20 I.U. HCG in 0.9% NaCl, while in the other groups, the HCG dose was augmented with either rFSH or rFSH-SA-PEG (1 kDa) at either 0.14 µg, 0.4 µg or 1.2 µg per dose. On day 4, the animals were euthanized by CO₂ inhalation. The ovaries were removed, trimmed and weighted. The average ovary weight was determined for each group.

Figure 171 presents the average ovary weight of the test groups on day 4. The groups receiving HCG alone (control) or the low dose (0.14 µg) of either rFSH or rFSH-SA-PEG (1 kDa) had ovary weights that were roughly equivalent. The groups receiving the medium (0.4 µg) or high (1.2 µg) doses of rFSH or rFSH-SA-PEG (1 kDa) had ovary weights roughly twice that of the control group. At the medium dose (0.4 µg), the glycoPEGylated rFSH had roughly the same *in vivo* activity (as determined by ovary weight) as the unPEGylated rFSH.

At the high dose (1.2 µg), the glycoPEGylated rFSH had somewhat higher *in vivo* activity than the unPEGylated rFSH.

G-CSF

5 28. GlycoPEGylation of G-CSF produced in CHO cells

Preparation of Asialo-Granulocyte-Colony Stimulation Factor (G-CSF). G-CSF produced in CHO cells is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 5 mM CaCl₂ and concentrated to 500 µL in a Centricon Plus 20 centrifugal filter. The solution is incubated with 300 mU/mL Neuraminidase II (*Vibrio cholerae*) for 16
10 hours at 32 °C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel performed. The reaction mixture is then added to prewashed N-(*p*-aminophenyl)oxamic acid-agarose conjugate (800 µL/mL reaction volume) and the washed beads gently rotated for 24 hours at 4 °C. The mixture is centrifuged at 10,000 rpm and the supernatant was collected. The beads are washed 3 times with Tris-EDTA buffer,
15 once with 0.4 mL Tris-EDTA buffer and once with 0.2 mL of the Tris-EDTA buffer and all supernatants are pooled. The supernatant is dialyzed at 4 °C against 50 mM Tris –HCl pH 7.4, 1 M NaCl, 0.05% NaN₃ and then twice more against 50 mM Tris –HCl pH 7.4, 1 M NaCl, 0.05% NaN₃. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter and stored at –20 °C. The conditions for the IEF gel were run according to
20 the procedures and reagents provided by Invitrogen. Samples of native and desialylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of G-CSF-(alpha2,3)-Sialyl-PEG. Desialylated G-CSF was dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of ST3Gal1 at 32°C for 2 days.
25 To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas
30 G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis

according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of G-CSF-(alpha2,8)-Sialyl-PEG. G-CSF produced in CHO cells, which contains an alpha2,3-sialylated O-linked glycan, is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of CST-II at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction has CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of G-CSF-(alpha2,6)-Sialyl-PEG. G-CSF, containing only O-linked GalNAc, is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of ST6GalNAcI or II at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction has CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

G-CSF produced in CHO cells was treated with Arthrobacter sialidase and was then purified by size exclusion on Superdex75 and was treated with ST3Gal1 or ST3 Gal2 and then with CMP-SA-PEG 20Kda. The resulting molecule was purified by ion exchange and

gel filtration and analysis by SDS PAGE demonstrated that the PEGylation was complete. This is the first demonstration of glycoPEGylation of an O-linked glycan.

Glucocerebrosidase

5 29. Glucocerebrosidase-mannose-6-phosphate produced in CHO cells

This example sets forth the procedure to glycoconjugate mannose-6-phosphate to a peptide produced in CHO cells such as glucocerebrosidase.

Preparation of asialo-glucoceramide. Glucocerebrosidase produced in CHO cells is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, and is
10 incubated with 300 mU/mL sialidase-agarose conjugate for 16 hours at 32 °C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel and SDS-PAGE performed according to Invitrogen procedures. The mixture is centrifuged at 10,000 rpm and the supernatant is collected. The beads are washed 3 times with Tris-EDTA buffer, once with 0.4 mL Tris-EDTA buffer, and once with 0.2 mL of the Tris-EDTA buffer.
15 All supernatants are pooled. The supernatant is dialyzed at 4 °C against 50 mM Tris-HCl pH 7.4, 1 M NaCl, 0.05% NaN₃ and then twice more against 50 mM Tris-HCl pH 7.4, 1 M NaCl, 0.05% NaN₃. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed
20 against water and analyzed by MALDI-TOF MS.

Preparation of Glucocerebrosidase-SA-linker-Mannose-6-phosphate (procedure

1). Asialo-glucocerebrosidase from above is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-linker-Man-6-phosphate and 0.1 U/mL of ST3Gal3 at 32°C for 2 days. To monitor the
25 incorporation of sialic acid-linker-Man-6-phosphate, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas TSK-Gel-3000 analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. When the reaction is complete, the reaction mixture is purified
30 using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using

SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Glucocerebrosidase-SA-linker-Mannose-6-phosphate (procedure 2). Glucocerebrosidase, produced in CHO but incompletely sialylated, is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-linker-Man-6-phosphate and 0.1 U/mL of ST3Gal3 at 32°C for 2 days. To monitor the incorporation of sialic acid-linker-Man-6-phosphate, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas TSK-Gel-3000 analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

30. Glucocerebrosidase-transferrin

This example sets forth the procedures for the glycoconjugation of proteins, and in particular, transferrin is glycoconjugated to glucocerebrosidase. The GlcNAc-ASN structures are created on glucoceraminidase, and Transferrin-SA-Linker-Gal-UDP is conjugated to GNDF GlcNAc-ASN structures using galactosyltransferase.

Preparation of GlcNAc-glucocerebrosidase (Cerezyme™). Cerezyme™ (glucocerebrosidase) produced in CHO cells is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, and is incubated with 300 mU/mL Endo-H-agarose conjugate for 16 hours at 32 °C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel and SDS-PAGE performed according to Invitrogen procedures. The mixture is centrifuged at 10,000 rpm and the supernatant is collected. The beads are washed 3 times with Tris-EDTA buffer, once with 0.4 mL Tris-EDTA buffer and once with 0.2 mL of the Tris-EDTA buffer and all supernatants are pooled. The supernatant is dialyzed at 4 °C against 50 mM Tris-HCl pH 7.4, 1 M NaCl, 0.05% NaN₃ and then twice

more against 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Transferrin-SA-Linker-Gal-glucocerebrosidase. Transferrin-SA-Linker-Gal-UDP from above is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl₂, 0.05% NaN₃, pH 7.2. The solution is incubated with 2.5 mg/mL GlcNAc-glucocerebrosidase and 0.1 U/mL of galactosyltransferase at 32°C for 2 days. To monitor the incorporation of glucocerebrosidase, the peptide is separated by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1) and the product detected by UV absorption. The reaction mixture is then purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

GM-CSF

31. Generation and PEGylation of GlcNAc-ASN Structures: GM-CSF produced in *Saccharomyces*

This example sets forth the preparation of Tissue-type Activator with PEGylated GlcNAc-Asn structures.

Recombinant GM-CSF expressed in yeast is expected to contain 2 N-linked and 2 O-linked glycans. The N-linked glycans should be of the branched mannan type. This recombinant glycoprotein is treated with an endoglycosidase from the group consisting of endoglycosidase H, endoglycosidase-F1, endoglycosidase-F2, endoglycosidase-F3, endoglycosidase-M either alone or in combination with mannosidases I, II and III to generate GlcNAc nubs on the asparagine (Asn) residues on the peptide/protein backbone.

The GlcNAc-Asn structures on the peptide/protein backbone is then be modified with galactose or galactose-PEG using UDP-galactose or UDP-galactose-6-PEG, respectively, and a galactosyltransferase such as GalT1. In one case the galactose-PEG is the terminal residue.

In the second case the galactose is further modified with SA-PEG using a CMP-SA-PEG donor and a sialyltransferase such as ST3GalIII. In another embodiment the GlcNAc-Asn structures on the peptide/protein backbone can be galactosylated and sialylated as described above, and then further sialylated using CMP-SA-PEG and an α 2,8-sialyltransferase such as the enzyme encoded by the *Campylobacter jejuni* cst-II gene.

Herceptin™

32. Glycoconjugation of mithramycin to Herceptin™

This example sets forth the procedures to glycoconjugate a small molecule, such as mithramycin to Fc region glycans of an antibody molecule produced in mammalian cells. Here, the antibody Herceptin™ is used, but one of skill in the art will appreciate that the method can be used with many other antibodies.

Preparation of Herceptin™-Gal-linker-mithramycin. Herceptin™ is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl₂, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM UDP-galactose-linker-mithramycin and 0.1 U/mL of galactosyltransferase at 32°C for 2 days to introduce the mithramycin in the Fc region glycans. To monitor the incorporation of galactose, a small aliquot of the reaction has ¹⁴C-galactose-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The fractions containing product are combined, concentrated, buffer exchanged and then freeze-dried. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Interferon α and Interferon β 33. GlycoPEGylation of Proteins expressed in Mammalian or Insect Systems:
EPO, Interferon α and Interferon β

5 This example sets forth the preparation of PEGylated peptides that are expressed in mammalian and insect systems.

Preparation of acceptor from mammalian expression systems. The peptides to be glycoPEGylated using CMP-sialic acid PEG need to have glycans terminating in galactose. Most peptides from mammalian expression systems will have terminal sialic acid that first
10 needs to be removed.

Sialidase digestion. The peptide is desialylated using a sialidase. A typical procedure involves incubating a 1 mg/mL solution of the peptide in Tris-buffered saline, pH 7.2, with 5 mM CaCl_2 added, with 0.2 U/mL immobilized sialidase from *Vibrio cholera* (Calbiochem) at 32°C for 24 hours. Microbial growth can be halted either by sterile filtration
15 or the inclusion of 0.02% sodium azide. The resin is then removed by centrifugation or filtration, and then washed to recover entrapped peptide. At this point, EDTA may be added to the solution to inhibit any sialidase that has leached from the resin.

Preparation from insect expression systems. EPO, interferon-alpha, and interferon-beta may also be expressed in non-mammalian systems such as yeast, plants, or
20 insect cells. The peptides to be glycoPEGylated using CMP-sialic acid PEG need to have glycans terminating in galactose. The majority of the N-glycans on peptides expressed in insect cells, for example, are the trimannosyl core. These glycans are first built out to glycans terminating in galactose before they are acceptors for sialyltransferase.

Building acceptor glycans from trimannosyl core. Peptide (1 mg/mL) in Tris-
25 buffered saline, pH 7.2, containing 5 mM MnCl_2 , 5 mM UDP-glcNAc, 0.05 U/mL GLCNACT I, 0.05 U/mL GLCNACT II, is incubated at 32°C for 24 hours or until the reaction is substantially complete. Microbial growth can be halted either by sterile filtration or the inclusion of 0.02% sodium azide. After buffer exchange to remove UDP and other small molecules, UDP-galactose and MnCl_2 are each added to 5 mM, galactosyltransferase is
30 added to 0.05 U/mL, and is incubated at 32°C for 24H or until the reaction is substantially

complete. Microbial growth can be halted either by sterile filtration or the inclusion of 0.02% sodium azide. The peptides are then ready for glycoPEGylation.

Building O-linked glycans. A similar strategy may be employed for interferon alpha to produce enzymatically the desired O-glycan Gal-GalNAc. If necessary, GalNAc linked to serine or threonine can be added to the peptide using appropriate peptide GalNAc transferases (e.g. GalNAc T1, GalNAc T2, T3, T4, etc.) and UDP-GalNAc. Also, if needed, galactose can be added using galactosyltransferase and UDP-galactose.

GlycoPEGylation using sialyltransferase. The glycopeptides (1 mg/mL) bearing terminal galactose in Tris buffered saline + 0.02% sodium azide are incubated with CMP-SA-PEG (0.75 mM) and 0.4 U/mL sialyltransferase (ST3Gal3 or ST3Gal4 for N-glycans on EPO and interferon beta; ST3Gal4, or ST3Gal1 for O-glycans on interferon alpha) at 32°C for 24 hours. Other transferases that may work include the 2,6 sialyltransferase from *Photobacterium damsella*. The acceptor peptide concentration is most preferably in the range of 0.1 mg/mL up to the solubility limit of the peptide. The concentration of CMP-SA-PEG should be sufficient for there to be excess over the available sites, but not so high as to cause peptide solubility problems due to the PEG, and may range from 50 µM up to 5 mM, and the temperature may range from 2°C up to 40°C. The time required for complete reaction will depend on the temperature, the relative amounts of enzyme to acceptor substrate, the donor substrate concentration, and the pH.

34. GlycoPEGylation of Interferon α produced in CHO cells

Preparation of Asialo-Interferon α . Interferon alpha produced from CHO cells is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, 5 mM CaCl₂ and concentrated to 500 µL in a Centricon Plus 20 centrifugal filter. The solution is incubated with 300 mU/mL Neuraminidase II (*Vibrio cholerae*) for 16 hours at 32 °C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel performed. The reaction mixture is then added to prewashed N-(*p*-aminophenyl)oxamic acid-agarose conjugate (800 µL/mL reaction volume) and the washed beads gently rotated for 24 hours at 4 °C. The mixture is centrifuged at 10,000 rpm and the supernatant was collected. The beads are washed 3 times with Tris-EDTA buffer, once with 0.4 mL Tris-EDTA buffer and once with 0.2 mL of the Tris-EDTA buffer and all

supernatants were pooled. The supernatant is dialyzed at 4 °C against 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃ and then twice more against 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃. The dialyzed solution is then concentrated using a Centricon Plus 20 centrifugal filter and stored at -20 °C. The conditions for the IEF gel are run according to the procedures and reagents provided by Invitrogen. Samples of native and desialylated G-CSF are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Interferon-alpha-(alpha2,3)-Sialyl-PEG. Desialylated interferon-alpha is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of ST3Gal1 at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and desialylated Interferon-alpha are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Interferon-alpha-(alpha2,8)-Sialyl-PEG. Interferon-alpha produced in CHO, which contains an alpha2,3-sialylated O-linked glycan, is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of CST-II at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction has CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis

according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated interferon-alpha are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Interferon-alpha-(alpha2,6)-Sialyl-PEG. Interferon-alpha, containing only O-linked GalNAc, was dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM CMP-sialic acid-PEG and 0.1 U/mL of ST6GalNAcI or II at 32°C for 2 days. To monitor the incorporation of sialic acid-PEG, a small aliquot of the reaction had CMP-SA-PEG-fluorescent ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The fluorescent label incorporation into the peptide is quantitated using an in-line fluorescent detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples of native and PEGylated interferon-alpha are dialyzed against water and analyzed by MALDI-TOF MS.

35. GlycoPEGylation of Interferon-β-1a with PEG (10 kDa) and PEG (20 kDa)

This example illustrates a procedure PEGylate Interferon-β with either PEG (10 kDa) or PEG (20 kDa).

Briefly, Interferon-β-1a (INF-β) was obtained from Biogen (Avonex™). The INF-β was first purified by Superdex-75 chromatography. The INF-β was then desialylated with *Vibrio cholerae* sialidase. The INF-β was then PEGylated with SA-PEG (10 kDa) or SA-PEG (20 kDa) and purified with Superdex-200 chromatography.

Superdex-75 chromatography purification. INF-β (150 μg) was applied to a Superdex-75 column (Amersham Biosciences, Arlington Heights, IL) and eluted with PBS with 0.5 M NaCl, 0.02 Tween-20, 20 mM histidine and 10% glycerol. The eluant was monitored for absorbance at 280 nm (Figure 172A and 172B) and fractions were collected. Peaks 4 and 5 were pooled, concentrated in an Amicon Ultra 15 spin filter (Millipore, Billerica, MA), and the buffer was exchanged to TBS with 5 mM CaCl₂, 0.02% Tween-20, 20 mM histidine and 10% glycerol.

Sialidase Reaction. The INF- β was then desialylated with *Vibrio cholera* sialidase (70 mU/ml, CALBIOCHEM®, EMD Biosciences, Inc., San Diego, CA) on agarose in TBS with 5 mM CaCl₂, 0.02% Tween-20, 20 mM histidine and 10% glycerol. The reaction was carried out at 32°C for 18 hours. The INF- β was removed from the agarose with a 0.22 μ m Spin-X™ filter (Corning Technology, Inc., Norcross, GA). Figure 173A depicts the MALDI analysis of glycans released from native INF- β . The native INF- β has many glycoforms containing terminal sialic acid moieties. Figure 173B depicts the MALDI analysis of glycans released from desialylated INF- β . The desialylated INF- β has primarily one glycoform which is bi-antennary with terminal galactose moieties.

Lectin Dot-Blot Analysis of Sialylation. Samples of the INF- β from the desialidase reaction were dot-blotted onto nitrocellulose and then blocked with Tris buffered saline (TBS: 0.05M Tris, 0.15M NaCl, pH 7.5) and DIG kit (glycan differentiation kit available from Roche #1 210 238) blocking buffer. Some of the blots were incubated with *Maackia amurensis* agglutinin (MAA) labeled with digoxigenin (DIG) (Roche Applied Science, Indianapolis, IL) to detect α 2,3-sialylation of INF- β . These blots were washed with TBS then incubated with anti-digoxigenin antibody labeled with alkaline phosphatase, then washed again with TBS and developed with NBT/X-phosphate solution, wherein NBT is 4-nitro blue tetrazolium chloride and X-phosphate is 5-bromo-4-chloro-3-indoyl phosphate. The left side of Figure 174 depicts the results of the MAA blot of INF- β after the desialylation reaction. The INF- β is partially desialylated, as indicated by the decrease in dot development as compared to native INF- β in the desialylated samples.

Other blots were incubated with *Erthrina cristagalli* lectin (ECL) labeled with biotin (Vector Laboratories, Burlingame, CA) to detect exposed galactose residues on INF- β . After incubation with 2.5 μ g/ml ECL, the blots were washed in TBS and incubated with streptavidin labeled with alkaline phosphatase. The blots were then washed again and developed. The right side of Figure 174 depicts the ECL blot after development. The increased intensity of the dot of desialylated INF- β as compared to the native INF- β indicate more exposed galactose moieties and therefore extensive desialylation.

PEGylation of Desialylated INF- β with SA-PEG (10 kDa). Desialylated INF- β (0.05 mg/ml) was PEGylated with ST3Gal3 (50 mU/ml) and CMP-SA-PEG (10 kDa) (250

μM) in an appropriate buffer of TBS + 5 mM CaCl₂, 0.02% Tween 20, 20 mM histidine, 10% glycerol for 50 hours at 32°C. Figure 175 depicts the SDS-PAGE analysis of the reaction products showing PEGylated INF-β at approximately 98 kDa.

PEGylation of Desialylated INF-β with SA-PEG (20 kDa). Desialylated INF-β (0.5 mg/ml) was PEGylated with ST3Gal3 (170 mU/ml) and CMP-SA-PEG (20 kDa) in an appropriate buffer of TBS + 5 mM CaCl₂, 0.02% Tween 20, 20 mM histidine, 10% glycerol for 50 hours at 32°C. Figure 176 depicts the SDS-PAGE analysis the products of the PEGylation reaction. The PEGylated INF-β has many higher molecular weight bands not found in the unmodified INF-β indicating extensive PEGylation.

Superdex-200 Purification of INF-β PEGylated with PEG (10 kDa). The products of the PEGylation reaction were separated on a Superdex-200 column (Amersham Biosciences, Arlington Heights, IL) in PBS with 0.5 NaCl, 0.02 Tween-20, 20 mM histidine and 10% glycerol at 1ml/min and 30 cm/hr flow. The eluant was monitored for absorbance at 280 nm (Figure 177) and fractions were collected. Peaks 3 and 4 were pooled and concentrated in an Amicon Ultra 15 spin filter.

Bioassay of INF-β PEGylated with PEG (10 kDa).

The test is inhibition of the proliferation of the lung carcinoma cell line, A549. The A549 cell line are lung carcinoma adherent cells growing in RPMI + 10% FBS at 37°C 5% CO₂. They can be obtained from ATCC # CCL-185. Wash the cells with 10 ml of PBS and remove the PBS. Add 5 ml of trypsin, incubate for 5 minutes at room temperature or 2 minutes at 37°C. When the cells are detached resuspend into 25 ml of media and count the cells. Dilute the cells at a concentration of 10000 cells/ml and add 200 ul / well (96 wells plate). Incubate for 4 hours at 37°C 5% CO₂. Prepare 1 ml of IFN B at a concentration of 0.1 ug/ml. Filter it under the hood with a 0.2 um filter. Add 100 ul per well (8 replicates = 1 lane). Incubate for 3 days (do not let the cells go to confluence). Remove 200 ul of media (only 100ul per well left). Add 25 μl of MTT (Sigma) (5 mg/ml filtered 0.22μm). Incubate for 4 hours at 37°C and 5% CO₂. Aspirate the media gently and add 100 μl of a mixture of isopropanol (100 ml and 6N HCl. Aspirate up and down to homogenize the crystal violet. Read OD 570nm (remove the background at 630 or 690 nm).

Figure 178 depicts the results of the bioassay of the peaks containing INF- β PEGylated with PEG (10 kDa) as eluted from the Superdex-200 column.

Superdex-200 Purification of INF- β PEGylated with PEG (20 kDa). The products of the PEG (20 kDa) PEGylation reaction were separated on a Superdex-200 column (Amersham Biosciences, Arlington Heights, IL) in PBS with 0.5 NaCl, 0.02 Tween-20, 20 mM histidine and 10% glycerol at 1 ml/min flow. The eluant was monitored for absorbance at 280 nm (Figure 179) and fractions were collected. Peak 3 contained most of the INF- β PEGylated with PEG (20 kDa).

Endotoxin test of INF- β PEGylated with PEG (20 kDa).

Limulus Lysate Test was performed, BioWhittaker # 50-647U

Table 24. Results of the endotoxin test of INF- β PEGylated with PEG (20 kDa).

	Concentration		
INF- β with PEG (20 kDa)	10 EU/ml	0.06 mg/ml	0.16 EU/ μ g
INF- β with PEG (20 kDa)	1 EU/ml	0.07 mg/ml	0.014 EU/ μ g
Native INF- β	40 EU/ml	0.1 mg/ml	0.4 EU/ μ g

RemicadeTM

36. GlycoPEGylation of RemicadeTM antibody

This example sets forth the procedure to glycoPEGylate a recombinant antibody molecule by introducing PEG molecules to the Fc region glycans. Here RemicadeTM, a TNF-R:IgG Fc region fusion protein, is the exemplary peptide.

Preparation of RemicadeTM-Gal-PEG (10 kDa). RemicadeTM is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl₂, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM UDP-galactose-PEG (10 kDa) and 0.1 U/mL of galactosyltransferase at 32°C for 2 days to introduce the PEG in the Fc region glycans. To monitor the incorporation of galactose, a small aliquot of the reaction has ¹⁴C-galactose-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The fractions containing product are combined, concentrated, buffer exchanged and then freeze-dried. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Rituxan™

37. Glycoconjugation of geldanamycin to Rituxan™

This example sets forth the glycoconjugation of a small molecule, such as geldanamycin, to the Fc region glycans of an antibody produced in CHO cells, such as Rituxan™. Here, the antibody Rituxan™ is used, but one of skill in the art will appreciate that the method can be used with many other antibodies.

Preparation of Rituxan™-Gal-linker-geldanamycin. Rituxan™ is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl₂, 0.05% NaN₃, pH 7.2. The solution is incubated with 1 mM UDP-galactose-linker-geldanamycin and 0.1 U/mL of galactosyltransferase at 32°C for 2 days to introduce the geldanamycin in the Fc region glycans. To monitor the incorporation of galactose, a small aliquot of the reaction has ¹⁴C-galactose-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

When the reaction is complete, the reaction mixture is purified using a Toso Haas TSK-Gel-3000 preparative column using PBS buffer (pH 7.1) and collecting fractions based on UV absorption. The fractions containing product are combined, concentrated, buffer exchanged and then freeze-dried. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Rnase38. Remodeling high mannose N-glycans to hybrid and complex N-glycans:
Bovine pancreatic RNase

This example sets forth the preparation of bovine pancreas RNase with hybrid or
5 complex N-glycans. The high mannose N-linked glycans of the RNase are enzymatically
digested and elaborated to create hybrid N-linked glycans. Additionally, the high mannose
N-linked glycans of the RNase are enzymatically digested and elaborated to create complex
N-linked glycans.

High mannose structures of *N*-linked oligosaccharides in glycopeptides can be
10 modified to hybrid or complex forms using the combination of α -mannosidases and
glycosyltransferases. This example summarizes the results in such efforts using a simple *N*-
Glycan as a model substrate.

Ribonuclease B (RNaseB) purified from bovine pancreas (Sigma) is a glycopeptide
consisting of 124 amino acid residues. It has a single potential *N*-glycosylation site modified
15 with high mannose structures. Due to its simplicity and low molecular weight (13.7 kDa to
15.5 kDa), ribonuclease B is a good candidate to demonstrate the feasibility of the *N*-Glycan
remodeling from high mannose structures to hybrid or complex *N*-linked oligosaccharides.
The MALDI-TOF spectrum of RNaseB (Figure 180A) and HPLC profile for the
oligosaccharides cleaved from RNaseB by *N*-Glycanase (Figure 180B) indicated that, other
20 than a small portion of the non-modified peptide, the majority of *N*-glycosylation sites of the
peptide are modified with high mannose oligosaccharides consisting of 5 to 9 mannose
residues.

Conversion of high mannose N-Glycans to hybrid N-Glycans. High mannose *N*-
Glycans were converted to hybrid *N*-Glycans using the combination of α 1,2-mannosidase,
25 GlcNAcT-I (β -1,2-*N*-acetyl glucosaminyl transferase), GalT-I (β 1,4-galactosyltransferase) and
 α 2,3-sialyltransferase /or α 2,6-sialyltransferase as shown in Figure 181.

As an example, high mannose structures in RNaseB were successfully converted to
hybrid structures.

Man₅GlcNAc₂-R was obtained from Man₅₋₉GlcNAc₂-R catalyzed by a single α 1,2-
30 mannosidase cloned from *Trichoderma reesei* (Figure 182). RNase B (1 g, about 67 μ mol)
was incubated at 30°C for 45 hr with 15 mU of the recombinant *T. reesei* α 1,2-mannosidase

in MES buffer (50 mM, pH 6.5) in a total volume of 10 mL. Man₆₋₉GlcNAc₂-protein structures have been successfully converted to Man₅GlcNAc₂-protein with high efficiency by the recombinant mannosidase.

Alternately, Man₅GlcNAc₂-R was obtained from Man₅₋₉GlcNAc₂-R catalyzed by a single α 1,2-mannosidase purified from *Aspergillus saitoi* (Figure 183). RNase B (40 μ g, about 2.7 nmol) was incubated at 37°C for 42.5 hr with 25 μ U of the commercial *A. saitoi* α 1,2-mannosidase (Glyko or CalBioChem) in NaOAc buffer (100 mM, pH 5.0) in a total volume of 20 μ l. Man₆₋₉GlcNAc₂-protein structures were successfully converted to Man₅GlcNAc₂-protein by the commercially available mannosidase. However, a new peak corresponding to the GlcNAc-protein appears in the spectrum, indicating the possible contamination of endoglycosidase H in the preparation. Although several mammalian alpha-mannosidases were required to achieve this step, the fungal α 1,2-mannosidase was very efficient to remove all α 1,2-linked mannose residues.

GlcNAcT-I then added a GlcNAc residue to the Man₅GlcNAc₂-R (Figure 184). The reaction mixture after the *T. reesei* α 1,2-mannosidase reaction containing RNase B (600 μ g, about 40 nmol) was incubated with non-purified recombinant GlcNAcT-I (34 mU) in MES buffer (50 mM, pH 6.5) containing MnCl₂ (20 mM) and UDP-GlcNAc (5 mM) in a total volume of 400 μ l. at 37°C for 42 hr. A GlcNAc residue was quantitatively added to Man₅GlcNAc₂-protein by the recombinant GlcNAcT-I.

A Gal residue was then added using GalT 1 (Figure 185). The reaction mixture after the GnT-I reaction containing RNase B (120 μ g, about 8 nmol) was incubated at 37°C for 20 hr with 3.3 mU of the recombinant GalT-1 in Tris-HCl buffer (100 mM, pH 7.3) containing UDP-Gal (7.5 mM) and MnCl₂ (20 mM) in a total volume of 100 μ l. A Gal residue was added to about 98% of the GlcNAc-Man₅GlcNAc₂-protein by the recombinant GalT 1.

The next step was the addition of a sialic acid using an α 2,3-sialyltransferase or an α 2,6-sialyltransferase (Figure 186). As an example, ST3Gal III, an α 2,3-sialyltransferase was used. The reaction mixture after the GalT-1 reaction containing RNase B (13 μ g, about 0.87 nmol) was incubated at 37°C for 16 hr with 8.9 mU of recombinant ST3Gal III in Tris-HCl buffer (100 mM, pH 7.3) containing CMP-Sialic acid (5 mM) and MnCl₂ (20 mM) in a total volume of 20 μ l. A sialic acid residue was added to about 90% of the Gal-GlcNAc-

Man₅GlcNAc₂-protein by recombinant ST3Gal III using CMP-SA as the donor. The yield can be further improved by adjusting the reaction conditions.

For convenience, no purification or dialysis step was required after each reaction described above. More interesting, GalT 1 and ST3Gal III can be combined in a one-pot reaction. Similar yields were obtained as compared with the separate reactions. The reaction mixture after the GlcNAcT-I reaction containing RNase B (60 µg, about 4 nmol) was incubated at 37°C for 20 hr with 1.7 mU of recombinant GalT 1, 9.8 mU of recombinant ST3Gal III in Tris-HCl buffer (100 mM, pH 7.3) containing UDP-Gal (7.5 mM), CMP-sialic acid (5 mM) and MnCl₂ (20 mM) in a total volume of 60 µl.

As shown in Figure 187, SA-PEG (10 kDa) was successfully added to the RNaseB. The reaction mixture after the GalT-1 reaction containing RNase B (6.7 µg, about 0.45 nmol) was dialyzed against H₂O for 1 hour at room temperature and incubated at 37°C for 15.5 hours with 55 mU of the recombinant ST3Gal III in Tris-HCl buffer (50 mM, pH 7.3) containing CMP-SA-PEG (10 kDa) (0.25 mM) and MnCl₂ (20 mM) in a total volume of 20 µl. PEG-modified sialic acid residues were successfully added to the Gal-GlcNAc-Man₅GlcNAc₂-peptide by the recombinant ST3Gal III. The yield can be further improved by adjusting the reaction conditions.

Conversion of high mannose N-Glycans to complex N-Glycans. To achieve this conversion, a GlcNAcβ1,2Man₃GlcNAc₂-peptide intermediate is obtained. As shown in Figure 188, there are at least four feasible routes to carry out the reaction from Man₅GlcNAc₂-peptide to this intermediate:

Route I: The Man₅GlcNAc₂-peptide produced by the fungal α1,2 mannosidase is a substrate of GlcNAc transferase I (GlcNAcT-I, enzyme 2) which adds one GlcNAc. The terminal α1,3- and α1,6-linked mannose residues of GlcNAcMan₅GlcNAc₂-peptide is removed by Golgi α-mannosidase II (ManII, enzyme 5). This route is a part of the natural pathway for the processing of N-linked oligosaccharides carried out in higher organisms.

Route II: Two mannose residues are first removed by an α-mannosidase (enzyme 6), then a GlcNAc is added by GlcNAcT-I (enzyme 2). Other than its natural acceptor Man₅GlcNAc₂-R, GlcNAcT-I can also recognize Man₃GlcNAc₂-R as its substrate and add one GlcNAc to the mannose core structure to form GlcNAcMan₃GlcNAc₂-peptide.

Route III: The α 1,6-linked mannose is removed by an α 1,6-mannosidase, followed by the addition of GlcNAc by GlcNAcT-I and removal of the terminal α 1,3-linked mannose by an α 1,3-mannosidase. From the experimental data obtained, GlcNAcT-I can recognize this Man₄GlcNAc₂-peptide as acceptor and add one GlcNAc residue to form

5 GlcNAcMan₄GlcNAc₂-peptide.

Route IV: Similar to Route III, α 1,3-linked mannose is removed by an α 1,3-mannosidase, followed by GlcNAcT-I reaction. Then the terminal α 1,6-linked mannose can be removed by an α 1,6-mannosidase.

After the function of GlcNAcT-I (responsible for the addition of the GlcNAc β 1,2-linked to the α 1,3-mannose on the mannose core) and GlcNAcT-II (responsible for the addition of a second GlcNAc β 1,2-linked to the α 1,6-mannose on the mannose core), the GlcNAc₂Man₃GlcNAc₂-peptide can be processed by GalT 1 and sialyltransferase to form bi-antennary complex N- Glycans. Other GlcNAc transferases such as GlcNAcT-IV, GlcNAcT-V, and/or GlcNAcT-VI (Figure 188 and Figure 189) can also glycosylate the

10 linked to the α 1,3-mannose on the mannose core) and GlcNAcT-II (responsible for the addition of a second GlcNAc β 1,2-linked to the α 1,6-mannose on the mannose core), the GlcNAc₂Man₃GlcNAc₂-peptide can be processed by GalT 1 and sialyltransferase to form bi-antennary complex N- Glycans. Other GlcNAc transferases such as GlcNAcT-IV, GlcNAcT-V, and/or GlcNAcT-VI (Figure 188 and Figure 189) can also glycosylate the

15 GlcNAc₂Man₃GlcNAc₂-peptide. Additional glycosylation by the GalT 1 and sialyltransferases will form multi-antennary complex N-glycans. The enzyme GlcNAcT-III catalyzes the insertion of a bisecting GlcNAc, thus preventing the actions of ManII and subsequent action of transferases GlcNAcT-II, GlcNAcT-IV and GlcNAcT-V.

20 Tissue-Type Plasminogen Activator (TPA)

39. Fucosylation of TPA to create Sialyl Lewis X

This example sets forth the preparation of Tissue Tissue-type Plasminogen Activator (TPA) with N-linked sialyl Lewis X antigen.

Sialylation. TPA expressed in mammalian cells will often contain a majority of the glycans terminating in sialic acid, but to ensure complete sialylation, it would be beneficial to first perform an *in vitro* sialylation. TPA in a suitable buffer (most preferably between pH 5.5 and 9, for example Tris buffered saline, pH 7.2) is incubated with CMP sialic acid and sialyltransferase for a time sufficient to convert any glycans lacking sialic acid to sialylated species. Typical conditions would be 1 mg/mL TPA, 3 mM CMP sialic acid, 0.02 U/mL

25 glycans terminating in sialic acid, but to ensure complete sialylation, it would be beneficial to first perform an *in vitro* sialylation. TPA in a suitable buffer (most preferably between pH 5.5 and 9, for example Tris buffered saline, pH 7.2) is incubated with CMP sialic acid and sialyltransferase for a time sufficient to convert any glycans lacking sialic acid to sialylated species. Typical conditions would be 1 mg/mL TPA, 3 mM CMP sialic acid, 0.02 U/mL

30 ST3Gal3, 32°C for 24 hours. Microbial growth can be halted either by sterile filtration or the

inclusion of 0.02% sodium azide. The TPA concentration is most preferably in the range 0.1 mg/mL up to the solubility limit of the peptide. The concentration of CMP-SA should be sufficient for there to be excess over the available sites, and might range from 50 μ M up to 50 mM, and the temperature from 2°C up to 40°C. The time required for complete reaction will depend on the temperature, the relative amounts of enzyme to acceptor substrate, the donor substrate concentration, and the pH. Other sialyltransferases that may be capable of adding sialic acid in 2,3 linkage include ST3Gal4; microbial transferases could also be used.

Fucosylation. Typical conditions for fucosylation would be 1 mg/mL TPA, 3 mM GDP-fucose, 0.02 U/mL FTVI, 5 mM MnCl₂, 32°C for 24H in Tris buffered saline.

Microbial growth can be halted either by sterile filtration or the inclusion of 0.02% sodium azide. The TPA concentration is most preferably in the range 0.1 mg/mL up to the solubility limit of the peptide. The concentration of GDP-fucose should be sufficient for there to be excess over the available sites, and might range from 50 μ M up to 50 mM, and the temperature from 2°C up to 40°C. The time required for complete reaction will depend on the temperature, the relative amounts of enzyme to acceptor substrate, the donor substrate concentration, and the pH. Other fucosyltransferases that may be capable of making sialyl Lewis x include FTVII, FTV, FTIII, as well as microbial transferases could also be used.

40. Trimming of high mannose to tri-mannose core structure: Tissue-type Plasminogen Activator produced in CHO

This example sets forth the preparation of Tissue-type Plasminogen Activator with a trimannose core by trimming back from a high mannose glycan.

Tissue-type plasminogen activator (TPA) is currently produced in Chinese Hamster Ovary (CHO) cells and contains a low amount of high mannose N-linked oligosaccharide.

The mannoses can be trimmed down using a variety of the specific mannosidases. The first step is to generate Man5GlcNAc2(Fuc0-1) from Man9GlcNAc2(Fuc0-1). This can be done using mannosidase I. Then either GlcNAcT1 (GlcNAc transferase I) is used to make GlcNAc1Man5GlcNAc2(Fuc0-1) or Mannosidase III is used to make Man3GlcNAc2(Fuc0-1). From Man3GlcNAc2(Fuc0-1), GlcNAc1Man3GlcNAc2(Fuc0-1) can be produced using GlcNAcT1 or from GlcNAc1Man5GlcNAc2(Fuc0-1), GlcNAc1Man3GlcNAc2(Fuc0-1) can be produced using Mannosidase II. GlcNAc1Man3GlcNAc2(Fuc0-1) is then converted into

GlcNAc2Man3GlcNAc2(Fuc0-1) using GlcNAcTransferase II (GlcNAcTII). The two terminal GlcNAc residues are then galactosylated using GalTI and then sialylated with SA-PEG using ST3GalIII.

Conversely, TPA can be produce in yeast or fungal systems. Similar processing
5 would be required for fungal derived material.

41. Generation and PEGylation of GlcNAc-Asn structures: TPA produced in Yeast

This example sets forth the preparation of PEGylated GlcNAc-Asn structures on a
10 peptide such as TPA expressed in yeast.

Yeast expression is expected to result in a TPA which contains a single N-linked mannan-type structure. This recombinant glycoprotein is first treated with endoglycosidase H to generate GlcNAc structures on the asparagine (Asn) residues on the peptide.

The GlcNAc-Asn structures on the peptide/protein backbone are then be modified
15 with galactose or galactose-PEG using UDP-galactose or UDP-galactose-6-PEG, respectively, and a galactosyltransferase such as GalT1. In one case, the galactose-PEG is the terminal residue. In the second case, the galactose is further modified with SA-PEG using a CMP-SA-PEG donor and a sialyltransferase such as ST3GalIII. In another embodiment, the GlcNAc-Asn structures on the peptide/protein backbone may be
20 galactosylated and sialylated as described above, and then further sialylated using CMP-SA-PEG and an α 2,8-sialyltransferase such as the enzyme encoded by the *Campylobacter jejuni* cst-II gene.

Transferrin

25 42. GlycoPEGylation of Transferrin

This example sets forth the preparation of asialotransferrin and its sialylation with PEG-CMP-sialic acid.

Preparation of Asialo-transferrin. Human-derived holo-Transferrin, (10 mg) was dissolved in 500 μ L of 50 mM NaOAc, 5 mM CaCl₂, pH 5.5. To this solution was added
30 500 mU Neuraminidase II (*Vibrio cholerae*) and the reaction mixture was shaken gently for 20.5 hours at 37 °C. The reaction mixture was added to the prewashed N-(p-

aminophenyl)oxamic acid-agarose conjugate (600 μ L) and the washed beads gently rotated for 24 hours at 4 °C. The mixture was centrifuged at 10,000 rpm and the supernatant was collected. The reaction mixture was adjusted to 5 mM EDTA by addition of 100 μ L of 30 mM EDTA to the washed beads, which were gently rotated for 20 hours at 4 °C. The suspension was centrifuged for 2 minutes at 10,000 rpm and the supernatant was collected. The beads were washed 5 times with 0.35 mL of 50 mM NaOAc, 5 mM CaCl₂, 5 mM EDTA, pH 5.5 and all supernatants were pooled. The enzyme solution was dialyzed twice at 4 °C into 15 mM Tris-HCl, 1 M NaCl, pH 7.4. 0.3 mL of the transferrin solution (3.3 mL total) was removed and dialyzed twice against water. The remainder was dialyzed twice more at 4 °C against phosphate buffered saline. The dialyzed solution was stored at -20 °C. Protein samples were analyzed by IEF Electrophoresis. Samples (9 μ L, 25 μ g) were diluted with 16 μ L Tris buffer and mixed with 25 μ L of the sample loading buffer and applied to Isoelectric Focusing Gels (pH 3-7). Gels were run and fixed using standard procedures. Gels were stained with Colloidal Blue Stain.

Sialyl-PEGylation of asialo-Transferrin. Desialylated transferrin (250 μ g) and CMP-sialic acid or CMP-SA-PEG (1 kDa or 10 kDa)(0.05 μ mol) were dissolved in 69 μ L 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2 in 1.5 mL plastic tubes. The tubes were vortexed briefly and 100 mU ST3Gal3 (90 μ L) were added (total volume 250 μ L). The tubes were vortexed again and mixed gently for 24 hours at 32 °C. The reactions were stopped by freezing at -80 °C. Novex Tris-Glycine 8-16% 1 mm gels were used for SDS PAGE analysis (Figure 190). Samples (25 μ L, 25 μ g) were mixed with 25 μ L of sample loading buffer and 0.4 μ L of β -mercaptoethanol and heated for 6 minutes at 85 °C. Gels were run using standard conditions and stained with Colloidal Blue Stain. IEF gels were also performed as described above Figure 191). Samples were also dialyzed against water analyzed by MALDI-TOF.

Results. MALDI was also performed. Native transferrin (78729); asialotransferrin (78197); resialylated transferrin (79626/80703); with SA-PEG 1k (79037 (1); 80961 (2); 82535 (3); 84778 (4)); with SA-PEG 5k (90003 (2); 96117 (3); 96117 (4)); with SA-PEG 10k (100336 (2); 111421 (3); 122510 (4)).

43. Transferrin-GDNF

This example sets forth the procedures for the glycoconjugation of proteins, and in particular, transferrin is glycoconjugated to GDNF. Transferrin-SA-Linker-Gal-UDP is prepared from transferrin. The galactose residue is removed from GDNF glycans, and
5 Transferrin-SA-Linker-Gal-UDP is conjugated to GDNF glycans using a galactosyltransferase.

Preparation of agalacto-GDNF. GDNF produced in NSO cells (NSO murine myeloma cells) is dissolved at 2.5 mg/mL in 50 mM Tris 50 mM Tris-HCl pH 7.4, 0.15 M NaCl, and is incubated with 300 mU/mL beta-galactosidase-agarose conjugate for 16 hours at
10 32°C. To monitor the reaction a small aliquot of the reaction is diluted with the appropriate buffer and a IEF gel performed according to Invitrogen procedures. The mixture is centrifuged at 10,000 rpm and the supernatant is collected. The supernatant is dialyzed at 4 °C against 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃ and then twice more against 50 mM Tris -HCl pH 7.4, 1 M NaCl, 0.05% NaN₃. The dialyzed solution is then concentrated
15 using a Centricon Plus 20 centrifugal filter and stored at -20 °C. The conditions for the IEF gel are run according to the procedures and reagents provided by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Transferrin-SA-Linker-Gal-UDP. Asialo-transferrin is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 0.05% NaN₃, pH 7.2. The solution is
20 incubated with CMP-sialic acid-linker-Gal-UDP (molar amount to add 1 molar equivalent of nucleotide sugar to transferrin) and 0.1 U/mL of ST3Gal3 at 32°C for 2 days. To monitor the incorporation of sialic acid, a small aliquot of the reaction has ¹⁴C-SA-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label
25 incorporation into the peptide is quantitated using an in-line radiation detector.

The solution is incubated with 5 mM CMP-sialic acid and 0.1 U/mL of ST3Gal3 (to cap any unreacted transferrin glycans) at 32°C for 2 days. The incorporation into the peptide is quantitated using an in-line UV detector. After 2 days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) and collecting
30 fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE

and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

Preparation of Transferrin-SA-Linker-Gal-GDNF. The transferrin-SA-Linker-Gal-UDP prepared as described above is dissolved at 2.5 mg/mL in 50 mM Tris-HCl, 0.15 M NaCl, 5 mM MnCl₂, 0.05% NaN₃, pH 7.2. The solution is incubated with 2.5 mg/mL agalacto-GDNF and 0.1 U/mL of galactosyltransferase at 32°C for 2 days. To monitor the incorporation of galactose, a small aliquot of the reaction has ¹⁴C-galactose-UDP ligand added; the label incorporated into the peptide is separated from the free label by gel filtration on a Toso Haas G3000SW analytical column using PBS buffer (pH 7.1). The radioactive label incorporation into the peptide is quantitated using an in-line radiation detector.

When the reaction is complete, the solution is incubated with 5 mM UDP-Gal and 0.1 U/mL of galactosyltransferase (to cap any unreacted transferrin glycans) at 32°C for 2 days followed by addition of 5 mM CMP-SA and 0.1 U/mL of ST3Gal3. After 2 additional days, the reaction mixture is purified using a Toso Haas G3000SW preparative column using PBS buffer (pH 7.1) collecting fractions based on UV absorption. The product of the reaction is analyzed using SDS-PAGE and IEF analysis according to the procedures and reagents supplied by Invitrogen. Samples are dialyzed against water and analyzed by MALDI-TOF MS.

The disclosures of each and every patent, patent application, and publication cited herein are hereby incorporated herein by reference in their entirety.

While this invention has been disclosed with reference to specific embodiments, it is apparent that other embodiments and variations of this invention may be devised by others skilled in the art without departing from the true spirit and scope of the invention. The appended claims are intended to be construed to include all such embodiments and equivalent variations.

What is claimed:

1. An EPO peptide comprising one or more glycans, having a glycoconjugate molecule covalently attached to said peptide.

2. The EPO peptide of claim 1, wherein said one or more glycans is a
5 monoantennary glycan.

3. The EPO peptide of claim 1, wherein said one or more glycans is a biantennary glycan.

4. The EPO peptide of claim 1, wherein said one or more glycans is a triantennary glycan.

10 5. The EPO peptide of claim 1, wherein said one or more glycans is at least a triantennary glycan.

6. The EPO peptide of claim 1, wherein said one or more glycans comprises at least two glycans comprising a mixture of mono or multiantennary glycans.

15 7. The EPO peptide of claim 1, wherein said one or more glycans is selected from an N-linked glycan and an O-linked glycan.

8. The EPO peptide of claim 1, wherein said one or more glycans is at least two glycans selected from an N-linked and an O-linked glycan.

9. The EPO peptide of claim 1, wherein said peptide is expressed in a cell selected from the group consisting of a prokaryotic cell and a eukaryotic cell.

20 10. The EPO peptide of claim 9, wherein said eukaryotic cell is selected from the group consisting of a mammalian cell, an insect cell and a fungal cell.

11. The EPO peptide of claim 10, wherein said fungal cell is a yeast cell.

12. A glycoPEGylated EPO peptide comprising an EPO peptide and at least one glycan and at least one poly(ethylene glycol) molecule covalently attached to said glycan,

wherein said poly(ethylene glycol) molecule is added to said EPO peptide using a glycosyltransferase.

13. The glycoPEGylated EPO peptide of claim 12, comprising at least one
5 mono-antennary glycan.

14. The glycoPEGylated EPO peptide of claim 12, wherein all of said glycans are N-linked and are mono-antennary.

10 15. The glycoPEGylated EPO peptide of claim 12, wherein all of said glycans are N-linked and at least one of said glycans comprise said poly(ethylene glycol).

16. The glycoPEGylated EPO peptide of claim 15, wherein more than one of said glycans comprises said poly(ethylene glycol).
15

17. The glycoPEGylated EPO peptide of claim 12, wherein all of said glycans are N-linked and all of said glycans comprise said poly(ethylene glycol).

18. The glycoPEGylated EPO peptide of claim 12, comprising at least three
20 mono-antennary glycans having said poly(ethylene glycol) covalently attached thereto.

19. A glycoPEGylated EPO peptide, wherein said EPO peptide comprises three or more glycans.

25 20. The glycoPEGylated EPO peptide of claim 9, wherein at least one of said glycans comprises said poly(ethylene glycol) covalently attached thereto.

21. The glycoPEGylated EPO peptide of claim 18, wherein more than one of said glycans comprises said poly(ethylene glycol) covalently attached thereto.
30

22. The glycoPEGylated EPO peptide of claim 18, wherein all of said glycans comprise said poly(ethylene glycol) covalently attached thereto.

5 23. The glycoPEGylated EPO peptide of claim 12 wherein said poly(ethylene glycol) is linked to at least one sugar moiety selected from the group consisting of fucose (Fuc), N-acetylglucosamine (GlcNAc), galactose (Gal) and a sialic acid (SA).

24. The glycoPEGylated EPO peptide of claim 23, wherein said sialic acid is N-acetylneuraminic acid.

10 25. The glycoPEGylated EPO peptide of claim 12, wherein said EPO peptide does not comprise an O-linked glycan.

26. The glycoPEGylated EPO peptide of claim 12 wherein said EPO peptide comprises at least one O-linked glycan.

15

27. The glycoPEGylated EPO peptide of claim 26, wherein said O-linked peptide comprises said poly(ethylene glycol) covalently attached thereto.

20 28. The glycoPEGylated EPO peptide of claim 27, wherein said EPO peptide is recombinantly expressed in a cell.

29. The glycoPEGylated EPO peptide of claim 28, wherein said cell is selected from the group consisting of an insect cell, a fungal cell and a mammalian cell.

25 30. The glycoPEGylated EPO peptide of claim 29, wherein said fungal cell is a yeast cell.

31. The glycoPEGylated EPO peptide of claim 29, wherein said cell is an insect cell.

30

32. The glycoPEGylated EPO peptide of claim 29, wherein said cell is a yeast cell.

33. The glycoPEGylated EPO peptide of claim 29, wherein said cell is a
5 mammalian cell.

34. The glycoPEGylated EPO peptide of claim 33, wherein said mammalian cell is a CHO cell.

10 35. The glycoPEGylated EPO peptide of claim 12, wherein said poly(ethylene glycol) has a molecular weight selected from the group consisting of about 1 kDa, 2 kDa, 5 kDa, 10 kDa, 20 kDa, 30 kDa and 40 kDa.

15 36. The glycoPEGylated EPO peptide of claim 35, wherein said poly(ethylene glycol) has a molecular weight of 20 kDa.

37. The glycoPEGylated EPO peptide of claim 12, wherein said EPO peptide is selected from the group consisting of a naturally occurring EPO peptide and a mutated EPO peptide.

20 38. The glycoPEGylated EPO peptide of claim 37, wherein said mutated EPO peptide comprises the amino acid sequence of SEQ ID NO:73 having at least one mutation selected from the group consisting of Arg¹³⁹ to Ala¹³⁹, Arg¹⁴³ to Ala¹⁴³ and Lys¹⁵⁴ to Ala¹⁵⁴.

25 39. A method of making a glycoPEGylated EPO peptide, said method comprising the step of:

(a) contacting an EPO peptide with a mixture comprising a nucleotide sugar covalently linked to poly(ethylene glycol) and a glycosyltransferase under conditions sufficient to transfer said poly(ethylene glycol) to said EPO peptide.

30

40. The method of claim 39, wherein the sugar of said nucleotide sugar is selected from the group consisting of fucose (Fuc), N-acetylglucosamine (GlcNAc), galactose (Gal) and a sialic acid (SA).

5 41. The method of claim 40, wherein said sialic acid is N-acetylneuraminic acid (NAN).

 42. The method of claim 39, wherein said poly(ethylene glycol) has a molecular weight selected from the group consisting of about 1 kDa, 2 kDa, 5 kDa, 10 kDa,
10 20 kDa, 30 kDa and 40 kDa.

 43. The method of claim 42, wherein said poly(ethylene glycol) has a molecular weight of 20 kDa.

15 44. The method of claim 39, wherein said EPO peptide is recombinantly expressed in a cell.

 45. The method of claim 44, wherein said cell is selected from the group consisting of an insect cell, a fungal cell and a mammalian cell.

20

 46. The method of claim 45, wherein said cell is an insect cell.

 47. The method of claim 45, wherein said cell is a yeast cell.

25 48. The method of claim 45, wherein said cell is a mammalian cell.

 49. The method of claim 48, wherein said mammalian cell is a CHO cell.

 50. The method of claim 39, wherein said EPO peptide is selected from the
30 group consisting of a naturally occurring EPO peptide and a mutated EPO peptide.

51. The method of claim 50, wherein said mature EPO peptide has the sequence of SEQ ID NO:73.

52. The method of claim 50, wherein said mutated EPO peptide comprises the amino acid sequence of SEQ ID NO: 73 having at least one mutation selected from the group consisting of Arg¹³⁹ to Ala¹³⁹, Arg¹⁴³ to Ala¹⁴³ and Lys¹⁵⁴ to Ala¹⁵⁴.

53. The method of claim 39, wherein before step (a):

(b) contacting said EPO peptide with a mixture comprising a nucleotide-N-acetylglucosamine (GlcNAc) molecule and an N-acetylglucosamine transferase (GnT) for which the nucleotide-GlcNAc is a substrate under conditions sufficient to form a bond between said GlcNAc and said EPO, wherein said GnT is selected from the group consisting of GnT I, GnT II, GnT III, GnT IV, GnT V and GnT VI.

54. The method of claim 53, wherein said mixture comprises one GnT selected from the group consisting of GnT I, GnT II, GnT IV, GnT V and GnT VI.

55. The method of claim 54, wherein said GnT is GnT I.

56. The method of claim 54, wherein said GnT is GnT II.

57. The method of claim 39, wherein said glycoPEGylated EPO peptide comprises at least one mono-antennary glycan.

58. The method of claim 39, wherein the sugar of said nucleotide sugar is galactose and said glycosyltransferase is galactosyl transferase I (GalT I).

59. The method of claim 53, wherein before step (a) but after step (b):
(c) contacting said EPO peptide with a mixture comprising a nucleotide galactose (Gal) and galactosyl transferase I (GalT I) under conditions sufficient to transfer galactose to said EPO peptide.

60. The method of claim 39, wherein in step (a), the sugar of said nucleotide sugar is sialic acid and said glycosyltransferase is a sialyltransferase.

5 61. The method of claim 60, wherein said sialic acid is N-acetylneuraminic acid (NAN).

62. The method of claim 60, wherein said sialyltransferase is selected from the group consisting of $\alpha(2,3)$ sialyltransferase, $\alpha(2,6)$ sialyltransferase and
10 (2,8)sialyltransferase.

63. A glycoPEGylated EPO peptide made by the method of claim 39.

64. A glycoPEGylated EPO peptide, said EPO peptide comprising the
15 sequence of SEQ ID NO:73.

65. A glycoPEGylated EPO peptide, said EPO peptide comprising the sequence of SEQ ID NO:73 and further comprising a mutation in said sequence.

20 66. A method of making a glycoPEGylated EPO peptide, said method comprising the steps of:

 (a) contacting an EPO peptide with a mixture comprising a nucleotide sugar covalently linked to poly(ethylene glycol) and a glycosyltransferase under conditions sufficient to transfer said poly(ethylene glycol) to said EPO peptide, wherein said
25 glycosyltransferase is a fucosyltransferase.

67. The method of claim 66, wherein said fucosyltransferase is selected from the group consisting of fucosyltransferase I, fucosyltransferase III, fucosyltransferase IV, fucosyltransferase V, fucosyltransferase VI and fucosyltransferase VII.

30 68. A glycoPEGylated EPO peptide made by the method of claim 66.

69. The method of claim 66, wherein said EPO peptide is expressed in a CHO cell.

5 70. A method of treating a mammal having anemia, said method comprising administering to said mammal an EPO peptide having one or more glycans having a glycoconjugate molecule attached to said peptide, wherein said EPO peptide is administered in an amount effective to increase the hematocrit level in said mammal.

10 71. The method of claim 70, wherein said mammal is a human.

 72. A method of providing erythropoietin therapy to a mammal, said method comprising administering an effective amount of a glycoPEGylated EPO peptide comprising an EPO peptide and at least one glycan and at least one poly(ethylene glycol) molecule covalently attached to said glycan, wherein said poly(ethylene glycol) molecule is added to
15 said EPO peptide using a glycosyltransferase, wherein said EPO peptide is administered in an amount effective to increase the hematocrit level in said mammal.

 73. The method of claim 72, wherein said mammal is a human.
20

 74. A method of treating a mammal having anemia, said method comprising administering to said mammal a glycoPEGylated EPO peptide comprising an EPO peptide and at least one glycan and at least one poly(ethylene glycol) molecule covalently attached to said glycan, wherein said poly(ethylene glycol) molecule is added to said EPO peptide using
25 a glycosyltransferase, wherein said EPO peptide is administered in an amount effective to increase the hematocrit level in said mammal..

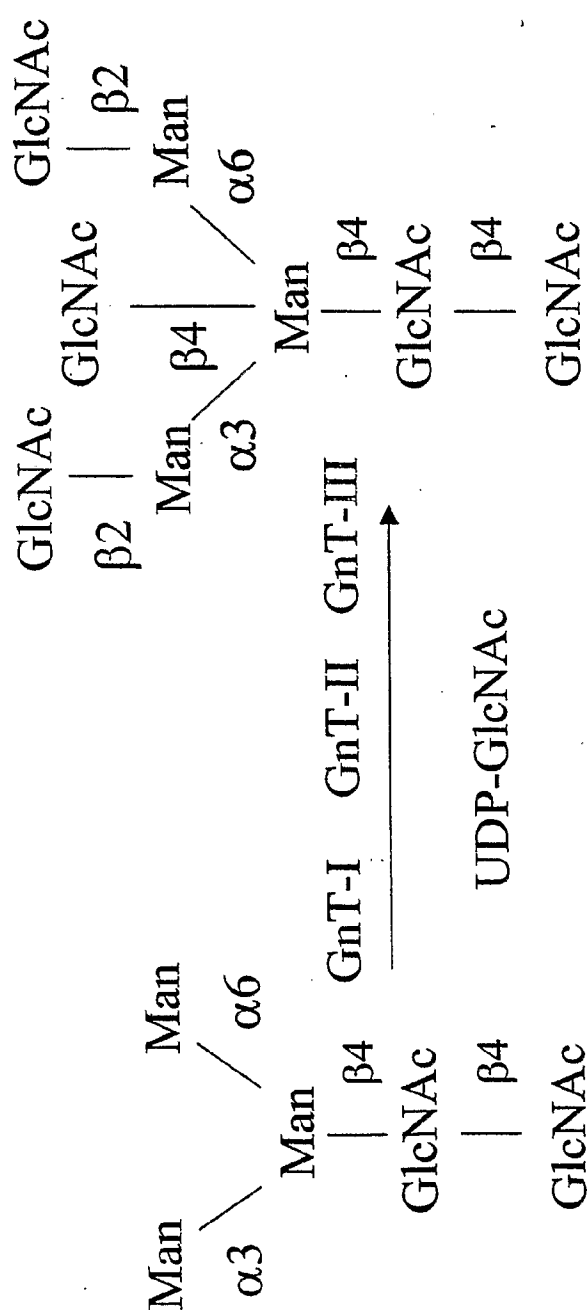
 75. The method of claim 74, wherein said mammal is a human.
30

76. The method of claim 75, wherein said anemia is associated with chemotherapy.

77. A method of treating a kidney dialysis patient, said method comprising
5 administering to said patient a glycoPEGylated EPO peptide comprising an EPO peptide and
at least one glycan and at least one poly(ethylene glycol) molecule covalently attached to said
glycan, wherein said poly(ethylene glycol) molecule is added to said EPO peptide using a
glycosyltransferase, wherein said EPO peptide is administered in an amount effective to
increase the hematocrit level in said patient.

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Trimannosyl core Trimannosyl core with
Bisecting GlcNAc

FIG. 1

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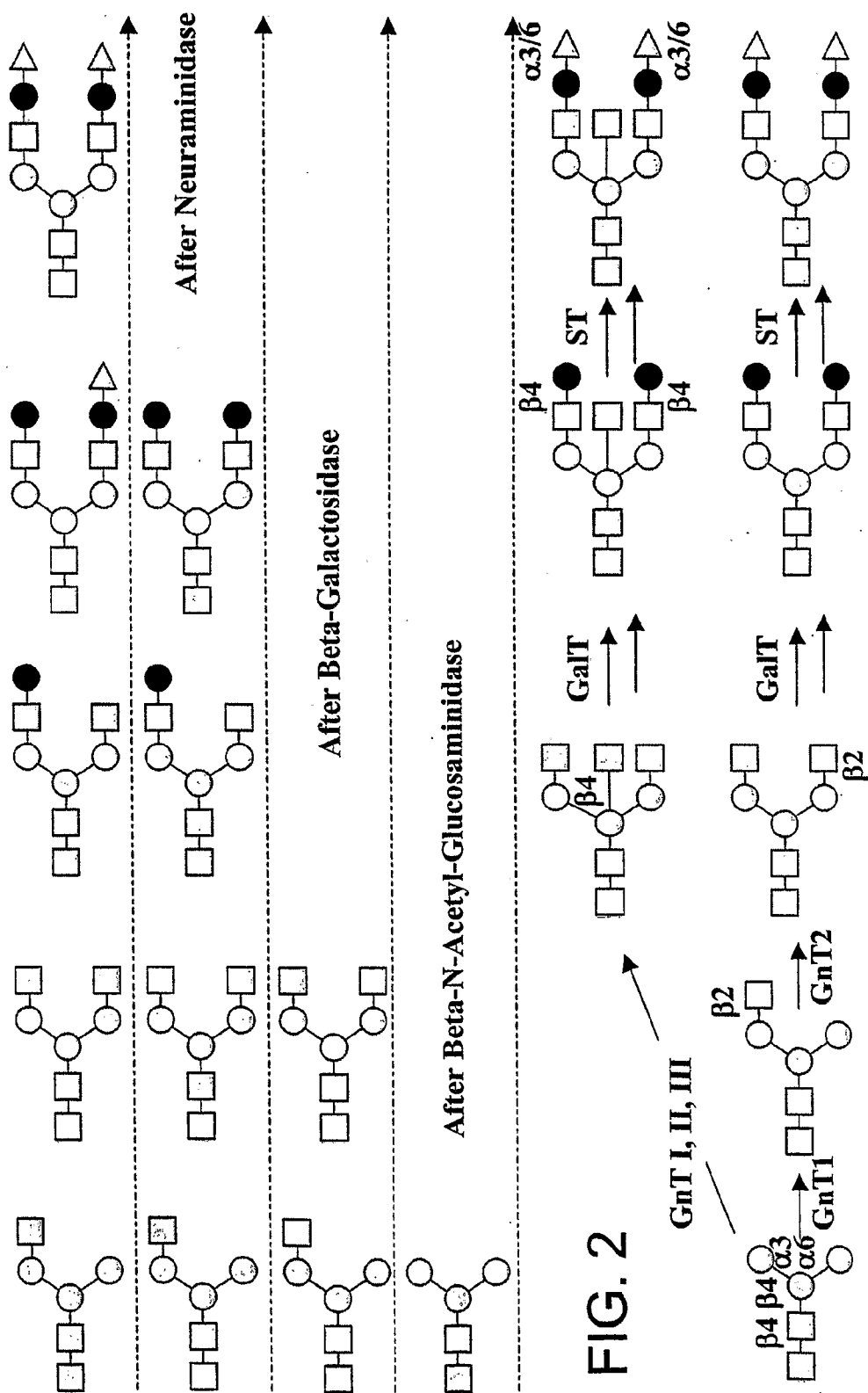


FIG. 2

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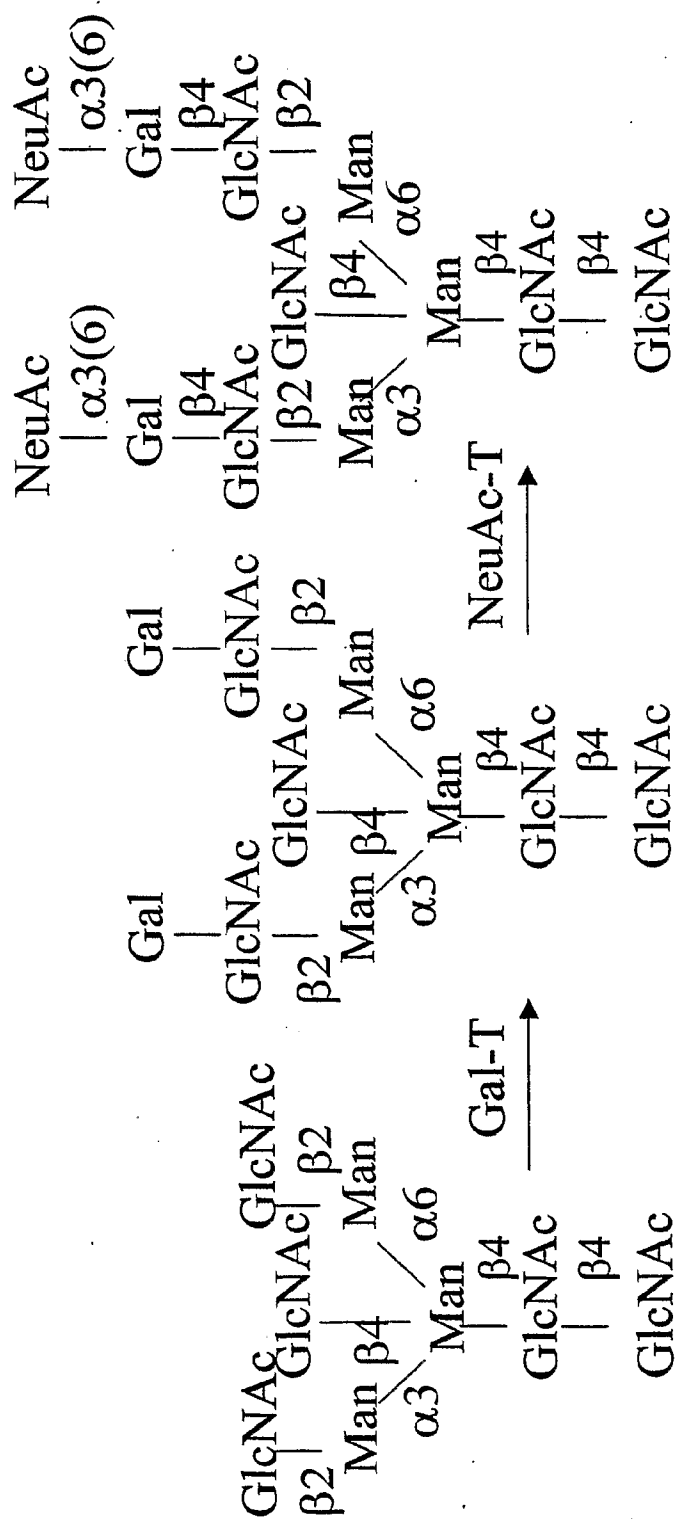


FIG. 3

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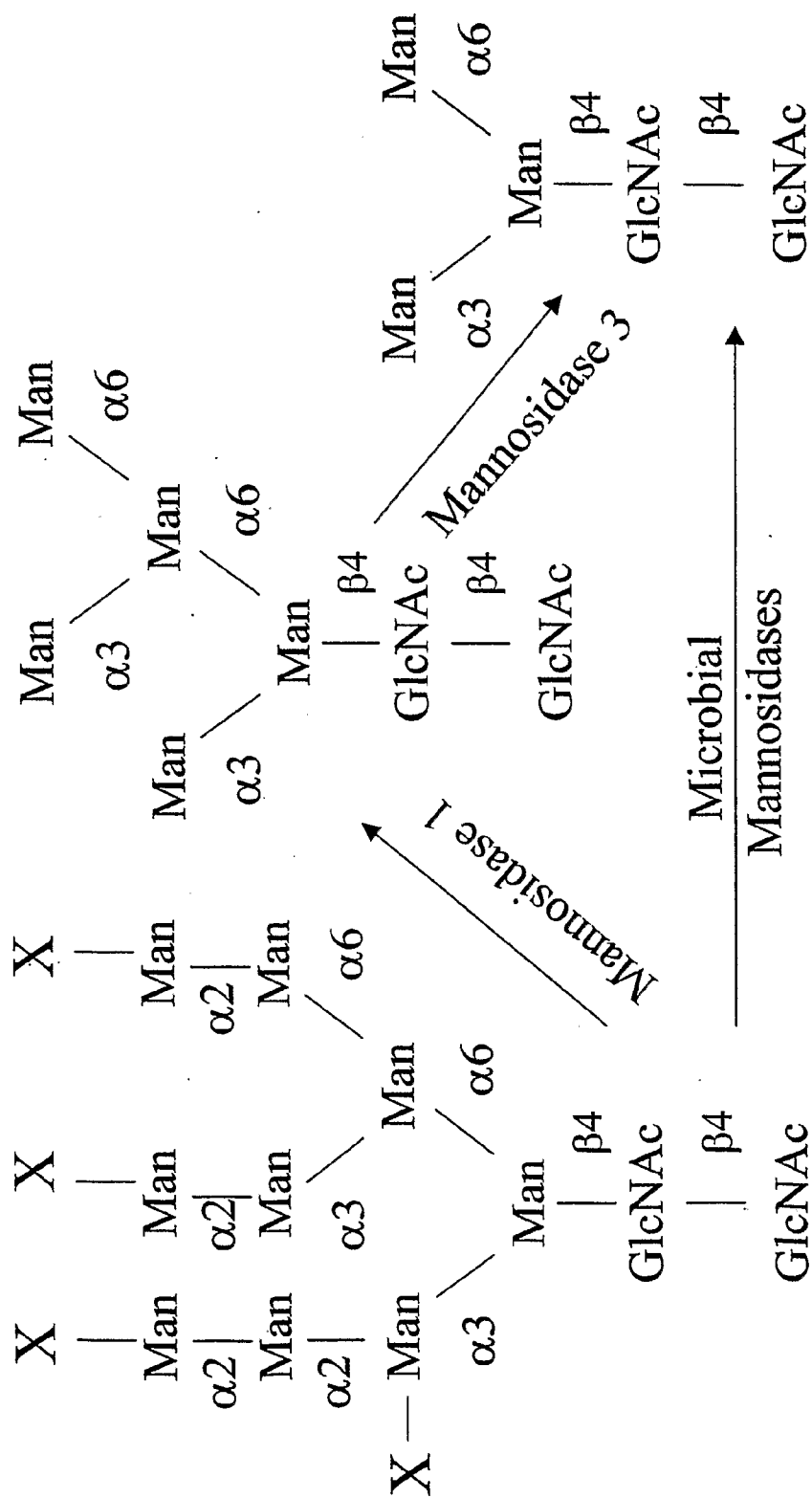


FIG. 4

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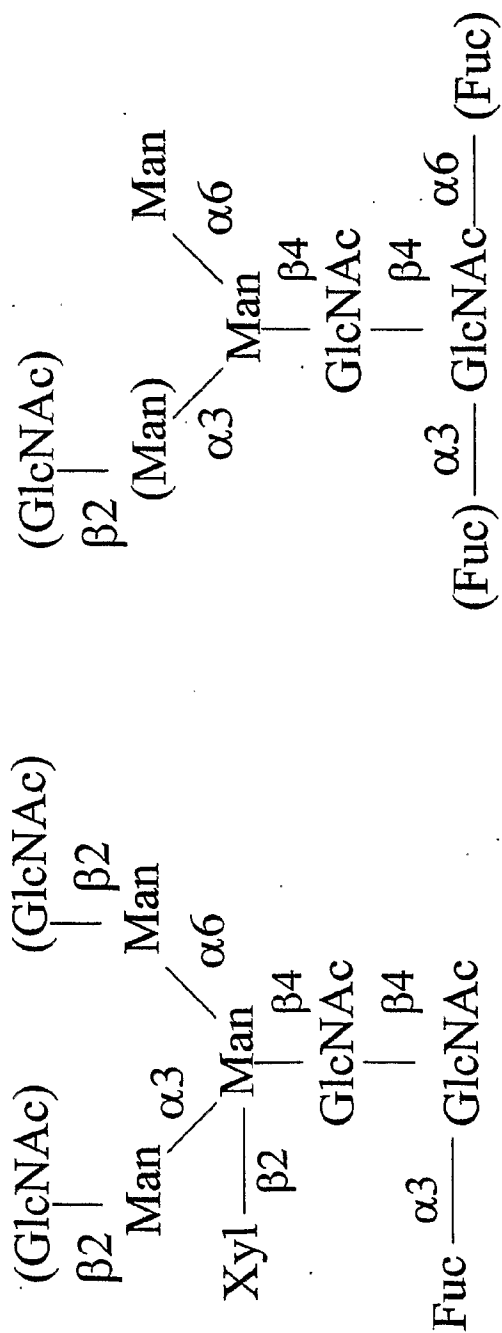


FIG. 5

FIG. 6

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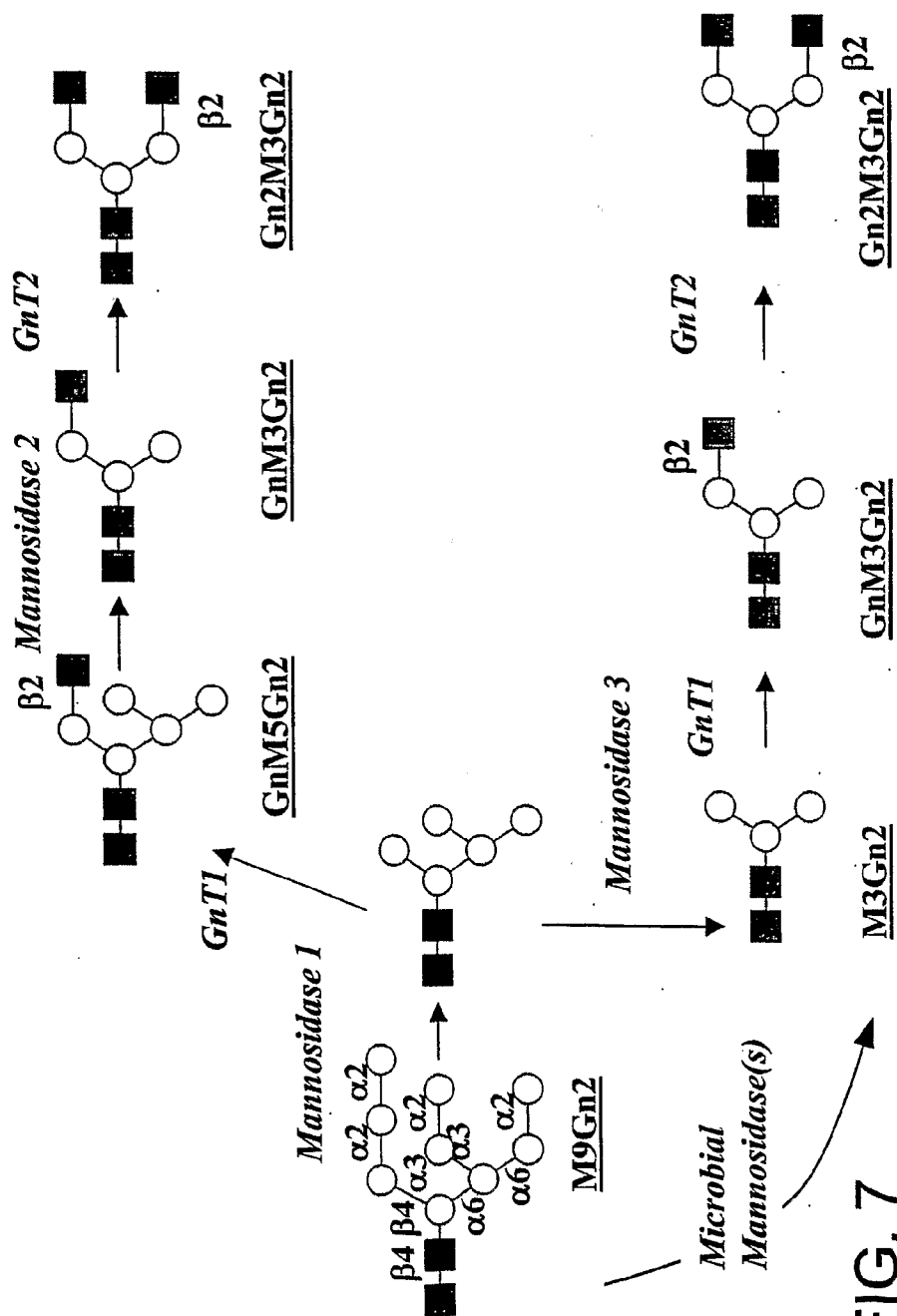


FIG. 7

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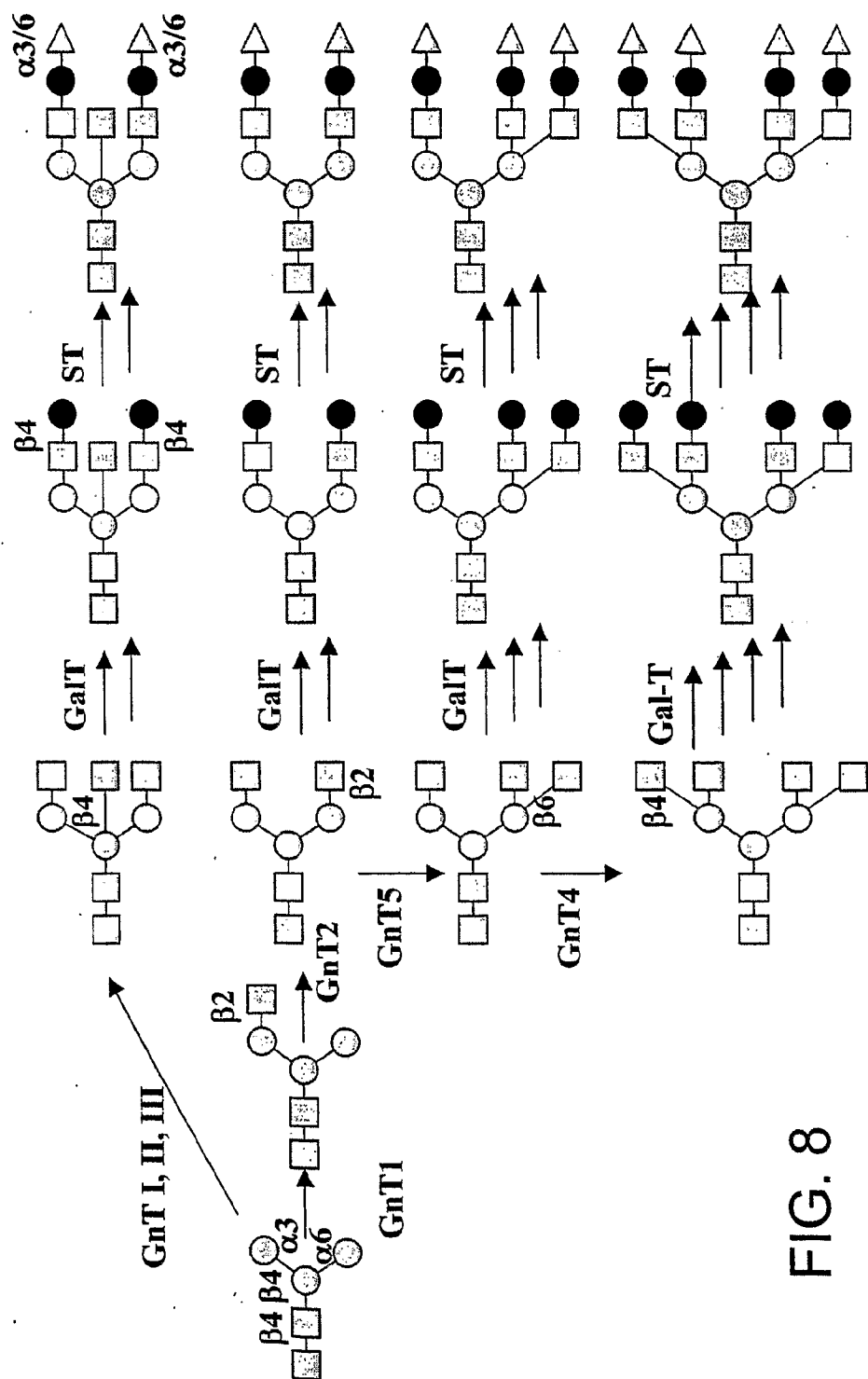


FIG. 8

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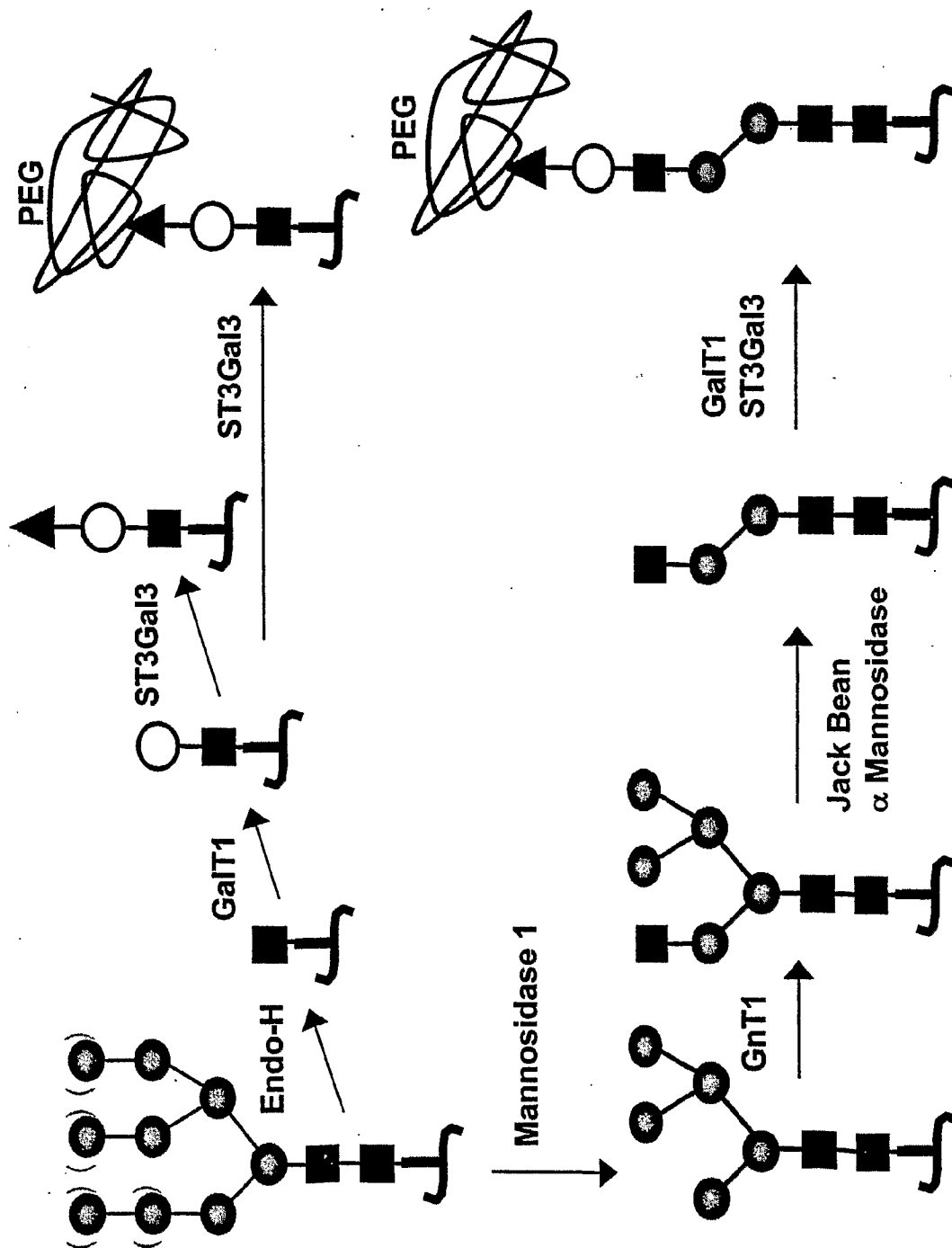


FIG. 9

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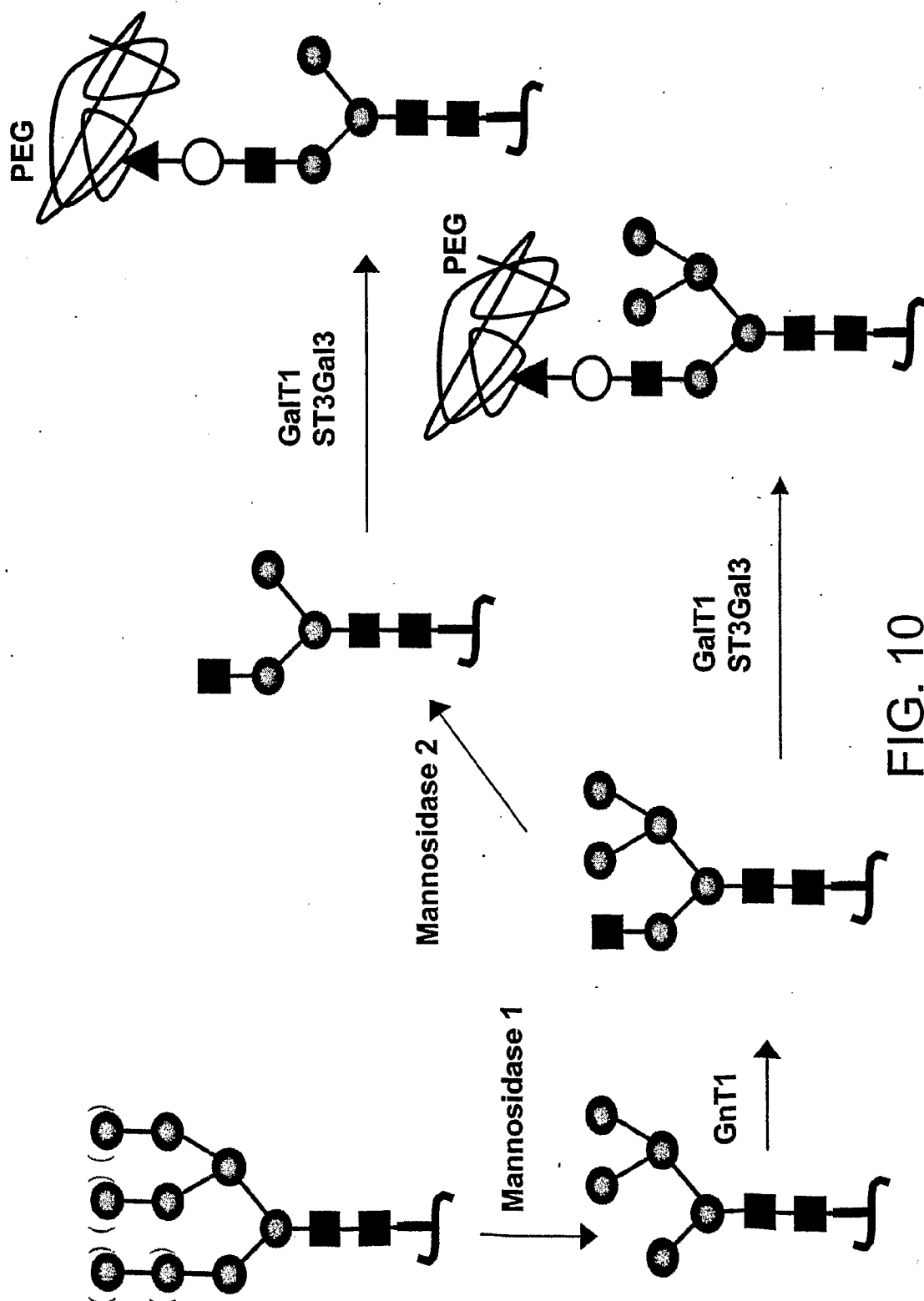


FIG. 10

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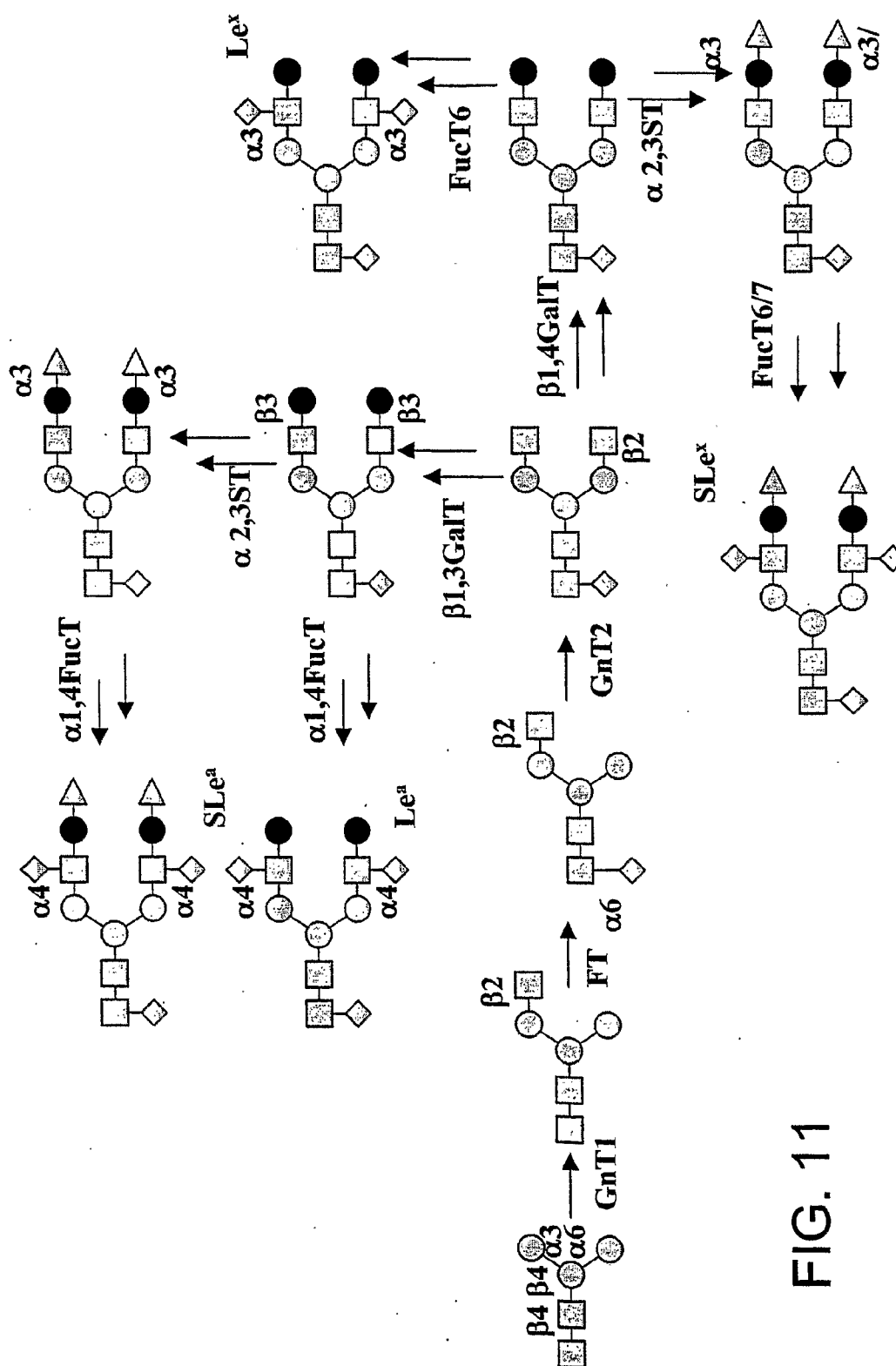


FIG. 11

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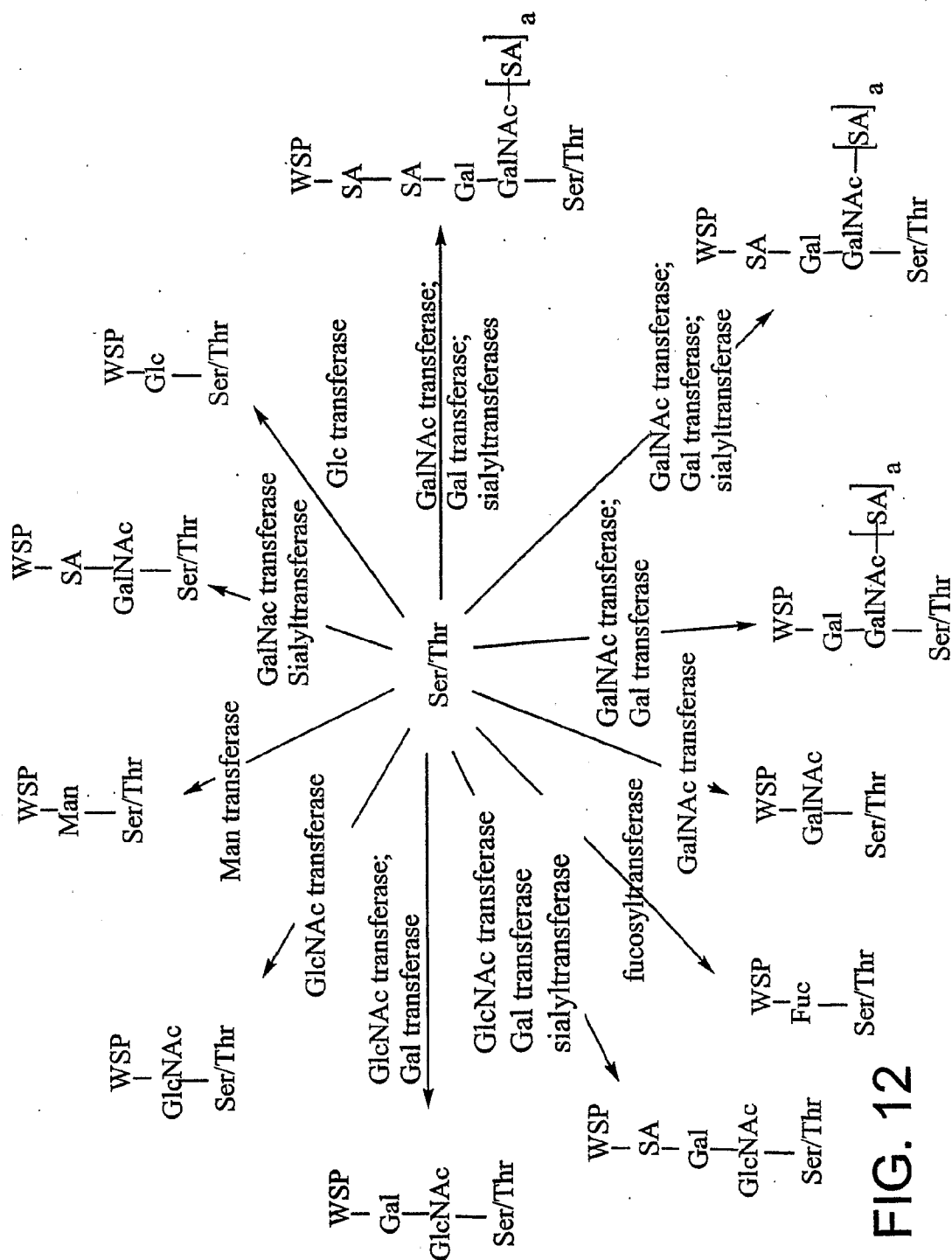


FIG. 12

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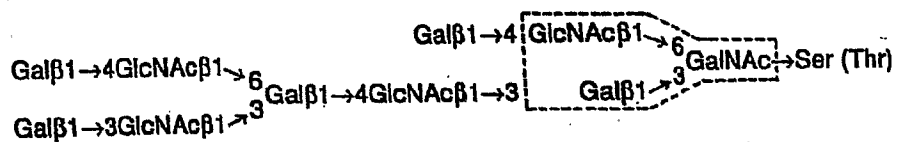
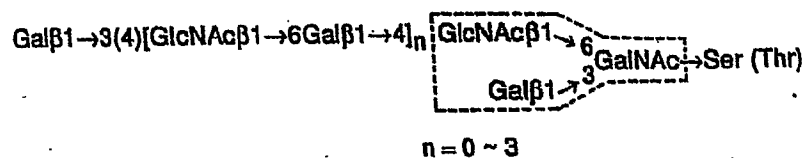
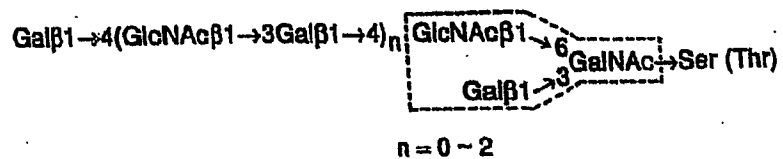
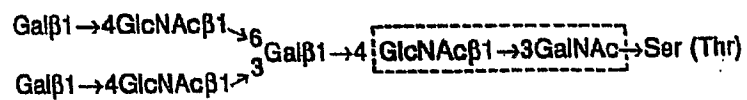
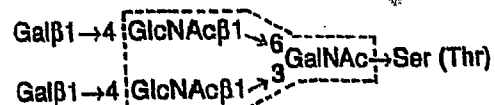
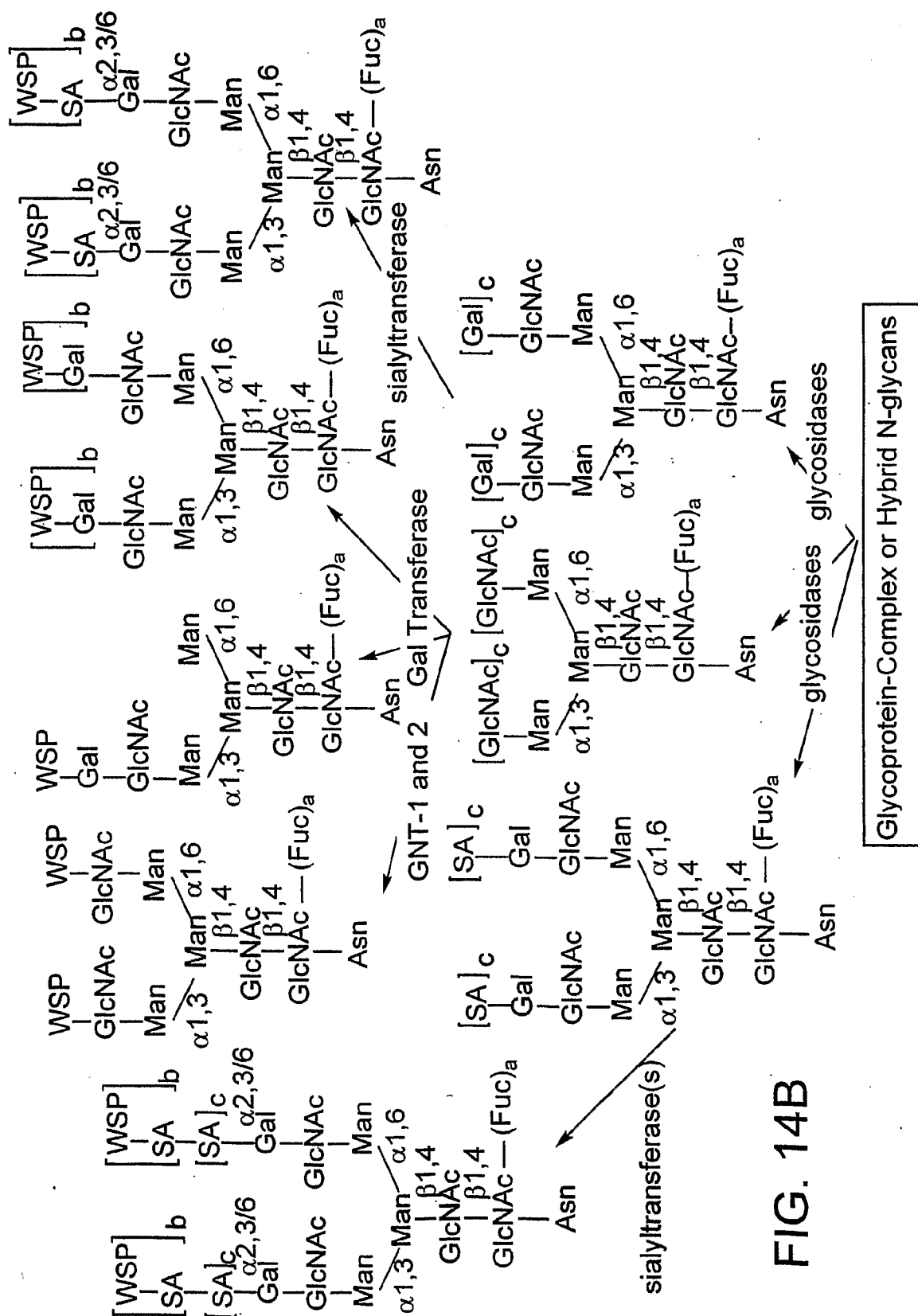
Core 1**Core 2****Core 3****Core 4**

FIG. 13

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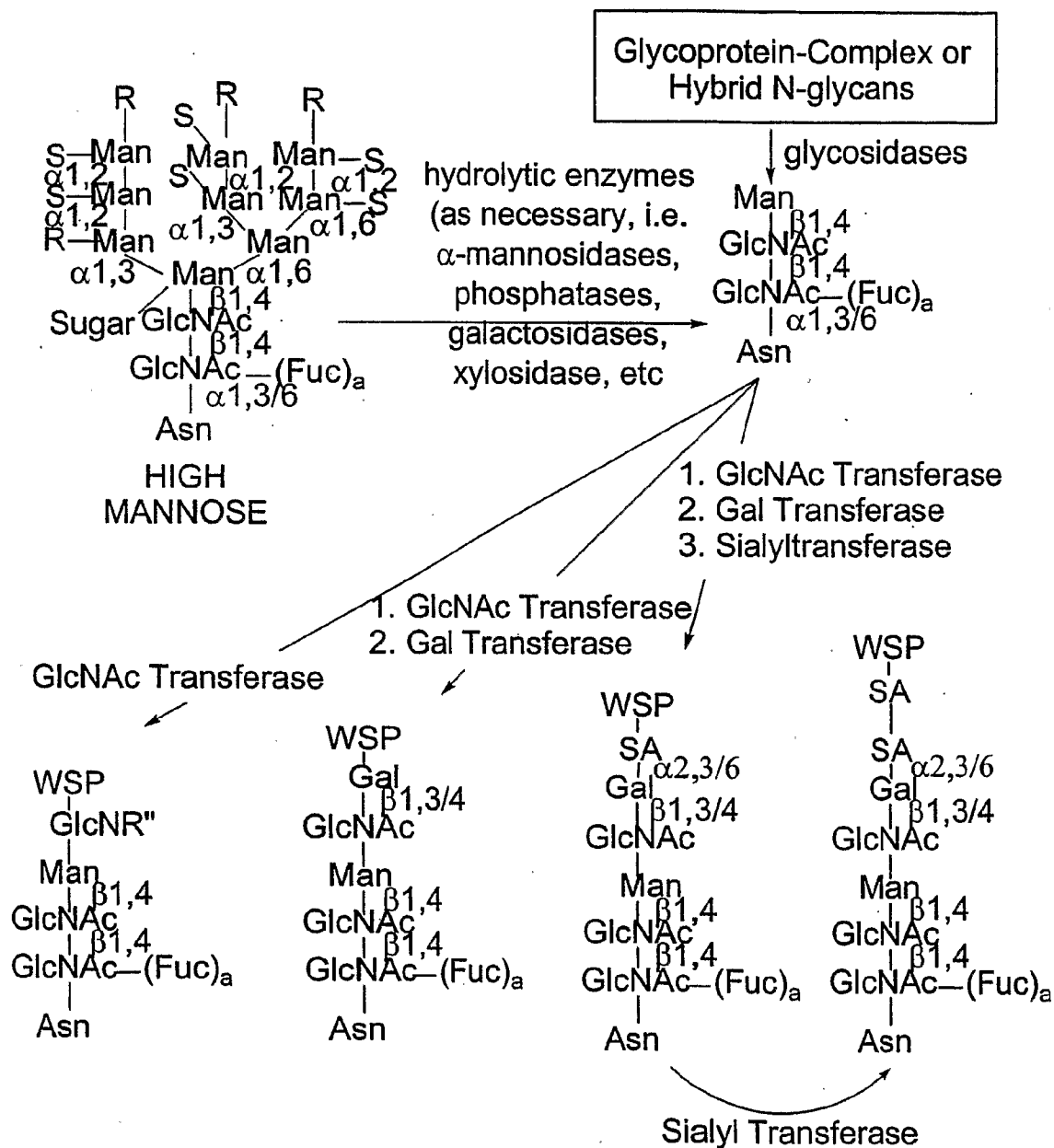


FIG. 15

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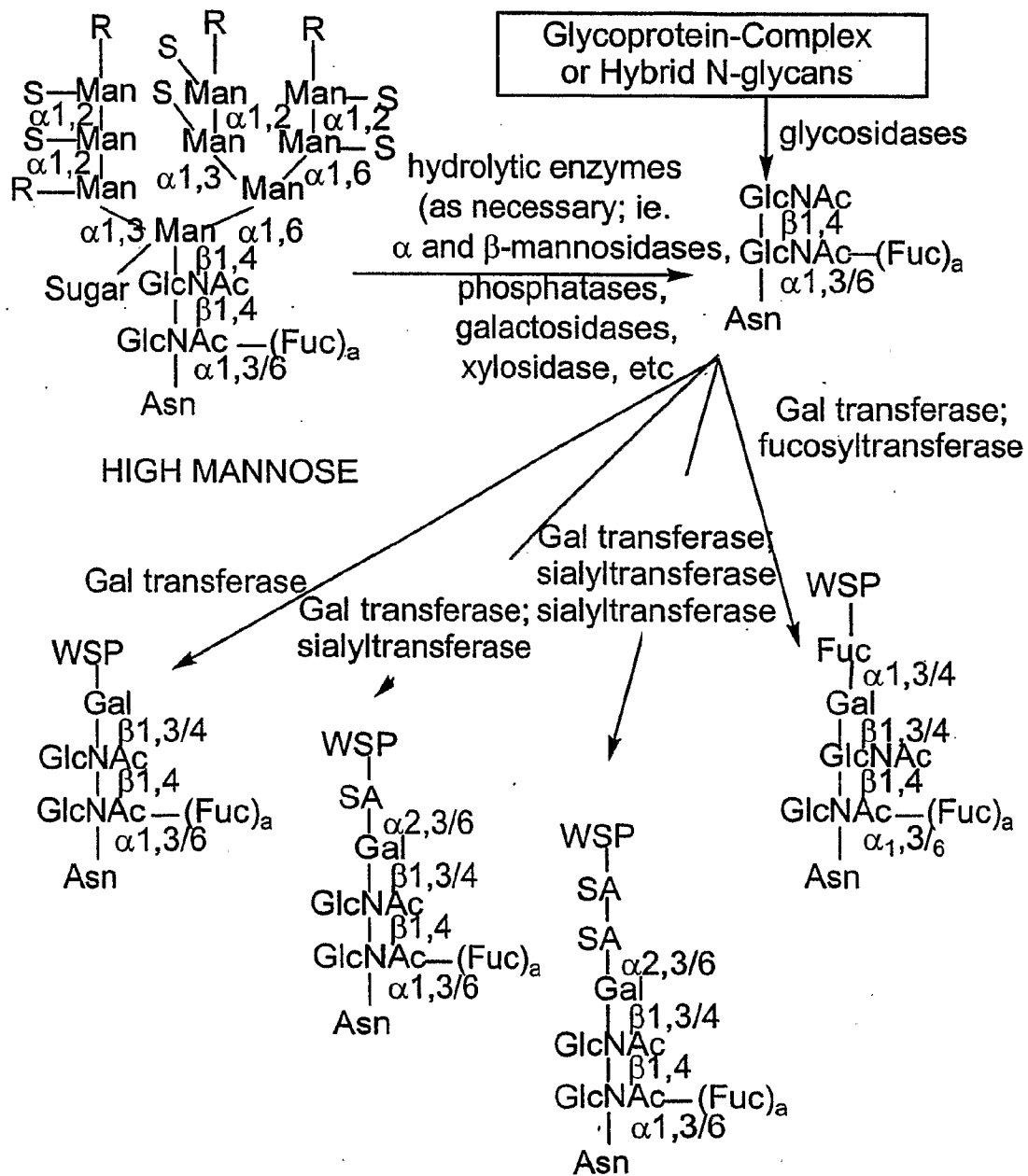


FIG. 16

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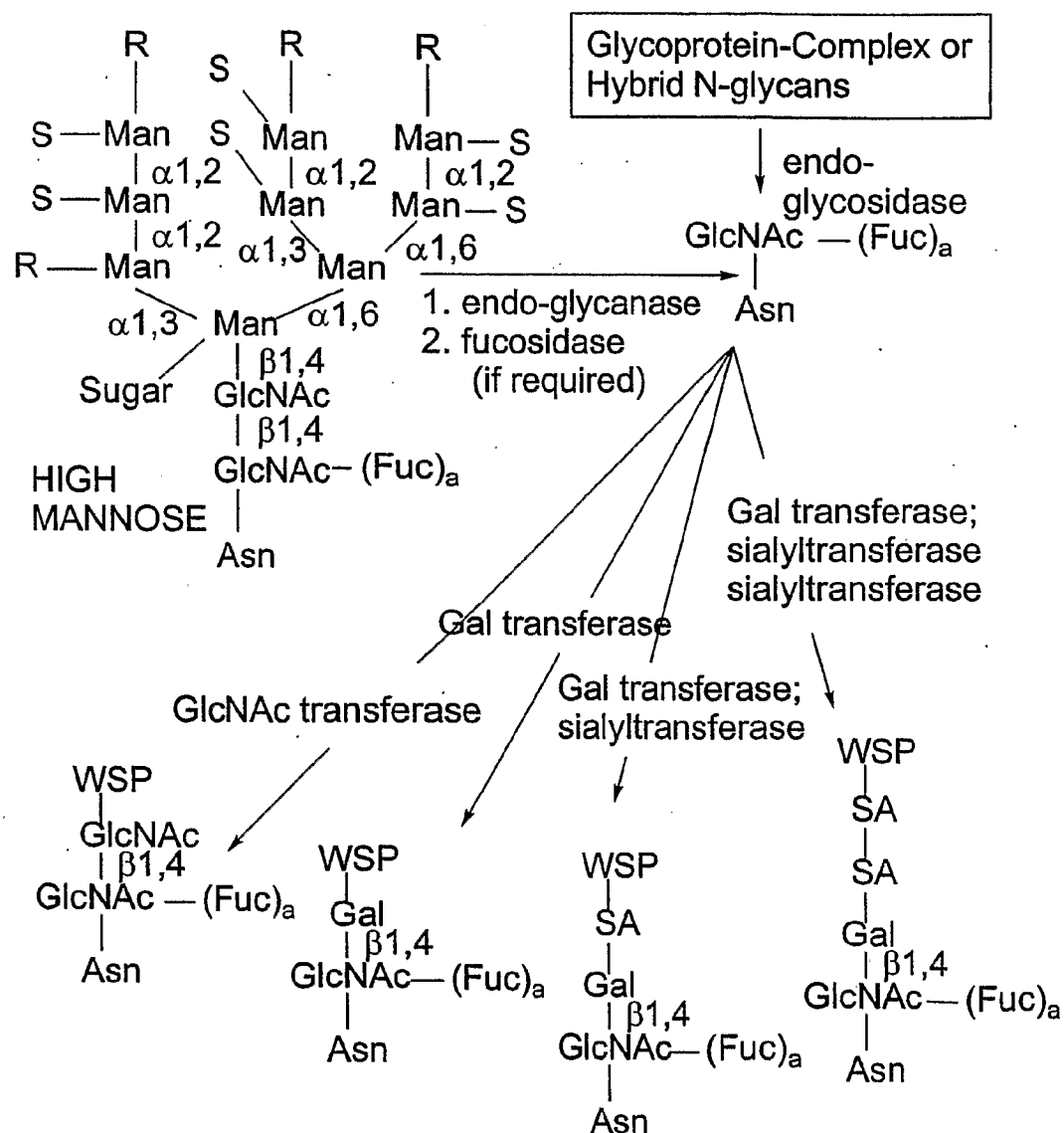


FIG. 17

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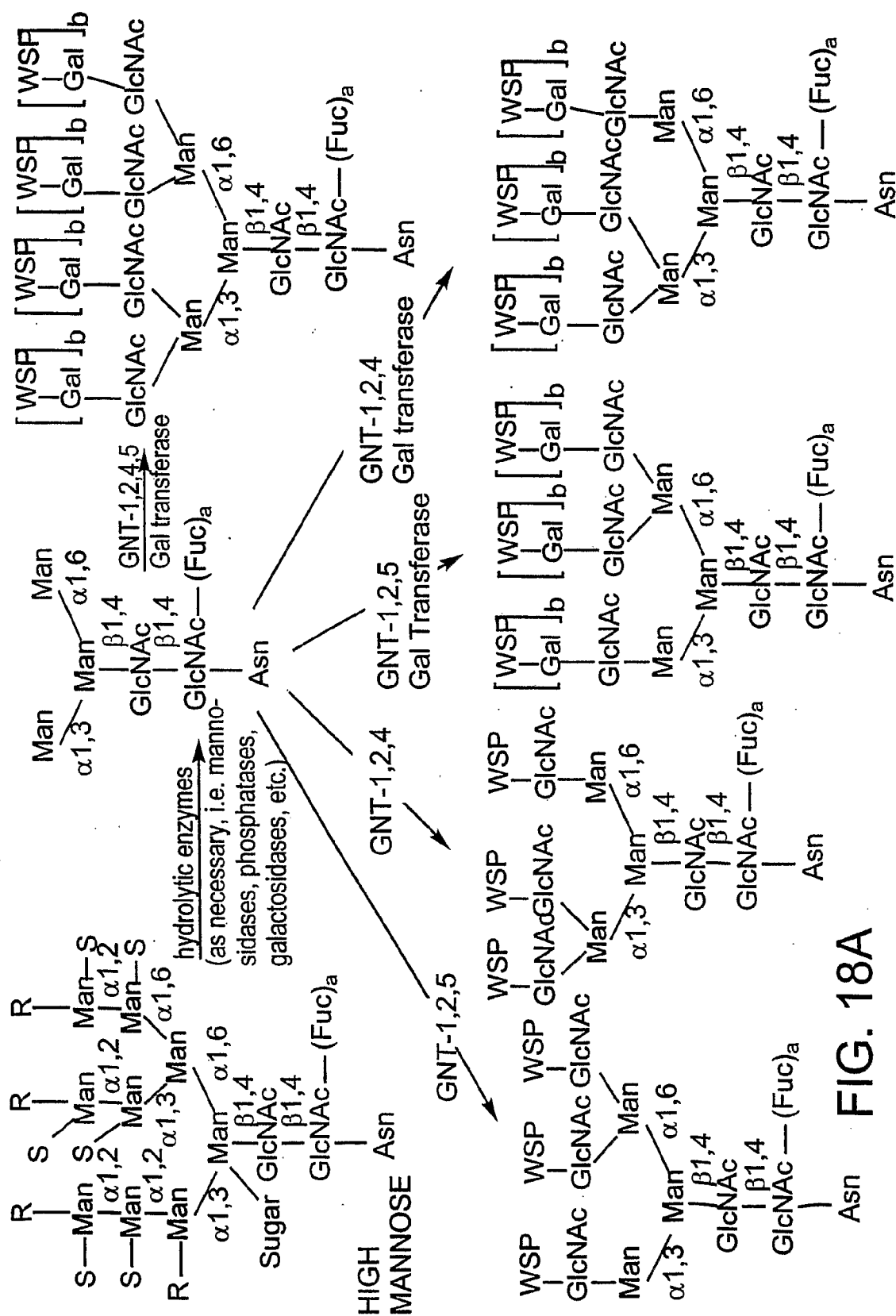


FIG. 18A

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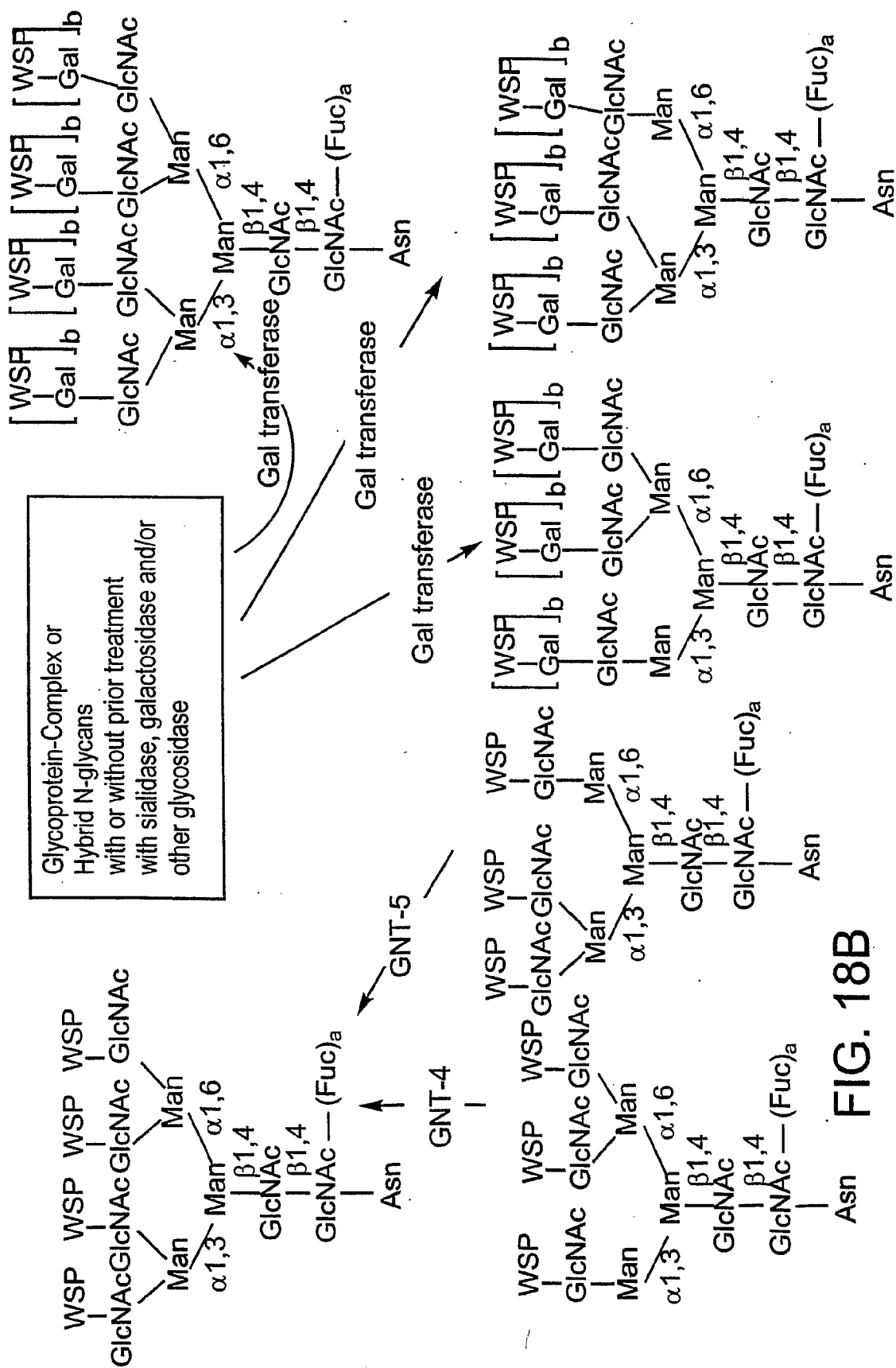
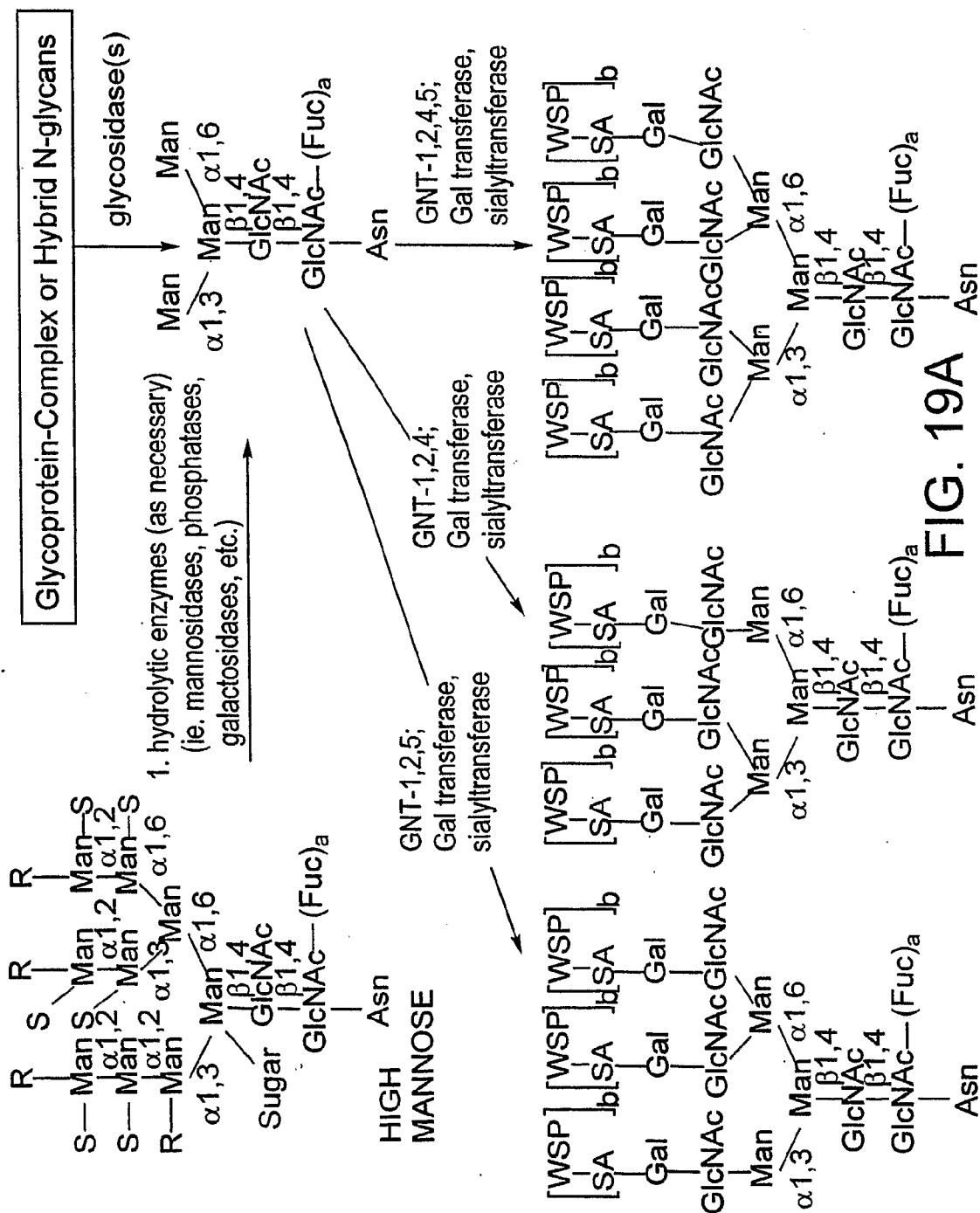
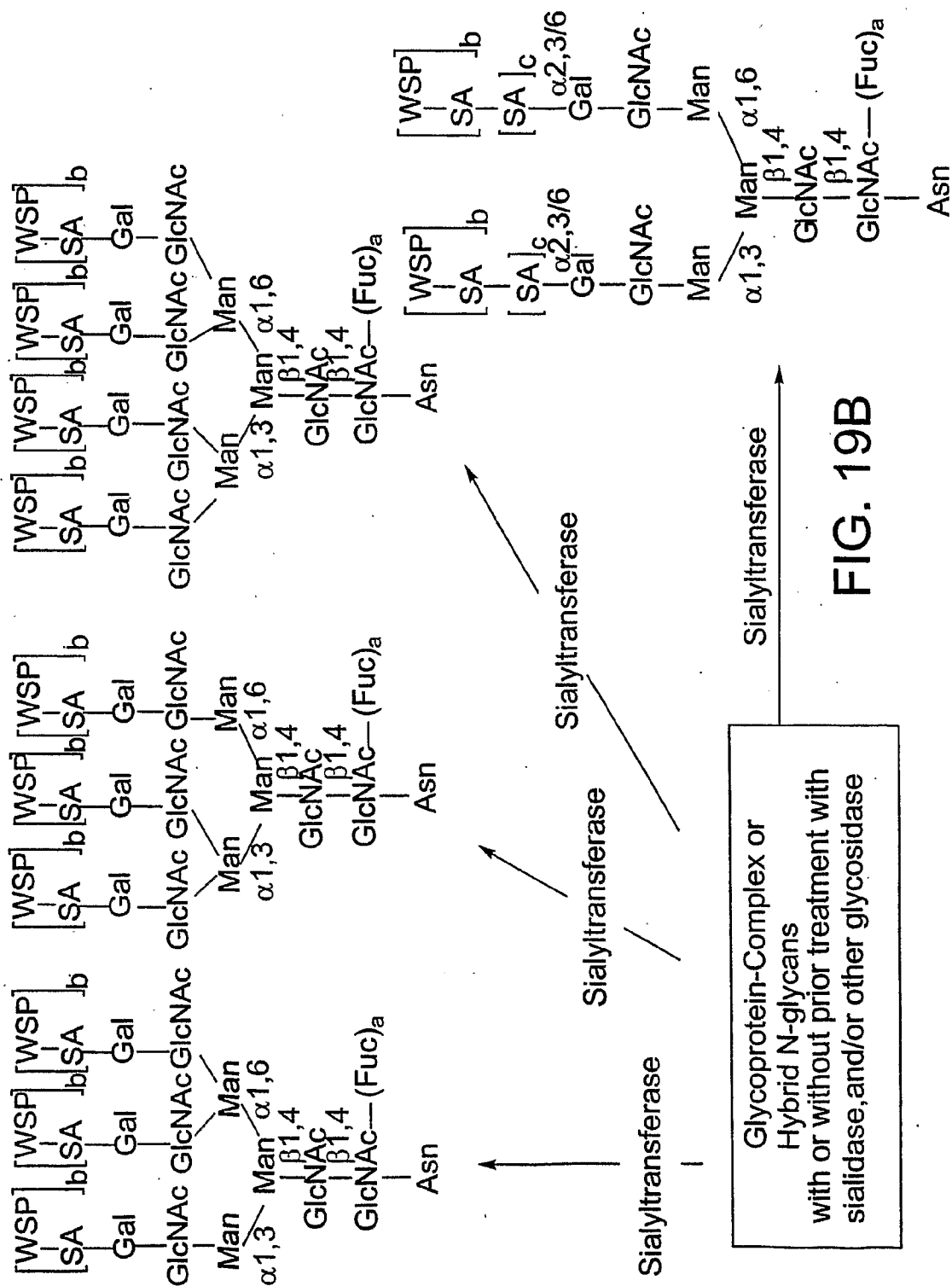


FIG. 18B

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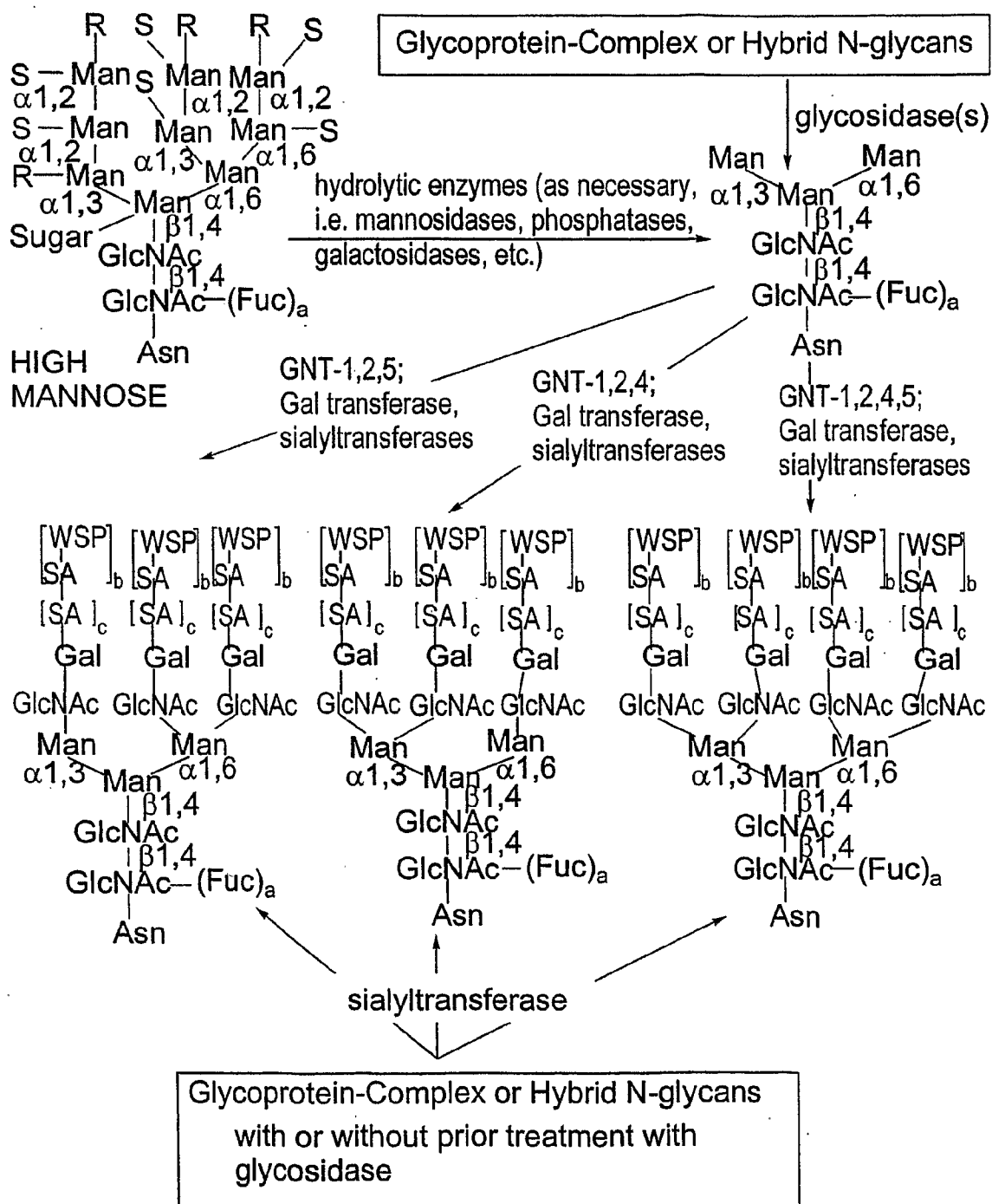


FIG. 20

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O-LINKED OLIGOSACCHARIDES

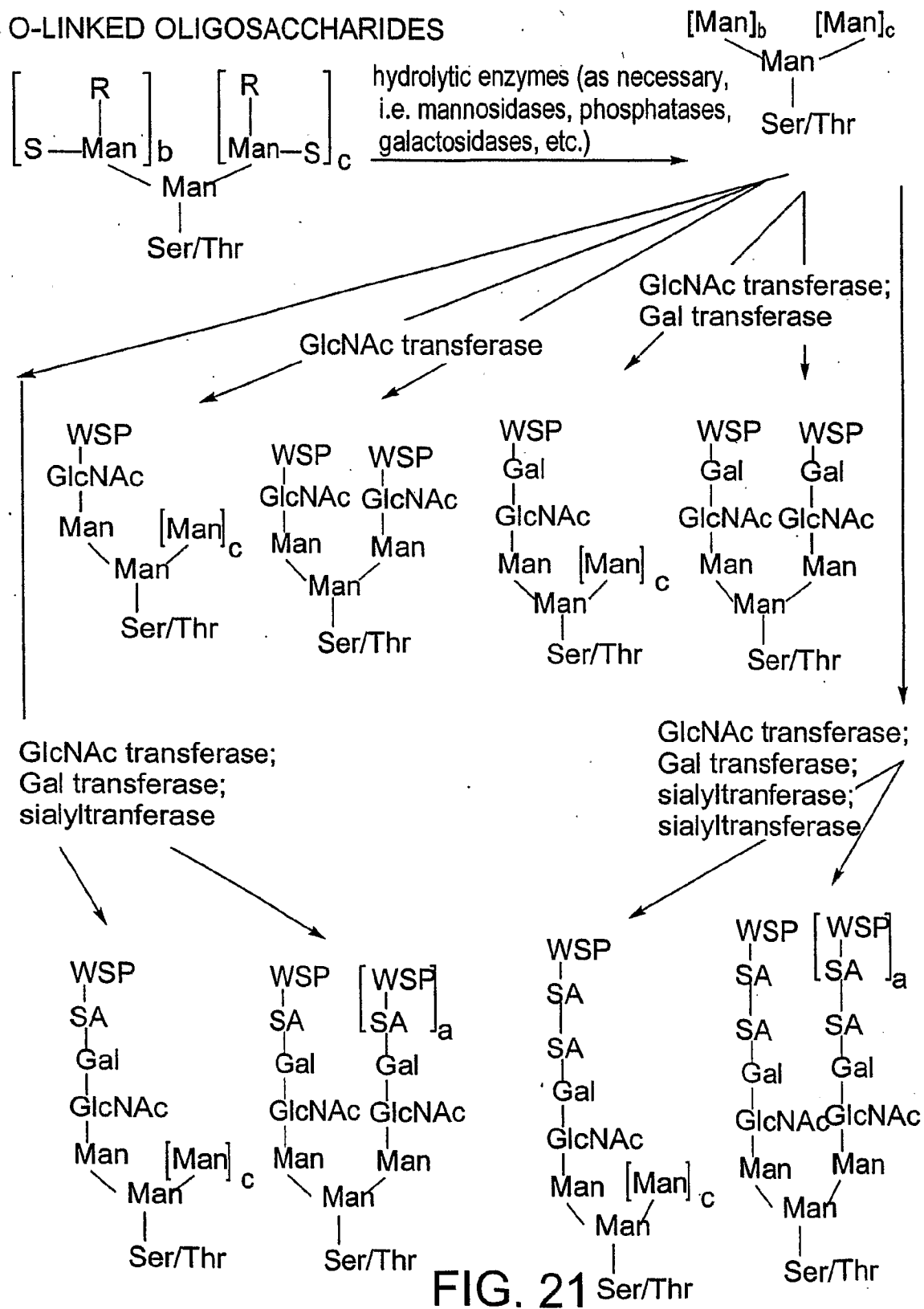
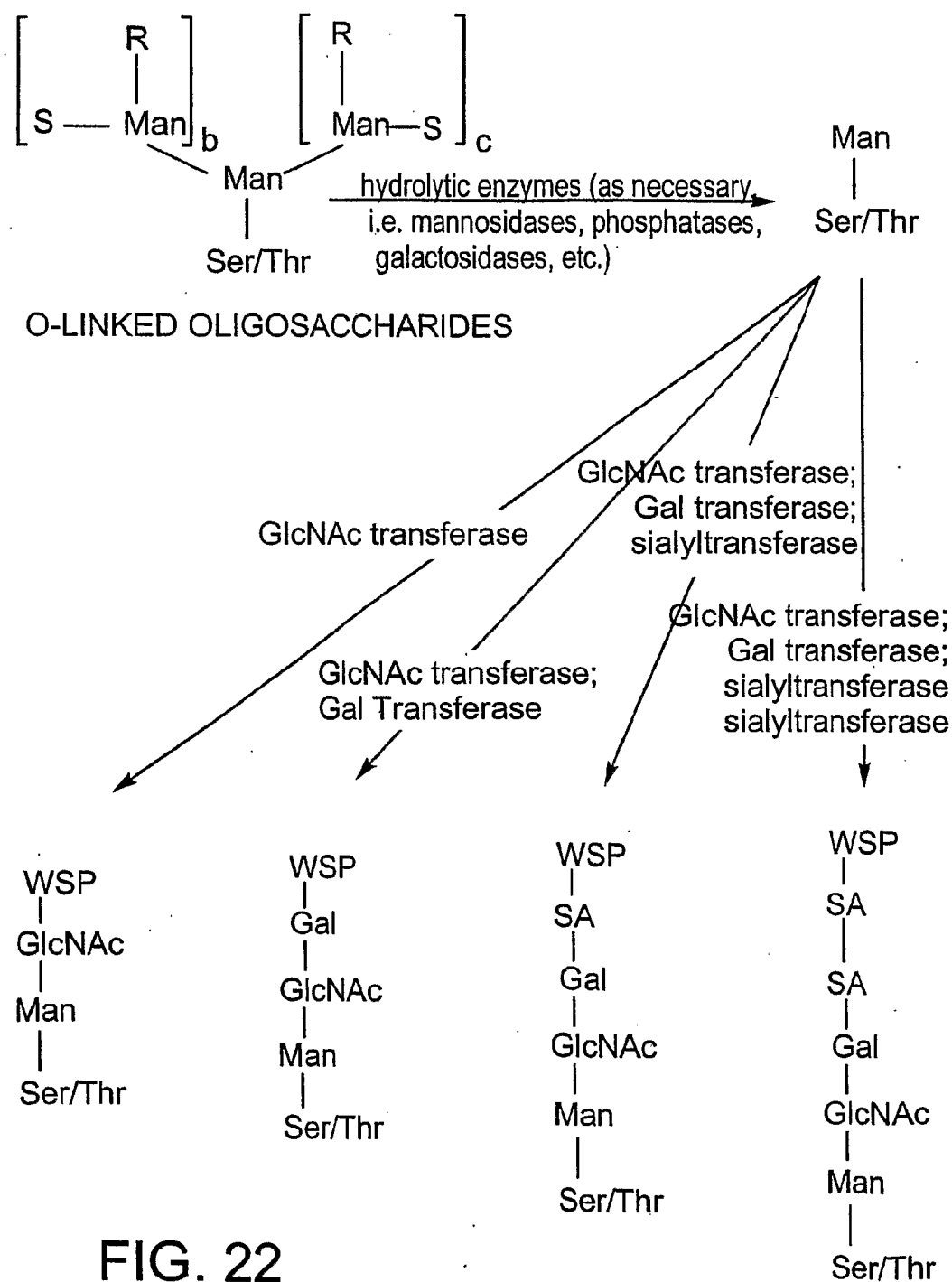
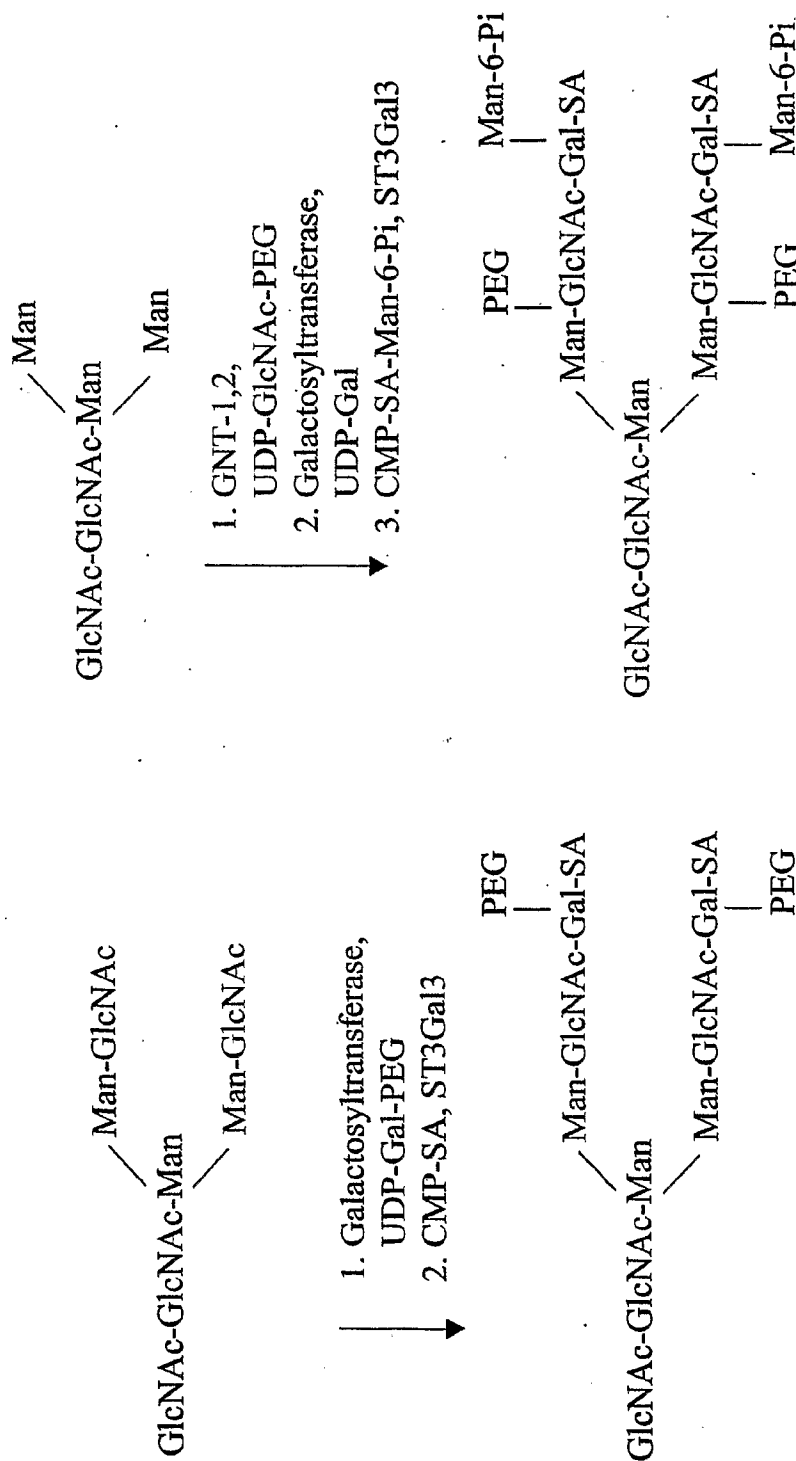


FIG. 21 Ser/Thr

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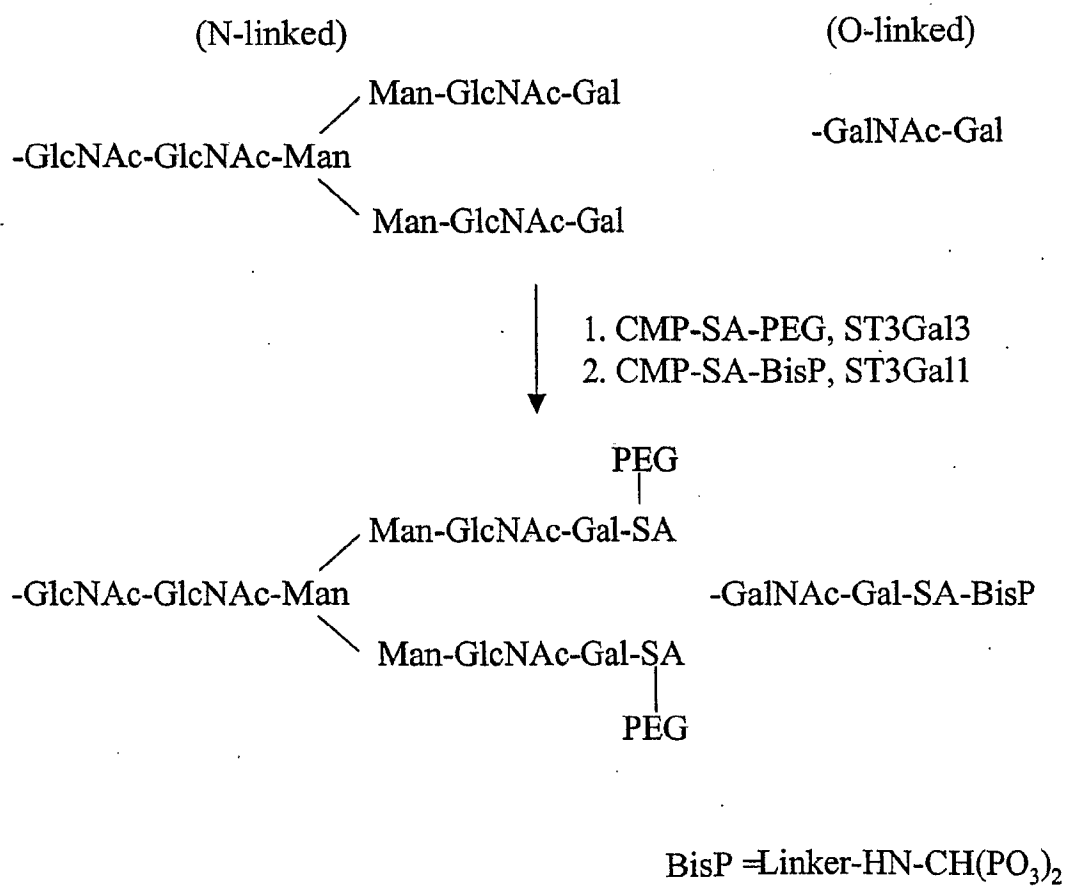


FIG. 23C

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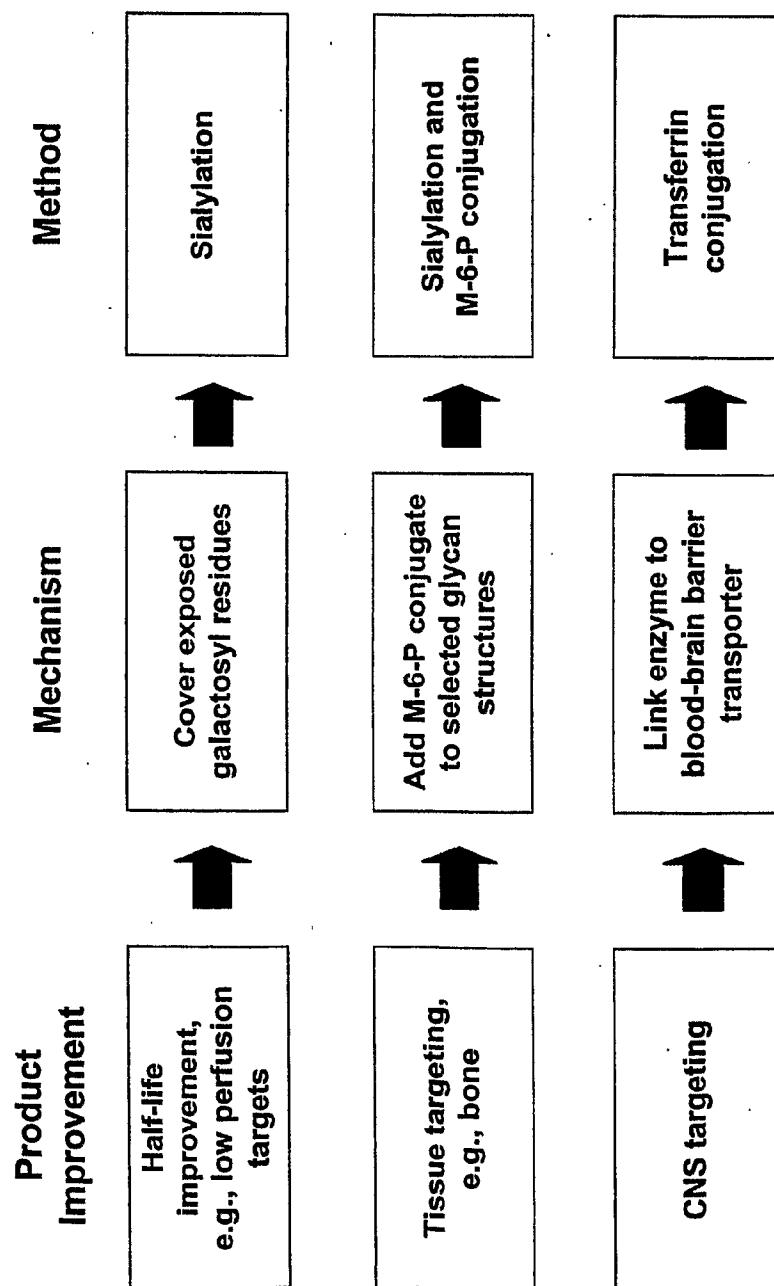


FIG. 24

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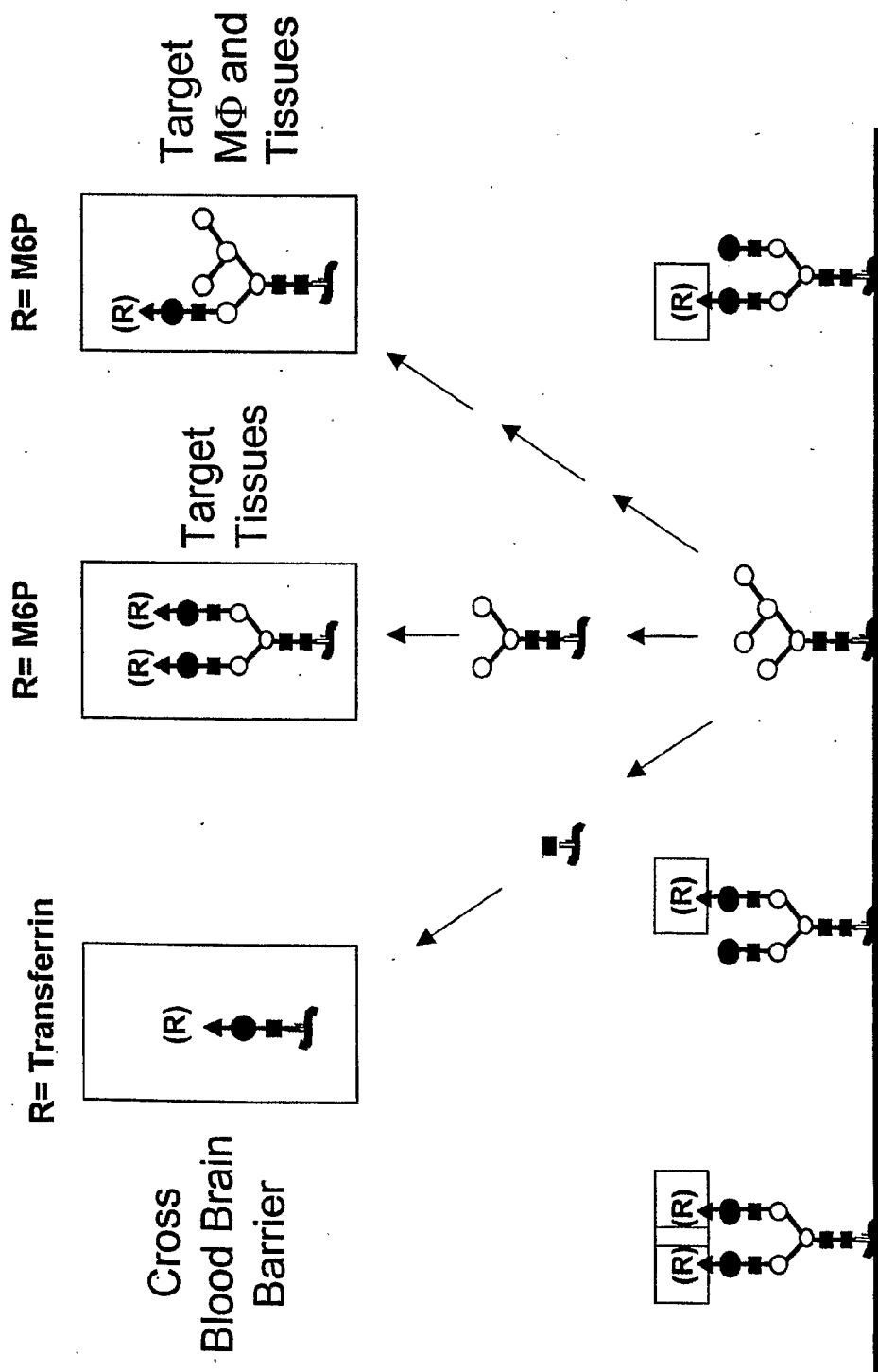


FIG. 25

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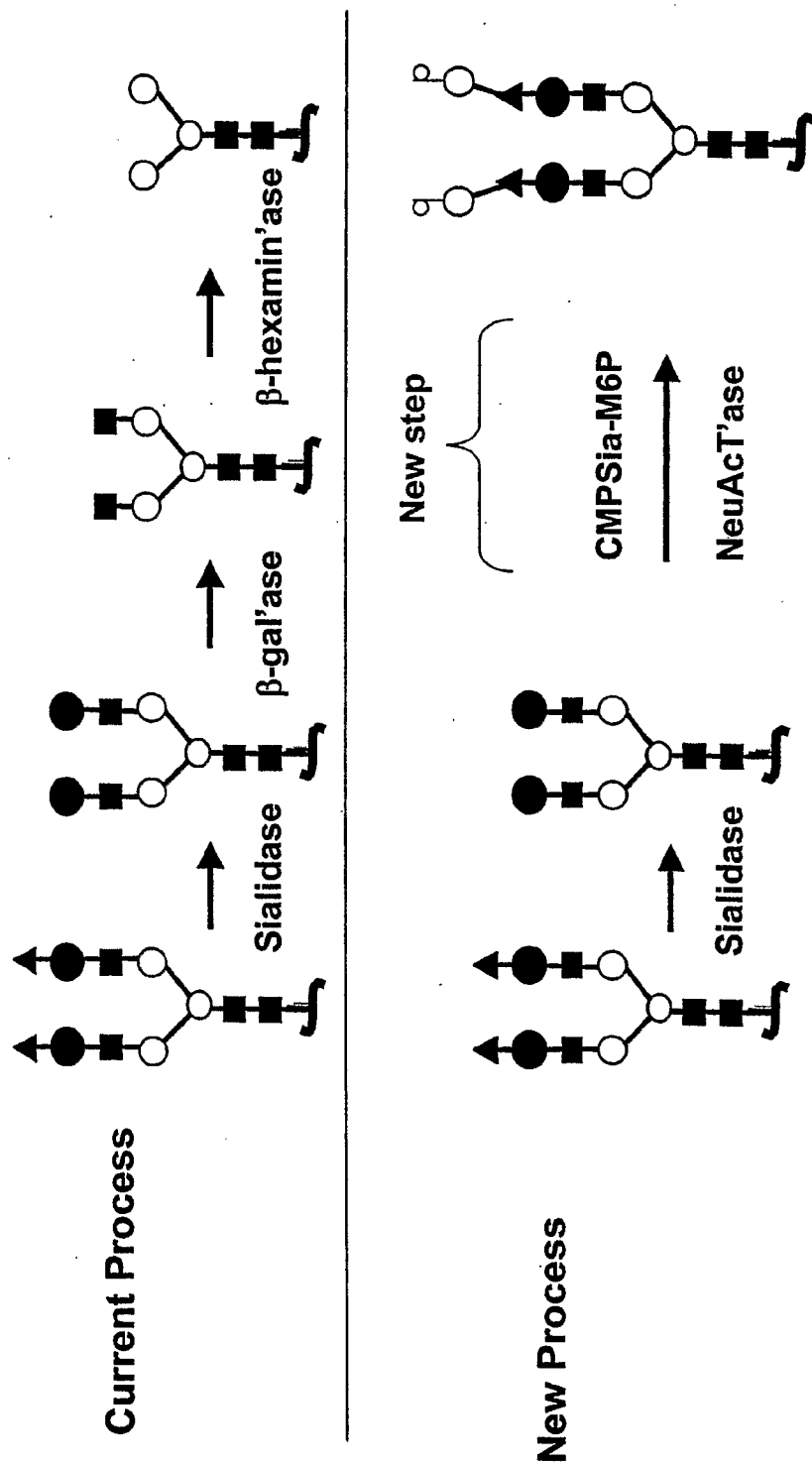


FIG. 26

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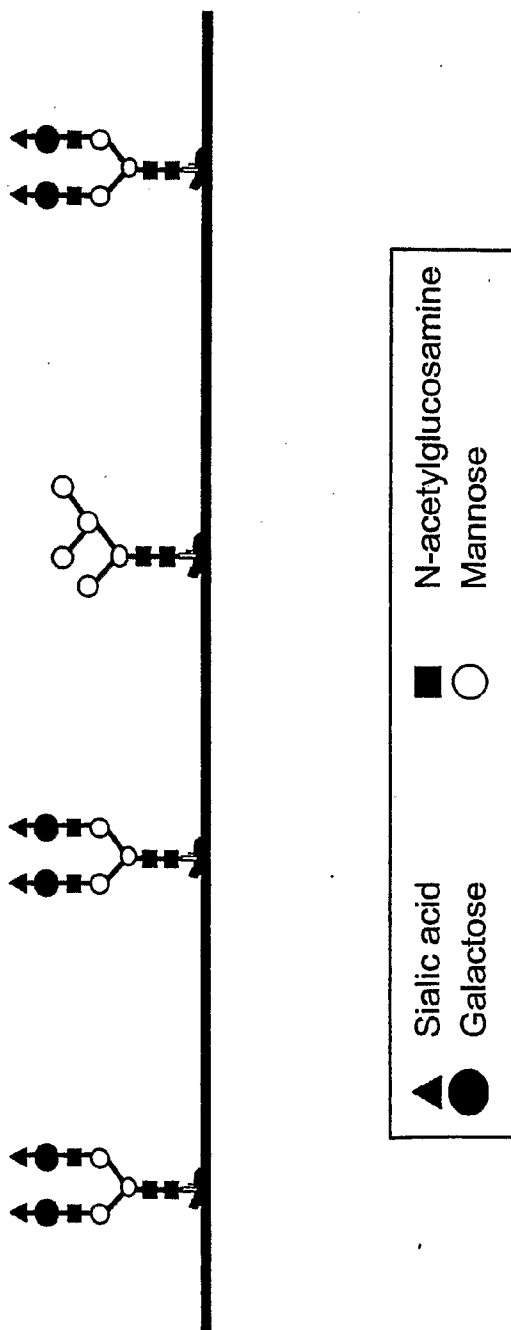


FIG. 27

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12AP1/E5 -- Viventia Biotech	AI-201 -- AutoImmune
1964 -- Aventis	AI-301 -- AutoImmune
20K growth hormone -- AMUR	AIDS vaccine -- ANRS, CIBG, Hesed
28P6/E6 -- Viventia Biotech	Biomed, Hollis-Eden, Rome, United
3-Hydroxyphthaloyl-beta-lactoglobulin --	Biomedical, American Home Products,
4-IBB ligand gene therapy --	Maxygen
64-Cu MAb conjugate TETA-1A3 --	airway receptor ligand -- IC Innovations
Mallinckrodt Institute of Radiology	AJvW 2 -- Ajinomoto
64-Cu MAb conjugate TETA-cT84.66	AK 30 NGF -- Alkermes
64-Cu Trastuzumab TETA conjugate --	Albuferon -- Human Genome Sciences
Genentech	albumin -- Biogen, DSM Anti-Infectives,
A 200 -- Amgen	Genzyme Transgenics, PPL Therapeutics,
A10255 -- Eli Lilly	TranXenoGen, Welfide Corp.
A1PDX -- Hedral Therapeutics	aldesleukin -- Chiron
A6 -- Angstrom	alefacept -- Biogen
aaAT-III -- Genzyme	Alemtuzumab
Abciximab -- Centocor	Allergy therapy -- ALK-Abello/Maxygen,
ABI.001 -- Atlantic BioPharmaceuticals	ALK-Abello/RP Scherer
ABT-828 -- Abbott	allergy vaccines -- Allergy Therapeutics
Accutin	Alnidofibatide -- Aventis Pasteur
Actinohivin	Alnorine -- SRC VB VECTOR
activin -- Biotech Australia, Human	ALP 242 -- Gruenenthal
Therapeutics, Curis	Alpha antitrypsin -- Arriva/Hyland
AD 439 -- Tanox	Immuno/ProMetic/Protease Sciences
AD 519 -- Tanox	Alpha-1 antitrypsin -- Cutter, Bayer, PPL
Adalimumab -- Cambridge Antibody Tech.	Therapeutics, Profile, ZymoGenetics,
Adenocarcinoma vaccine -- Biomira -- NIS	Arriva
Adenosine deaminase -- Enzond	Alpha-1 protease inhibitor -- Genzyme
Adenosine A2B receptor antagonists --	Transgenics, Welfide Corp.
Adenosine Therapeutics	Alpha-galactose fusion protein --
ADP-001 -- Axis Genetics	Immunomedics
AF 13948 -- Affymax	Alpha-galactosidase A -- Research
Afelimomab -- Knoll	Corporation Technologies, Genzyme
AFP-SCAN -- Immunomedics	Alpha-glucosidase -- Genzyme, Novazyme
AG 2195 -- Corixa	Alpha-lactalbumin
agalsidase alfa -- Transkaryotic Therapies	Alpha-L-iduronidase -- Transkaryotic
agalsidase beta -- Genzyme	Therapies, BioMarin
AGENT-- Antisoma	alteplase -- Genentech
AI 300 -- AutoImmune	alvircept sudotox -- NIH
AI-101 -- Teva	ALX-0600, a GLP-2 agonist -- NPS Allelix
AI-102 -- Teva	Corp.

FIG. 28A

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ALX1-11 --sNPS Pharmaceuticals	Anti-alphav β 3 integrin MAb -- Applied
Alzheimer's disease gene therapy	Molecular Evolution
AM-133 -- AMRAD	Anti-angiogenesis monoclonal antibodies --
Amb a 1 immunostim conj. -- Dynavax	KS Biomedix/Schering AG
AMD 3100 -- AnorMED -- NIS	Anti-B4 MAb-DC1 conjugate -- ImmunoGen
AMD 3465 -- AnorMED -- NIS	Anti-B7 antibody PRIMATIZED -- IDEC
AMD 3465 -- AnorMED -- NIS	Anti-B7-1 MAb 16-10A1
AMD Fab -- Genentech	Anti-B7-1 MAb 1G10
Amediplase -- Menarini, Novartis	Anti-B7-2 MAb GL-1
AM-F9	Anti-B7-2-gelonin immunotoxin --
Amoebiasis vaccine	Antibacterials/antifungals --
Amphiregulin -- Octagene	Diversa/IntraBiotics
anakinra -- Amgen	Anti-beta-amyloid monoclonal antibodies --
analgesic -- Nobex	Cambridge Antibody Tech., Wyeth-Ayerst
ancestim -- Amgen	Anti-BLyS antibodies -- Cambridge
AnergiX.RA -- Corixa, Organon	Antibody Tech. /Human Genome Sciences
Angiocidin -- InKine	Antibody-drug conjugates -- Seattle
angiogenesis inhibitors -- ILEX	Genetics/Eos
AngioMab -- Antisoma	Anti-C5 MAb BB5-1 -- Alexion
Angiopoietins -- Regeneron/Procter &	Anti-C5 MAb N19-8 -- Alexion
Gamble	Anti-C8 MAb
angiostatin -- EntreMed	anticancer cytokines -- BioPulse
Angiostatin/endostatin gene therapy --	anticancer matrix -- Telios Integra
Genetix Pharmaceuticals	Anticancer monoclonal antibodies -- ARIUS,
angiotensin-II, topical -- Maret	Immunex
Anthrax -- EluSys Therapeutics/US Army	anticancer peptides -- Maxygen, Micrologix
Medical Research Institute	Anticancer prodrug Tech. -- Alexion
Anthrax vaccine	Antibody Technologies
Anti platelet-derived growth factor D human	anticancer Troy-Bodies -- Affite -- Affitech
monoclonal antibodies -- CuraGen	anticancer vaccine -- NIH
Anti-17-1A MAb 3622W94 --	anticancers -- Epimmune
GlaxoSmithKline	Anti-CCR5/CXCR4 sheep MAb -- KS
Anti-2C4 MAb -- Genentech	Biomedix Holdings
anti-4-1BB monoclonal antibodies -- Bristol-	Anti-CD11a MAb KBA --
Myers Squibb	Anti-CD11a MAb M17
Anti-Adhesion Platform Tech. -- Cytovax	Anti-CD11a MAb TA-3 --
Anti-adipocyte MAb -- Cambridge Antibody	Anti-CD11a MAb WT.1 --
Tech./ObeSys	Anti-CD11b MAb -- Pharmacia
antiallergics -- Maxygen	Anti-CD11b MAb LM2
antiallergy vaccine -- Acambis	Anti-CD154 MAb -- Biogen
Anti-alpha-4-integrin MAb	Anti-CD16-anti-CD30 MAb -- Biotest

FIG. 28B

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Anti-CD18 MAb -- Pharmacia	Anti-CD4 MAb -- Centocor, IDEC
Anti-CD19 MAb B43 --	Pharmaceuticals, Xenova Group
Anti-CD19 MAb -liposomal sodium butyrate conjugate --	Anti-CD4 MAb 16H5
Anti-CD147	Anti-CD4 MAb 4162W94 -- GlaxoSmithKline
Anti-CD19 MAb-saporin conjugate --	Anti-CD4 MAb B-F5 -- Diaclone
Anti-CD19-dsFv-PE38-immunotoxin --	Anti-CD4 MAb GK1-5
Anti-CD2 MAb 12-15 --	Anti-CD4 MAb KT6
Anti-CD2 MAb B-E2 -- Diaclone	Anti-CD4 MAb OX38
Anti-CD2 MAb OX34 --	Anti-CD4 MAb PAP conjugate -- Bristol-Myers Squibb
Anti-CD2 MAb OX54 --	Anti-CD4 MAb RIB 5-2
Anti-CD2 MAb OX55 --	Anti-CD4 MAb W3/25
Anti-CD2 MAb RM2-1	Anti-CD4 MAb YTA 3.1.2
Anti-CD2 MAb RM2-2	Anti-CD4 MAb YTS 177-9
Anti-CD2 MAb RM2-4	Anti-CD40 ligand MAb 5c8 -- Biogen
Anti-CD20 MAb BCA B20	Anti-CD40 MAb
Anti-CD20-anti-Fc alpha RI bispecific MAb -- Medarex, Tenovus	Anti-CD40 MAb 5D12 -- Tanox
Anti-CD22 MAb-saporin-6 complex --	Anti-CD44 MAb A3D8
Anti-CD3 immunotoxin --	Anti-CD44 MAb GKWA3
Anti-CD3 MAb 145-2C11 -- Pharming	Anti-CD44 MAb IM7
Anti-CD3 MAb CD4IgG conjugate -- Genentech	Anti-CD44 MAb KM81
Anti-CD3 MAb humanised -- Protein Design, RW Johnson	Anti-CD44 variant monoclonal antibodies -- Corixa/Hebrew University
Anti-CD3 MAb WT32	Anti-CD45 MAb BC8-I-131
Anti-CD3 MAb-ricin-chain-A conjugate --	Anti-CD45RB MAb
Anti-CD3 MAb-xanthine-oxidase conjugate --	Anti-CD48 MAb HuLy-m3
Anti-CD30 MAb BerH2 -- Medac	Anti-CD48 MAb WM-63
Anti-CD30 MAb-saporin conjugate	Anti-CD5 MAb -- Becton Dickinson
Anti-CD30-scFv-ETA'-immunotoxin	Anti-CD5 MAb OX19
Anti-CD38 MAb AT13/5	Anti-CD6 MAb
Anti-CD38 MAb-saporin conjugate	Anti-CD7 MAb-PAP conjugate
Anti-CD3-anti-CD19 bispecific MAb	Anti-CD7 MAb-ricin-chain-A conjugate
Anti-CD3-anti-EGFR MAb	Anti-CD8 MAb -- Amerimmune, Cytodyn,
Anti-CD3-anti-interleukin-2-receptor MAb	Becton Dickinson
Anti-CD3-anti-MOV18 MAb -- Centocor	Anti-CD8 MAb 2-43
Anti-CD3-anti-SCLC bispecific MAb	Anti-CD8 MAb OX8
Anti-CD4 idiotype vaccine	Anti-CD80 MAb P16C10 -- IDEC
	Anti-CD80 MAb P7C10 -- ID Vaccine
	Anti-CD8-idarubicin conjugate
	Anti-CEA MAb CE-25
	Anti-CEA MAb MN 14 -- Immunomedics

FIG. 28C

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Anti-CEA MAb MN14-PE40 conjugate -- Immunomedics	Anti-heparanase human monoclonal antibodies -- Oxford Glycosciences/Medarex
Anti-CEA MAb T84.66-interleukin-2 conjugate	Anti-hepatitis C virus human monoclonal antibodies -- XTL Biopharmaceuticals
Anti-CEA sheep MAb -- KS Biomedix Holdings	Anti-HER-2 antibody gene therapy
Anti-cell surface monoclonal antibodies -- Cambridge Antibody Tech. /Pharmacia	Anti-herpes antibody -- Epicyte
Anti-c-erbB2-anti-CD3 bifunctional MAb -- Otsuka	Anti-HIV antibody -- Epicyte
Anti-CMV MAb -- Scotgen	anti-HIV catalytic antibody -- Hersed Biomed
Anti-complement	anti-HIV fusion protein -- Idun
Anti-CTLA-4 MAb	anti-HIV proteins -- Cangene
Anti-EGFR catalytic antibody -- Hersed Biomed	Anti-HM1-24 MAb -- Chugai
anti-EGFR immunotoxin -- IVAX	Anti-hR3 MAb
Anti-EGFR MAb -- Abgenix	Anti-Human-Carcinoma-Antigen MAb -- Epicyte
Anti-EGFR MAb 528	Anti-ICAM-1 MAb -- Boehringer Ingelheim
Anti-EGFR MAb KSB 107 -- KS Biomedix	Anti-ICAM-1 MAb 1A-29 -- Pharmacia
Anti-EGFR MAb-DM1 conjugate -- ImmunoGen	Anti-ICAM-1 MAb HA58
Anti-EGFR MAb-LA1 --	Anti-ICAM-1 MAb YN1/1.7.4
Anti-EGFR sheep MAb -- KS Biomedix	Anti-ICAM-3 MAb ICM3 -- ICOS
Anti-FAP MAb F19-I-131	Anti-idiotypic breast cancer vaccine 11D10
Anti-Fas IgM MAb CH11	Anti-idiotypic breast cancer vaccine ACA14C5 --
Anti-Fas MAb Jo2	Anti-idiotypic cancer vaccine -- ImClone Systems/Merck KGaA ImClone, Viventia Biotech
Anti-Fas MAb RK-8	Anti-idiotypic cancer vaccine 1A7 -- Titan
Anti-Flt-1 monoclonal antibodies -- ImClone	Anti-idiotypic cancer vaccine 3H1 -- Titan
Anti-fungal peptides -- State University of New York	Anti-idiotypic cancer vaccine TriAb -- Titan
antifungal tripeptides -- BTG	Anti-idiotypic Chlamydia trachomatis vaccine
Anti-ganglioside GD2 antibody-interleukin-2 fusion protein -- Lexigen	Anti-idiotypic colorectal cancer vaccine -- Novartis
Anti-GM2 MAb -- Kyowa	Anti-idiotypic colorectal cancer vaccine -- Onyvax
Anti-GM-CSF receptor monoclonal antibodies -- AMRAD	Anti-idiotypic melanoma vaccine -- IDEC Pharmaceuticals
Anti-gp130 MAb -- Tosoh	Anti-idiotypic ovarian cancer vaccine ACA 125
Anti-HCA monoclonal antibodies -- AltaRex/Epigen	Anti-idiotypic ovarian cancer vaccine AR54 - AltaRex
Anti-hCG antibodies -- Abgenix/AVI BioPharma	

FIG. 28D

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Anti-idiotypic ovarian cancer vaccine CA-125 – AltaRex, Biomira	Anti-L-selectin monoclonal antibodies -- Protein Design Labs, Abgenix, Stanford University
Anti-IgE catalytic antibody -- Hersed Biomed	Anti-MBL monoclonal antibodies -- Alexion/Brigham and Women's Hospital
Anti-IgE MAb E26 -- Genentech	Anti-MHC monoclonal antibodies
Anti-IGF-1 MAb	Anti-MIF antibody humanised – IDEC, Cytokine PharmaSciences
anti-inflammatory -- GeneMax	Anti-MRSA/VRSA sheep MAb -- KS Biomedix Holdings
anti-inflammatory peptide -- BTG	Anti-mu MAb -- Novartis
anti-integrin peptides -- Burnha	Anti-MUC-1 MAb
Anti-interferon-alpha-receptor MAb 64G12 -- Pharma Pacific Management	Anti-MUC 18
Anti-interferon-gamma MAb -- Protein Design Labs	Anti-Nogo-A MAb IN1
Anti-interferon-gamma polyclonal antibody - Advanced Biotherapy	Anti-nuclear autoantibodies -- Procyon
Anti-interleukin-10 MAb –	Anti-ovarian cancer monoclonal antibodies - Dompe
Anti-interleukin-12 MAb –	Anti-p185 monoclonal antibodies
Anti-interleukin-1-beta polyclonal antibody -- R&D Systems	Anti-p43 MAb
Anti-interleukin-2 receptor MAb 2A3	Antiparasitic vaccines
Anti-interleukin-2 receptor MAb 33B3-1 -- Immunotech	Anti-PDGF/bFGF sheep MAb -- KS Biomedix
Anti-interleukin-2 receptor MAb ART-18	Anti-properdin monoclonal antibodies -- Abgenix/Gliatech
Anti-interleukin-2 receptor MAb LO-Tact-1	Anti-PSMA (prostate specific membrane antigen)
Anti-interleukin-2 receptor MAb Mikbeta1	Anti-PSMA MAb J591 -- BZL Biologics
Anti-interleukin-2 receptor MAb NDS61	Anti-Rev MAb gene therapy –
Anti-interleukin-4 MAb 11B11	Anti-RSV antibodies – Epicyte, Intracell
Anti-interleukin-5 MAb -- Wallace Laboratories	Anti-RSV monoclonal antibodies -- Medarex/MedImmune, Applied Molecular Evolution/MedImmune
Anti-interleukin-6 MAb – Centocor, Diaclone, Pharmadigm	Anti-RSV MAb, inhalation -- Alkermes/MedImmune
Anti-interleukin-8 MAb -- Abgenix	Anti-RT gene therapy
Anti-interleukin-8 MAb – Xenotech	Antisense K-ras RNA gene therapy
Anti-JL1 MAb	Anti-SF-25 MAb
Anti-Klebsiella sheep MAb -- KS Biomedix Holdings	Anti-sperm antibody -- Epicyte
Anti-Laminin receptor MAb-liposomal doxorubicin conjugate	Anti-Tac(Fv)-PE38 conjugate
Anti-LCG MAb -- Cytoclonal	Anti-TAPA/CD81 MAb AMP1
Anti-lipopolysaccharide MAb -- VitaResc	Anti-tat gene therapy

FIG. 28E

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Anti-TCR-alphabeta MAb H57-597	AOP-RANTES -- Senetek
Anti-TCR-alphabeta MAb R73	Apan-CH -- Praecis Pharmaceuticals
Anti-tenascin MAb BC-4-I-131	APC-8024 -- Demegen
Anti-TGF-beta human monoclonal antibodies -- Cambridge Antibody Tech., Genzyme	ApoA-1 -- Milano, Pharmacia
Anti-TGF-beta MAb 2G7 -- Genentech	Apogen -- Alexion
Antithrombin III -- Genzyme Transgenics, Aventis, Bayer, Behringwerke, CSL, Myriad	apolipoprotein A1 -- Avanir
Anti-Thy1 MAb	Apolipoprotein E -- Bio-Tech. General
Anti-Thy1.1 MAb	Applaggin -- Biogen
Anti-tissue factor/factor VIIA sheep MAb -- KS Biomedix	aprotinin -- ProdiGene
Anti-TNF monoclonal antibodies -- Centocor, Chiron, Peptech, Pharacia, Serono	APT-070C -- AdProTech
Anti-TNF sheep MAb -- KS Biomedix Holdings	AR 177 -- Aronex Pharmaceuticals
Anti-TNFalpha MAb -- Genzyme	AR 209 -- Aronex Pharmaceuticals, Antigenics
Anti-TNFalpha MAb B-C7 -- Diaclone	AR545C
Anti-tooth decay MAb -- Planet BioTech.	ARGENT gene delivery systems -- ARIAD
Anti-TRAIL receptor-1 MAb -- Takeda	Arresten
Antitumour RNases -- NIH	ART-123 -- Asahi Kasei
Anti-VCAM MAb 2A2 -- Alexion	arylsulfatase B -- BioMarin
Anti-VCAM MAb 3F4 -- Alexion	Arylsulfatase B, Recombinant human -- BioMarin
Anti-VCAM-1 MAb	AS 1051 -- Ajinomoto
Anti-VEC MAb -- ImClone	ASI-BCL -- Intracell
Anti-VEGF MAb -- Genentech	Asparaginase - Merck
Anti-VEGF MAb 2C3	ATL-101 -- Alizyme
Anti-VEGF sheep MAb -- KS Biomedix Holdings	Atrial natriuretic peptide -- Pharis
Anti-VLA-4 MAb HP1/2 -- Biogen	Aurintricarboxylic acid-high molecular weight
Anti-VLA-4 MAb PS/2	Autoimmune disorders -- GPC Biotech/MorphoSys
Anti-VLA-4 MAb R1-2	Autoimmune disorders and transplant rejection -- Bristol-Myers Squibb/Genzyme
Anti-VLA-4 MAb TA-2	Tra
Anti-VAP-1 human MAb	Autoimmune disorders/cancer -- Abgenix/Chiron, CuraGen
Anti-VRE sheep MAb -- KS Biomedix Holdings	Autotaxin
ANUP -- TranXenoGen	Avicidin -- NeoRx
ANUP-1 -- Pharis	axogenesis factor-1 -- Boston Life Sciences
	Axokine -- Regeneron
	B cell lymphoma vaccine -- Biomira
	B7-1 gene therapy --
	BABS proteins -- Chiron

FIG. 28F

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BAM-002 -- Novelos Therapeutics	BMP 2 -- Genetics Institute/Medtronic-
Basiliximab (anti CD25 MAb) -- Novartis	Sofamor Danek, Genetics Institute/
Bay-16-9996 -- Bayer	Collagenesis, Genetics
Bay-39-9437 -- Bayer	Institute/Yamanouch
Bay-50-4798 -- Bayer	BMP 2 gene therapy
BB-10153 -- British Biotech	BMP 52 -- Aventis Pasteur, Biopharm
BBT-001 -- Bolder BioTech.	BMP-2 -- Genetics Institute
BBT-002 -- Bolder BioTech.	BMS 182248 -- Bristol-Myers Squibb
BBT-003 -- Bolder BioTech.	BMS 202448 -- Bristol-Myers Squibb
BBT-004 -- Bolder BioTech.	bone growth factors -- IsoTis
BBT-005 -- Bolder BioTech.	BPC-15 -- Pfizer
BBT-006 -- Bolder BioTech.	brain natriuretic peptide --
BBT-007 -- Bolder BioTech.	Breast cancer -- Oxford
BCH-2763 -- Shire	GlycoSciences/Medarex
BCSF -- Millenium Biologix	Breast cancer vaccine -- Therion Biologics,
BDNF -- Regeneron -- Amgen	Oregon
Becaplermin -- Johnson & Johnson, Chiron	BSSL -- PPL Therapeutics
Bectumomab -- Immunomedics	BST-2001 -- BioStratum
Beriplast -- Aventis	BST-3002 -- BioStratum
Beta-adrenergic receptor gene therapy --	BTI 322 --
University of Arkansas	butyrylcholinesterase -- Shire
bFGF -- Scios	C 6822 -- COR Therapeutics
BI 51013 -- Behringwerke AG	C1 esterase inhibitor -- Pharming
BIBH 1 -- Boehringer Ingelheim	C3d adjuvant -- AdProTech
BIM-23190 -- Beaufour-Ipsen	CAB-2.1 -- Millennium
birch pollen immunotherapy -- Pharmacia	calcitonin -- Inhale Therapeutics Systems,
bispecific fusion proteins -- NIH	Aventis, Genetronics, TranXenoGen,
Bispecific MAb 2B1 -- Chiron	Unigene, Rhone Poulenc Rohrer
Bitistatin	calcitonin -- oral -- Nobex, Emisphere,
BIWA 4 -- Boehringer Ingelheim	Pharmaceutical Discovery
blood substitute -- Northfield, Baxter Intl.	Calcitonin gene-related peptide -- Asahi
BLP-25 -- Biomira	Kasei -- Unigene
BLS-0597 -- Boston Life Sciences	calcitonin, human -- Suntory
BLyS -- Human Genome Sciences	calcitonin, nasal -- Novartis, Unigene
BLyS radiolabelled -- Human Genome	calcitonin, Panoderm -- Elan
Sciences	calcitonin, Peptitrol -- Shire
BM 06021 -- Boehringer Mannheim	calcitonin, salmon -- Therapicon
BM-202 -- BioMarin	calin -- Biopharm
BM-301 -- BioMarin	Calphobindin I
BM-301 -- BioMarin	calphobindin I -- Kowa
BM-302 -- BioMarin	calreticulin -- NYU

FIG. 28G

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Campath-1G	CD4 fusion toxin -- Senetek
Campath-1M	CD4 IgG -- Genentech
cancer therapy -- Cangene	CD4 receptor antagonists --
cancer vaccine -- Aixlie, Aventis Pasteur,	Pharmacoepia/Progenics
Center of Molecular Immunology, YM	CD4 soluble -- Progenics
BioSciences, Cytos, Genzyme,	CD4, soluble -- Genzyme Transgenics
Transgenics, GlobelImmune, Igeneon,	CD40 ligand -- Immunex
ImClone, Virogenetics, InterCell, Iomai,	CD4-ricin chain A -- Genentech
Jenner Biotherapies, Memorial Sloan-	CD59 gene therapy -- Alexion
Kettering Cancer Center, Sydney Kimmel	CD8 TIL cell therapy -- Aventis Pasteur
Cancer Center, Novavax, Protein	CD8, soluble -- Avidex
Sciences, Argonex, SIGA	CD95 ligand -- Roche
Cancer vaccine ALVAC-CEA B7.1 --	CDP 571 -- Celltech
Aventis Pasteur/Therion Biologics	CDP 850 -- Celltech
Cancer vaccine CEA-TRICOM -- Aventis	CDP-860 (PEG-PDGF MAb) -- Celltech
Pasteur/Therion Biologics	CDP 870 -- Celltech
Cancer vaccine gene therapy -- Cantab	CDS-1 -- Ernest Orlando
Pharmaceuticals	Cedelizumab -- Ortho-McNeil
Cancer vaccine HER-2/neu -- Corixa	Cetermin -- Insmad
Cancer vaccine THERATOPE -- Biomira	CETP vaccine -- Avant
cancer vaccine, PolyMASC -- Valentis	Cetrorelix
Candida vaccine -- Corixa, Inhibitex	Cetuximab
Canstatin -- ILEX	CGH 400 -- Novartis
CAP-18 -- Panorama	CGP 42934 -- Novartis
Cardiovascular gene therapy -- Collateral	CGP 51901 -- Tanox
Therapeutics	CGRP -- Unigene
carperitide -- Suntory	CGS 27913 -- Novartis
Casocidin-1 -- Pharis	CGS 32359 -- Novartis
CAT 152 -- Cambridge Antibody Tech.	Chagas disease vaccine -- Corixa
CAT 192 -- Cambridge Antibody Tech.	chemokines -- Immune Response
CAT 213 -- Cambridge Antibody Tech.	CHH 380 -- Novartis
Catalase-- Enzon	chitinase -- Genzyme, ICOS
Cat-PAD -- Circassia	Chlamydia pneumoniae vaccine -- Antex
CB 0006 -- Celltech	Biologics
CCK(27-32)-- Akzo Nobel	Chlamydia trachomatis vaccine -- Antex
CCR2-64I -- NIH	Biologics
CD, Procept -- Paligent	Chlamydia vaccine -- GlaxoSmithKline
CD154 gene therapy	Cholera vaccine CVD 103-HgR -- Swiss
CD39 -- Immunex	Serum and Vaccine Institute Berne
CD39-L2 -- Hyseq	Cholera vaccine CVD 112 -- Swiss Serum
CD39-L4 -- Hyseq	and Vaccine Institute Berne

FIG. 28H

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Cholera vaccine inactivated oral -- SBL Vaccin	CRL 1605 -- CytRx
Chrysalin -- Chrysalis BioTech.	CS-560 -- Sankyo
CI-782 -- Hitachi Kase	CSF -- ZymoGenetics
Ciliary neurotrophic factor -- Fidia, Roche	CSF-G -- Hangzhou, Dong-A, Hanmi
CIM project -- Active Biotech	CSF-GM -- Cangene, Hunan, LG Chem
CL 329753 -- Wyeth-Ayerst	CSF-M -- Zarix
CL22, Cobra -- ML Laboratories	CT 1579 -- Merck Frosst
Clenoliximab -- IDEC	CT 1786 -- Merck Frosst
Clostridium difficile antibodies -- Epicyte	CT-112 [^] -- BTG
clotting factors -- Octagene	CTB-134L -- Xenova
CMB 401 -- Celltech	CTC-111 -- Kaketsuken
CNTF -- Sigma-Tau	CTGF -- FibroGen
Cocaine abuse vaccine -- Cantab, ImmuLogic, Scripps	CTLA4-Ig -- Bristol-Myers Squibb
coccidiomycosis vaccine -- Arizo	CTLA4-Ig gene therapy --
collagen -- Type I -- Pharming	CTP-37 -- AVI BioPharma
Collagen formation inhibitors -- FibroGen	C-type natriuretic peptide -- Suntory
Collagen/hydroxyapatite/bone growth factor -- Aventis Pasteur, Biopharm, Orquest	CVS 995 -- Corvas Intl.
collagenase -- BioSpecifics	CX 397 -- Nikko Kyodo
Colorectal cancer vaccine -- Wistar Institute	CY 1747 -- Epimmune
Component B, Recombinant -- Serono	CY 1748 -- Epimmune
Connective tissue growth factor inhibitors -- FibroGen/Taisho	Cyanovirin-N
Contortrostatin	Cystic fibrosis therapy -- CBR/IVAX
contraceptive vaccine -- Zonagen	CYT 351
Contraceptive vaccine hCG	cytokine Traps -- Regeneron
Contraceptive vaccine male reversible -- IMMUCON	cytokines -- Enzon, Cytoclonal
Contraceptive vaccine zona pellucida -- Zonagen	Cytomegalovirus glycoprotein vaccine -- Chiron, Aquila Biopharmaceuticals, Aventis Pasteur, Virogenetics
Copper-64 labelled MAb TETA-1A3 -- NCI	Cytomegalovirus vaccine live -- Aventis Pasteur
Coralyne	Cytosine deaminase gene therapy -- GlaxoSmithKline
Corsevin M	DA-3003 -- Dong-A
C-peptide analogues -- Schwarz	DAB389interleukin-6 -- Senetek
CPI-1500 -- Consensus	DAB389interleukin-7
CRF -- Neurobiological Tech.	DAC:GLP-2 -- ConjuChem, Inc.
cRGDfV pentapeptide --	Daclizumab (anti-IL2R MAb) -- Protein Design Labs
CRL 1095 -- CytRx	DAMP [^] -- Incyte Genomics
CRL 1336 -- CytRx	Daniplestim -- Pharmacia
	darbepoetin alfa -- Amgen

FIG. 28I

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DBI-3019 -- Diabetogen	dural graft matrix -- Integra
DCC -- Genzyme	Duteplase -- Baxter Intl.
DDF -- Hyseq	DWP-401 -- Daewoong
decorin -- Integra, Telios	DWP-404 -- Daewoong
defensins -- Large Scale Biology	DWP-408 -- Daewoong
DEGR-VIIa	Dx 88 (Epi-KAL2) -- Dyax
Delimmunised antibody 3B6/22 AGEN	Dx 890 (elastin inhibitors) -- Dyax
Deimmunised anti-cancer antibodies -- Biovation/Viragen	E coli O157 vaccine -- NIH
Dendroamide A	E21-R -- BresaGen
Dengue vaccine -- Bavarian Nordic, Merck	Eastern equine encephalitis virus vaccine --
denileukin diftitox -- Ligand	Echicetin --
DES-1101 -- Desmos	Echinhibin 1 --
desirudin -- Novartis	Echistatin -- Merck
desmopressin -- Unigene	Echitamine --
Desmoteplase -- Merck, Schering AG	Ecromeximab -- Kyowa Hakko
Destabilase	EC-SOD -- PPL Therapeutics
Diabetes gene therapy -- DeveloGen, Pfizer	Eculizumab (5G1.1) -- Alexion
Diabetes therapy -- Crucell	EDF -- Ajinomoto
Diabetes type 1 vaccine -- Diamyd Therapeutics	EDN derivative -- NIH
DiaCIM -- YM BioSciences	EDNA -- NIH
dialytic oligopeptides -- Research Corp	Edobacomab -- XOMA
Diamyd -- Diamyd Therapeutics	Edrecolomab -- Centocor
DiaPep227 -- Pepgen	EF 5077
DiavaX -- Corixa	Efalizumab -- Genentech
Digoxin MAb -- Glaxo	EGF fusion toxin -- Seragen, Ligand
Diphtheria tetanus pertussis-hepatitis B vaccine -- GlaxoSmithKline	EGF-P64k vaccine -- Center of Molecular Immunology
DIR therapy -- Solis Therapeutics --	EL 246 -- LigoCyte
DNase -- Genentech	elastase inhibitor -- Synergen
Dornase alfa -- Genentech	elcatonin -- Therapicon
Dornase alfa, inhalation -- Genentech	EMD 72000 -- Merck KGaA
Doxorubicin-anti-CEA MAb conjugate -- Immunomedics	Emdogain -- BIORA
DP-107 -- Trimeris	emfilermin -- AMRAD
drotrecogin alfa -- Eli Lilly	Emoctakin -- Novartis
DTctGMCSF	enamel matrix protein -- BIORA
DTP-polio vaccine -- Aventis Pasteur	Endo III -- NYU
DU 257-KM231 antibody conjugate -- Kyowa	endostatin -- EntreMed, Pharis
	Enhancins -- Micrologix
	Enlimomab -- Isis Pharm.
	Enoxaparin sodium -- Pharmuka

FIG. 28J

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enzyme linked antibody nutrient depletion therapy -- KS Biomedix Holdings	Factor IX gene therapy -- Cell Genesys
Eosinophil-derived neutralizing agent -- EP-51216 -- Asta Medica	Factor VII -- Novo Nordisk, Bayer, Baxter Intl.
EP-51389 -- Asta Medica	Factor VIIa -- PPL Therapeutics, ZymoGenetics
EPH family ligands -- Regeneron	Factor VIII -- Bayer Genentech, Beaufour-Ipsen, CLB, Inex, Octagen, Pharmacia, Pharming
Epidermal growth factor -- Hitachi Kasei, Johnson & Johnson	Factor VIII -- PEGylated -- Bayer
Epidermal growth factor fusion toxin -- Senetek	Factor VIII fragments -- Pharmacia
Epidermal growth factor-genistein -- EPI-HNE-4 -- Dyax	Factor VIII gene therapy -- Targeted Genetics
EPI-KAL2 -- Dyax	Factor VIII sucrose formulation -- Bayer, Genentech
Epoetin-alfa -- Amgen, Dragon Pharmaceuticals, Nanjing Huaxin	Factor VIII-2 -- Bayer
Epratuzumab -- Immunomedics	Factor VIII-3 -- Bayer
Epstein-Barr virus vaccine -- Aviron/SmithKline Beecham, Bioresearch	Factor Xa inhibitors -- Merck, Novo Nordisk, Mochida
Eptacog alfa -- Novo Nordisk	Factor XIII -- ZymoGenetics
Eptifibatide -- COR Therapeutics	Factors VIII and IX gene therapy -- Genetics Institute/Targeted Genetics
erb-38 --	Famoxin -- Genset
Erlizumab -- Genentech	Fas (delta) TM protein -- LXR BioTech.
erythropoietin -- Alkermes, ProLease, Dong-A, Elanex, Genetics Institute, LG Chem, Protein Sciences, Serono, Snow Brand, SRC VB VECTOR, Transkaryotic Therapies	Fas TR -- Human Genome Sciences
Erythropoietin Beta -- Hoffman La Roche	Felvizumab -- Scotgen
Erythropoietin/Epoetin alfa -- Chugai	FFR-VIIa -- Novo Nordisk
Escherichia coli vaccine -- North American Vaccine, SBL Vaccin, Swiss Serum and Vaccine Institute Berne	FG-001 -- F-Gene
etanercept -- Immunex	FG-002 -- F-Gene
examorelin -- Mediolanum	FG-004 -- F-Gene
Exendin 4 -- Amylin	FG-005 -- F-Gene
exonuclease VII	FGF + fibrin -- Repair
F 105 -- Centocor	Fibrimage -- Bio-Tech. General
F-992 -- Fornix	fibrin-binding peptides -- ISIS Innovation
Factor IX -- Alpha Therapeutics, Welfide Corp., CSL, enetics Institute/AHP, Pharmacia, PPL Therapeutics	fibrinogen -- PPL Therapeutics, Pharming
	fibroblast growth factor -- Chiron, NYU, Ramot, ZymoGenetics
	fibrolase conjugate -- Schering AG
	Filgrastim -- Amgen
	filgrastim -- PDA modified -- Xencor
	FLT-3 ligand -- Immunex
	FN18 CRM9 --

FIG. 28K

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follistatin -- Biotech Australia, Human Therapeutics	Glucocerebrosidase -- Genzyme
follitropin alfa -- Alkermes, ProLease, PowderJect, Serono, Akzo Nobel	glutamate decarboxylase -- Genzyme Transgenics
Follitropin Beta -- Bayer, Organon	Glycoprotein S3 -- Kureha
FP 59	GM-CSF -- Immuhex
FSH -- Ferring	GM-CSF tumour vaccine -- PowderJect
FSH + LH -- Ferring	GnRH immunotherapeutic -- Protherics
F-spondin -- CeNeS	Goserelin (LhRH antagonist) -- AstraZeneca
fusion protein delivery system -- UAB Research Foundation	gp75 antigen -- ImClone
fusion toxins -- Boston Life Sciences	gp96 -- Antigenics
G 5598 -- Genentech	GPI 0100 -- Galenica
GA-II -- Transkaryotic Therapies	GR 4991W93 -- GlaxoSmithKline
Gamma-interferon analogues -- SRC VB VECTOR	Granulocyte colony-stimulating factor -- Dong-A
Ganirelix -- Roche	Granulocyte colony-stimulating factor conjugate
gastric lipase -- Meristem	grass allergy therapy -- Dynavax
Gavilimomab --	GRF1-44 -- ICN
G-CSF -- Amgen, SRC VB VECTOR	Growth Factor -- Chiron, Atrigel, Atrix, Innogenetics, ZymoGenetics, Novo
GDF-1 -- CeNeS	growth factor peptides -- Biotherapeutics
GDF-5 -- Biopharm	growth hormone -- LG Chem
GDNF (glial derived neurotrophic factor) -- Amgen	growth hormone, Recombinant human -- Serono
gelsolin -- Biogen	GT 4086 -- Gliatech
Gemtuzumab ozogamicin -- Celltech	GW 353430 -- GlaxoSmithKline
Gene-activated epoetin-alfa -- Aventis Pharma -- Transkaryotic Therapies	GW-278884 -- GlaxoSmithKline
Glanzmann thrombasthenia gene therapy --	H 11 -- Viventia Biotech
Glatiramer acetate -- Yeda	H5N1 influenza A virus vaccine -- Protein Sciences
glial growth factor 2 -- CeNeS	haemoglobin -- Biopure
GLP-1 -- Amylin, Suntory, TheraTech, Watson	haemoglobin 3011, Recombinant -- Baxter Healthcare
GLP-1 peptide analogues -- Zealand Pharmaceuticals	haemoglobin crosfumaril -- Baxter Intl.
GLP-2 -- Novo Nordisk, Ontario, Inc., Suntory Limited	haemoglobin stabilized -- Ajinomoto
glucagon -- Eli Lilly, ZymoGenetics	haemoglobin, recombinant -- Apex
Glucagon-like peptide-1 7-36 amide -- Suntory	HAF -- Immune Response
Glucogen-like peptide -- Amylin	Hantavirus vaccine
	HB 19
	HBNF -- Regeneron
	HCC-1 -- Pharis

FIG. 28L

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hCG -- Milkhaus	Herpes simplex glycoprotein DNA vaccine --
hCG vaccine -- Zonagen	Merck, Wyeth-Lederle Vaccines-Malvern,
HE-317 -- Hollis-Eden Pharmaceuticals	Genentech, GlaxoSmithKline, Chiron,
Heat shock protein cancer and influenza	Takeda
vaccines -- StressGen	Herpes simplex vaccine -- Cantab
Helicobacter pylori vaccine -- Acambis,	Pharmaceuticals, CEL-SCI, Henderson
AstraZeneca/CSL, Chiron, Provalis	Morley
Helistat-G -- GalaGen	Herpes simplex vaccine live -- ImClone
Hemolink -- Hemosol	Systems/Wyeth-Lederle, Aventis Pasteur
hepapoietin -- Snow Brand	HGF derivatives -- Dompe
heparanase -- InSight	hIAPP vaccine -- Crucell
heparinase I -- Ibex	Hib-hepatitis B vaccine -- Aventis Pasteur
heparinase III -- Ibex	HIC 1
Hepatitis A vaccine -- American Biogenetic	HIP-- Altachem
Sciences	Hirudins -- Biopharma, Cangene, Dongkook,
Hepatitis A vaccine inactivated	Japan Energy Corporation, Pharmacia
Hepatitis A vaccine Nothav -- Chiron	Corporation, SIR International, Sanofi-
Hepatitis A-hepatitis B vaccine --	Synthelabo, Sotragene, Rhein Biotech
GlaxoSmithKline	HIV edible vaccine -- ProdiGene
hepatitis B therapy -- Tripep	HIV gp120 vaccine -- Chiron, Ajinomoto,
Hepatitis B vaccine -- Amgen, Chiron SpA,	GlaxoSmithKline, ID Vaccine, Progenics,
Meiji Milk, NIS, Prodeva, PowderJect,	VaxGen
Rhein Biotech	HIV gp120 vaccine gene therapy --
Hepatitis B vaccine recombinant -- Evans	HIV gp160 DNA vaccine -- PowderJect,
Vaccines, Epitec Combiotech, Genentech,	Aventis Pasteur, Oncogen, Hyland
MedImmune, Merck Sharp & Dohme,	Immuno, Protein Sciences
Rhein Biotech, Shantha Biotechnics,	HIV gp41 vaccine -- Panacos
Vector, Yeda	HIV HGP-30W vaccine -- CEL-SCI
Hepatitis B vaccine recombinant TGP 943 --	HIV immune globulin -- Abbott, Chiron
Takeda	HIV peptides -- American Home Products
Hepatitis C vaccine -- Bavarian Nordic,	HIV vaccine -- Applied bioTech., Axis
Chiron, Innogenetics Acambis,	Genetics, Biogen, Bristol-Myers Squibb,
Hepatitis D vaccine -- Chiron Vaccines	Genentech, Korea Green Cross, NIS,
Hepatitis E vaccine recombinant --	Oncogen, Protein Sciences Corporation,
Genelabs/GlaxoSmithKline, Novavax	Terumo, Tonen Corporation, Wyeth-
hepatocyte growth factor -- Panorama,	Ayerst, Wyeth-Lederle Vaccines-Malvern,
Sosei	Advanced BioScience Laboratories,
hepatocyte growth factor kringle fragments -	Bavarian Nordic, Bavarian Nordic/Statens
- EntreMed	Serum Institute, GeneCure, Immune
Her-2/Neu peptides -- Corixa	Response, Progenics, Therion Biologics,
	United Biomedical, Chiron

FIG. 28M

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HIV vaccine vCP1433 -- Aventis Pasteur	Human monoclonal antibodies --
HIV vaccine vCP1452 -- Aventis Pasteur	Medarex/Northwest Biotherapeutics,
HIV vaccine vCP205 -- Aventis Pasteur	Medarex/Seattle Genetics
HL-9 -- American BioScience	human netrin-1 -- Exelixis
HM-9239 -- Cytran	human papillomavirus antibodies -- Epicyte
HML-103 -- Hemosol	Human papillomavirus vaccine -- Biotech
HML-104 -- Hemosol	Australia, IDEC, StressGen
HML-105 -- Hemosol	Human papillomavirus vaccine MEDI 501 --
HML-109 -- Hemosol	MedImmune/GlaxoSmithKline
HML-110 -- Hemosol	Human papillomavirus vaccine MEDI
HML-121 -- Hemosol	503/MEDI 504 --
hNLP -- Pharis	MedImmune/GlaxoSmithKline
Hookworm vaccine	Human papillomavirus vaccine TA-CIN --
host-vector vaccines -- Henogen	Cantab Pharmaceuticals
HPM 1 -- Chugai	Human papillomavirus vaccine TA-HPV --
HPV vaccine -- MediGene	Cantab Pharmaceuticals
HSA -- Meristem	Human papillomavirus vaccine TH-GW --
HSF -- StressGen	Cantab/GlaxoSmithKline
HSP carriers --Weizmann, Yeda, Peptor	human polyclonal antibodies -- Biosite/Eos
HSPPC-70 -- Antigenics	BioTech./ Medarex
HSPPC-96, pathogen-derived -- Antigenics	human type II anti factor VIII monoclonal
HSV 863 -- Novartis	antibodies -- ThromboGenics
HTLV-I DNA vaccine	humanised anti glycoprotein Ib murine
HTLV-I vaccine	monoclonal antibodies -- ThromboGenics
HTLV-II vaccine -- Access	HumaRAD -- Intracell
HU 901 -- Tanox	HuMax EGFR -- Genmab
Hu23F2G -- ICOS	HuMax-CD4 -- Medarex
HuHMFG1	HuMax-IL15 -- Genmab
HumaLYM -- Intracell	HYB 190 -- Hybridon
Human krebs statika -- Yamanouchi	HYB 676 -- Hybridon
human monoclonal antibodies --	I-125 MAb A33 -- Celltech
Abgenix/Biogen, Abgenix/ Corixa,	Ibritumomab tiuxetan -- IDEC
Abgenix/Immunex, Abgenix/Lexicon,	IBT-9401 -- Ibex
Abgenix/ Pfizer, Athersys/Medarex,	IBT-9402 -- Ibex
Biogen/MorphoSys, CAT/Searle,	IC 14 -- ICOS
Centocor/Medarex, Corixa/Kirin Brewery,	Idarubicin anti-Ly-2.1 --
Corixa/Medarex, Eos BioTech./Medarex,	IDEC 114 -- IDEC
Eos/Xenerex, Exelixis/Protein Design	IDEC 131 -- IDEC
Labs, ImmunoGen/ Raven, Medarex/	IDEC 152 -- IDEC
B.Twelve, MorphoSys/ImmunoGen, XTL	IDM 1 -- IDM
Biopharmaceuticals/Dyax,	IDPS -- Hollis-Eden Pharmaceuticals

FIG. 28N

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iduronate-2-sulfatase -- Transkaryotic Therapies	insulin -- AutoImmune, Altea, Biobras, BioSante, Bio-Tech. General, Chong Kun Dang, Emisphere, Flamel, Provalis, Rhein Biotech, TranXenoGen
IGF/IBP-2-13 -- Pharis	insulin (bovine) -- Novartis
IGN-101 -- Igeneon	insulin analogue -- Eli Lilly
IK HIR02 -- Iketon	Insulin Aspart -- Novo Nordisk
IL-11 -- Genetics Institute/AHP	insulin detemir -- Novo Nordisk
IL-13-PE38 -- NeoPharm	insulin glargine -- Aventis
IL-17 receptor -- Immunex	insulin inhaled -- Inhale Therapeutics Systems, Alkermes
IL-18BP -- Yeda	insulin oral -- Inovax
IL-1Hy1 -- Hyseq	insulin, AeroDose -- AeroGen
IL-1 β -- Celltech	insulin, AERx -- Aradigm
IL-1 β adjuvant -- Celltech	insulin, BEODAS -- Elan
IL-2 -- Chiron	insulin, Biphax -- Helix
IL-2 + IL-12 -- Hoffman La-Roche	insulin, buccal -- Genex
IL-6/sIL-6R fusion -- Hadasit	insulin, I2R -- Flemington
IL-6R derivative -- Tosoh	insulin, intranasal -- Bentley
IL-7-Dap 389 fusion toxin -- Ligand	insulin, oral -- Nobex, Unigene
IL-21 -- Novo Nordisk, ZymoGenetics	insulin, Orasome -- Endorex
IM-862 -- Cytran	insulin, ProMaxx -- Epic
IMC-1C11 -- ImClone	insulin, Quadrant -- Elan
imiglucerase -- Genzyme	insulin, recombinant -- Aventis
Immune globulin intravenous (human) -- Hoffman La Roche	insulin, Spiros -- Elan
immune privilege factor -- Proneuron	insulin, Transfersome -- IDEA
Immunocal -- Immunotec	insulin, Zymo, recombinant -- Novo Nordisk
Immunogene therapy -- Briana Bio-Tech	insulinotropin -- Scios
Immunoliposomal 5-fluorodeoxyuridine-dipalmitate --	Insulysin gene therapy --
immunosuppressant vaccine -- Aixlie	integrin antagonists -- Merck
immunotoxin -- Antisoma, NIH	interferon (Alpha2) -- SRC VB VECTOR, Viragen, Dong-A, Hoffman La-Roche, Genentech
ImmuRAIT-Re-188 -- Immunomedics	interferon -- BioMedicines, Human Genome Sciences
imreg-1 -- Imreg	interferon (Alfa-n3) -- Interferon Sciences Intl.
infertility -- Johnson & Johnson, E-TRANS	interferon (Alpha), Biphax -- Helix
Infliximab -- Centocor	
Influenza virus vaccine -- Aventis Pasteur, Protein Sciences	
inhibin -- Biotech Australia, Human Therapeutics	
Inhibitory G protein gene therapy	
INKP-2001 -- InKine	
Inolimomab -- Diaclone	

FIG. 280

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interferon (Alpha)—Amgen, BioNative,	IL-2/ diphtheria toxin -- Ligand
Novartis, Genzyme Transgenics,	Interleukin-3 -- Cangene
Hayashibara, Inhale Therapeutics	Interleukin-4 -- Immunology Ventures,
Systems, Medusa, Flamel, Dong-A,	Sanofi Winthrop, Schering-Plough,
GeneTrol, Nastech, Shantha,	Immunex/ Sanofi Winthrop, Bayer, Ono
Wassermann, LG Chem, Sumitomo,	interleukin-4 + TNF-Alpha -- NIH
Aventis, Behring EGIS, Pepgen, Servier,	interleukin-4 agonist -- Bayer
Rhein Biotech,	interleukin-4 fusion toxin -- Ligand
interferon (Alpha2A)	Interleukin-4 receptor -- Immunex, Immun
interferon (Alpha2B) -- Enzon, Schering-	Interleukin-6 -- Ajinomoto, Cangene, Yeda,
Plough, Biogen, IDEA	Genetics Institute, Novartis
interferon (Alpha-N1) -- GlaxoSmithKline	interleukin-6 fusion protein
interferon (beta) -- Rentschler, GeneTrol,	interleukin-6 fusion toxin -- Ligand, Serono
Meristem, Rhein Biotech, Toray, Yeda,	interleukin-7 -- IC Innovations
Daiichi, Mochida	interleukin-7 receptor -- Immunex
interferon (Beta1A) -- Serono, Biogen	interleukin-8 antagonists -- Kyowa
interferon (beta1A),inhale -- Biogen	Hakko/Millennium/Pfizer
interferon (β1b)-- Chiron	interleukin-9 antagonists -- Genaera
interferon (tau)-- Pepgen	Interleukin-10 -- DNAX, Schering-Plough
Interferon alfacon-1 -- Amgen	Interleukin-10 gene therapy --
Interferon alpha-2a vaccine	interleukin-12 -- Genetics Institute, Hoffman
Interferon Beta 1b -- Schering/Chiron,	La-Roche
InterMune	interleukin-13 -- Sanofi
Interferon Gamma -- Boehringer Ingelheim,	interleukin-13 antagonists -- AMRAD
Sheffield, Rentschler, Hayashibara	Interleukin-13-PE38QQR
interferon receptor , Type I -- Serono	interleukin-15 -- Immunex
interferon(Gamma1B) -- Genentech	interleukin-16 -- Research Corp
Interferon-alpha-2b + ribavirin -- Biogen,	interleukin-18 -- GlaxoSmithKline
ICN	Interleukin-18 binding protein -- Serono
Interferon-alpha-2b gene therapy --	Ior-P3 -- Center of Molecular Immunology
Schering-Plough	IP-10 -- NIH
Interferon-con1 gene therapy --	IPF -- Metabolex
interleukin-1 antagonists -- Dompe	IR-501 -- Immune Response
Interleukin-1 receptor antagonist -- Abbott	ISIS 9125 -- Isis Pharmaceuticals
Bioresearch, Pharmacia	ISURF No. 1554 -- Millennium
Interleukin-1 receptor type I -- Immunex	ISURF No. 1866 -- Iowa State Univer.
interleukin-1 receptor Type II -- Immunex	ITF-1697 -- Italfarmaco
Interleukin-1 trap -- Regeneron	IxC 162 -- Ixion
Interleukin-1-alpha -- Immunex/Roche	J 695 -- Cambridge Antibody Tech.,
interleukin-2 -- SRC VB VECTOR,	Genetics Inst., Knoll
Ajinomoto, Biomira, Chiron	Jagged + FGF -- Repair

FIG. 28P

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JKC-362 -- Phoenix Pharmaceuticals	leptin, 2nd-generation -- Amgen
JTP-2942 -- Japan Tobacco	leridistim -- Pharmacia
Juman monoclonal antibodies -- Medarex/Raven	leuprolide, ProMaxx -- Epic
K02 -- Axys Pharmaceuticals	leuprorelin, oral -- Unigene
Keliximab -- IDEC	LeuTech -- Papatin
Keyhole limpet haemocyanin	LEX 032 -- SuperGen
KGF -- Amgen	LiDEPT -- Novartis
KM 871 -- Kyowa	Lintuzumab (anti-CD33 MAb) -- Protein Design Labs
KPI 135 -- Scios	lipase -- Altus Biologics
KPI-022 -- Scios	lipid A vaccine -- EntreMed
Kringle 5	lipid-linked anchor Tech. -- ICRT, ID Biomedical
KSB 304	liposome-CD4 Tech. -- Sheffield
KSB-201 -- KS Biomedix	Listeria monocytogenes vaccine
L 696418 -- Merck	LMB 1
L 703801 -- Merck	LMB 7
L1 -- Acorda	LMB 9 -- Battelle Memorial Institute, NIH
L-761191 -- Merck	LM-CD45 -- Cantab Pharmaceuticals
lactoferrin -- Meristem, Pharming, Agennix	lovastatin -- Merck
lactoferrin cardio -- Pharming	LSA-3
LAG-3 -- Serono	LT- β receptor -- Biogen
LAIT -- GEMMA	lung cancer vaccine -- Corixa
LAK cell cytotoxin -- Arizona	lusupultide -- Scios
lamellarins -- PharmaMar/University of Malaga	L-Vax -- AVAX
laminin A peptides -- NIH	LY 355455 -- Eli Lilly
lanotepase -- Genetics Institute	LY 366405 -- Eli Lilly
laronidase -- BioMarin	LY-355101 -- Eli Lilly
Lassa fever vaccine	Lyme disease DNA vaccine -- Vical/Aventis Pasteur
LCAT -- NIH	Lyme disease vaccine -- Aquila
LDP 01 -- Millennium	Biopharmaceuticals, Aventis, Pasteur, Symbicom, GlaxoSmithKline, Hyland
LDP 02 -- Millennium	Immuno, MedImmune
Lecithinized superoxide dismutase -- Seikagaku	Lymphocytic choriomeningitis virus vaccine
LeIF adjuvant -- Corixa	lymphoma vaccine -- Biomira, Genitope
leishmaniasis vaccine -- Corixa	LYP18
lenercept -- Hoffman La-Roche	lys plasminogen, recombinant
Lenograstim -- Aventis, Chugai	Lysosomal storage disease gene therapy -- Avigen
lepirudin -- Aventis	lysostaphin -- Nutrition 21
leptin -- Amgen, IC Innovations	
Leptin gene therapy -- Chiron Corporation	

FIG. 28Q

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M 23 -- Gruenenthal	MEDI 507 -- BioTransplant
M1 monoclonal antibodies -- Acorda	melanin concentrating hormone --
Therapeutics	Neurocrine Biosciences
MA 16N7C2 -- Corvas Intl.	melanocortins -- OMRF
malaria vaccine -- GlaxoSmithKline,	Melanoma monoclonal antibodies -- Viragen
AdProTech, Antigenics, Apovia, Aventis	melanoma vaccine -- GlaxoSmithKline,
Pasteur, Axis Genetics, Behringwerke,	Akzo Nobel, Avant, Aventis Pasteur,
CDCP, Chiron Vaccines, Genzyme	Bavarian Nordic, Biovector, CancerVax,
Transgenics, Hawaii, MedImmune, NIH,	Genzyme Molecular Oncology, Humbolt,
NYU, Oxxon, Roche/Saramane, Biotech	ImClone Systems, Memorial, NYU, Oxxon
Australia, Rx Tech	Melanoma vaccine Magevac -- Therion
Malaria vaccine CDC/NIIMALVAC-1	memory enhancers -- Scios
malaria vaccine, multicomponent	meningococcal B vaccine -- Chiron
mammaglobin -- Corixa	meningococcal vaccine -- CAMR
mammastatin -- Biotherapeutics	Meningococcal vaccine group B conjugate -
mannan-binding lectin -- NatlImmu	- North American Vaccine
mannan-MUC1 -- Psiron	Meningococcal vaccine group B
MAP 30	recombinant -- BioChem Vaccines,
Marinovir -- Phytera	Microscience
MARstem -- Maret	Meningococcal vaccine group Y conjugate -
MB-015 -- Mochida	- North American Vaccine
MBP -- ImmuLogic	Meningococcal vaccine groups A B and C
MCI-028 -- Mitsubishi-Tokyo	conjugate -- North American Vaccine
MCIF -- Human Genome Sciences	Mepolizumab -- GlaxoSmithKline
MDC -- Advanced BioScience -- Akzo	Metastatin -- EntreMed, Takeda
Nobel, ICOS	Met-CkB7 -- Human Genome Sciences
MDX 11 -- Medarex	met-enkephalin -- TNI
MDX 210 -- Medarex	METH-1 -- Human Genome Sciences
MDX 22 -- Medarex	methioninase -- AntiCancer
MDX 22	Methionine lyase gene therapy --
MDX 240 -- Medarex	AntiCancer
MDX 33	Met-RANTES -- Genexa Biomedical,
MDX 44 -- Medarex	Serono
MDX 447 -- Medarex	Metreleptin
MDX H210 -- Medarex	Microtubule inhibitor MAb
MDX RA -- Houston BioTech., Medarex	Immunogen/Abgenix
ME-104 -- Pharmexa	MGDF -- Kirin
Measles vaccine	MGV -- Progenics
Mecasermin -- Cephalon/Chiron, Chiron	micrin -- Endocrine
MEDI 488 -- MedImmune	microplasmin -- ThromboGenics
MEDI 500	MIF -- Genetics Institute

FIG. 28R

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migration inhibitory factor -- NIH	MAB 45-2D9- -- haematoporphyrin conjugate
Mim CD4.1 -- Xyte Therapies	MAB 4B4
mirostipen -- Human Genome Sciences	MAB 4E3-CPA conjugate -- BCM Oncologia
Mitumomab (BEC-2) -- ImClone Systems, Merck KGaA	MAB 4E3-daunorubicin conjugate
MK 852 -- Merck	MAB 50-6
MLN 1202 (Anti-CCR2 monoclonal antibody) -- Millenium Pharmaceuticals	MAB 50-61A -- Institut Pasteur
Mobenakin -- NIS	MAB 5A8 -- Biogen
molgramostim -- Genetics Institute, Novartis	MAB 791T/36-methotrexate conjugate
monoclonal antibodies -- Abgenix/Celltech, Immusol/ Medarex, Viragen/ Roslin Institute, Cambridge Antibody Tech./Elan	MAB 7c11.e8
MAB 108 --	MAB 7E11 C5-selenocystamine conjugate
MAB 10D5 --	MAB 93KA9 -- Novartis
MAB 14.18-interleukin-2 immunocytokine -- Lexigen	MAB A5B7-cisplatin conjugate -- Biodynamics Research, Pharmacia
MAB 14G2a --	MAB A5B7-I-131
MAB 15A10 --	MAB A7
MAB 170 -- Biomira	MAB A717 -- Exocell
MAB 177Lu CC49 --	MAB A7-zinostatin conjugate
MAB 17F9	MAB ABX-RB2 -- Abgenix
MAB 1D7	MAB ACA 11
MAB 1F7 -- Immune Network	MAB AFP-I-131 -- Immunomedics
MAB 1H10-doxorubicin conjugate	MAB AP1
MAB 26-2F	MAB AZ1
MAB 2A11	MAB B3-LysPE40 conjugate
MAB 2E1 -- RW Johnson	MAB B4 -- United Biomedical
MAB 2F5	MAB B43 Genistein-conjugate
MAB 31.1 -- International BioImmune Systems	MAB B43.13-Tc-99m -- Biomira
MAB 32 -- Cambridge Antibody Tech., Peptech	MAB B43-PAP conjugate
MAB 323A3 -- Centocor	MAB B4G7-gelonin conjugate
MAB 3C5	MAB BCM 43-daunorubicin conjugate -- BCM Oncologia
MAB 3F12	MAB BIS-1
MAB 3F8	MAB BMS 181170 -- Bristol-Myers Squibb
MAB 42/6	MAB BR55-2
MAB 425 -- Merck KGaA	MAB BW494
MAB 447-52D -- Merck Sharp & Dohme	MAB C 242-DM1 conjugate -- ImmunoGen
	MAB C242-PE conjugate
	MAB c30-6
	MAB CA208-cytorhodin-S conjugate -- Hoechst Japan
	MAB CC49 -- Enzon

FIG. 28S

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MAB ch14.18 –	MAB LL2-I-131 – Immunomedics
MAB CH14.18-GM-CSF fusion protein -- Lexigen	MAB LL2-Y-90
MAB chCE7	MAB LS2D617 -- Hybritech
MAB CI-137 -- AMRAD	MAB LYM-1-gelonin conjugate
MAB cisplatin conjugate	MAB LYM-1-I-131
MAB CLB-CD19	MAB LYM-1-Y-90
MAB CLB-CD19v	MAB LYM-2 -- Peregrine
MAB CLL-1 -- Peregrine	MAB M195
MAB CLL-1-GM-CSF conjugate	MAB M195-bismuth 213 conjugate -- Protein Design Labs
MAB CLL-1-IL-2 conjugate -- Peregrine	MAB M195-gelonin conjugate
MAB CLN IgG -- doxorubicin conjugates	MAB M195-I-131
MAB conjugates – Tanox	MAB M195-Y-90
MAB D612	MAB MA 33H1 -- Sanofi
MAB Dal B02	MAB MAD11
MAB DC101 -- ImClone	MAB MGb2
MAB EA 1 –	MAB MINT5
MAB EC708 -- Biovation	MAB MK2-23
MAB EP-5C7 -- Protein Design Labs	MAB MOC31 ETA(252-613) conjugate
MAB ERIC-1 -- ICRT	MAB MOC-31-In-111
MAB F105 gene therapy	MAB MOC-31-PE conjugate
MAB FC 2.15	MAB MR6 –
MAB G250 -- Centocor	MAB MRK-16 -- Aventis Pasteur
MAB GA6	MAB MS11G6
MAB GA733	MAB MX-DTPA BrE-3
MAB Gliomab-H -- Viventia Biotech	MAB MY9
MAB HB2-saporin conjugate	MAB Nd2 -- Tosoh
MAB HD 37 –	MAB NG-1 -- Hygeia
MAB HD37-ricin chain-A conjugate	MAB NM01 – Nissin Food
MAB HNK20 -- Acambis	MAB OC 125
MAB huN901-DM1 conjugate -- ImmunoGen	MAB OC 125-CMA conjugate
MAB I-131 CC49 -- Corixa	MAB OKI-1 -- Ortho-McNeil
MAB ICO25	MAB OX52 -- Bioproducts for Science
MAB ICR12-CPG2 conjugate	MAB PMA5
MAB ICR-62	MAB PR1
MAB IRac-ricin A conjugate	MAB prost 30
MAB K1	MAB R-24
MAB KS1-4-methotrexate conjugate	MAB R-24 α Human GD3 -- Celltech
MAB L6 -- Bristol-Myers Squibb, Oncogen	MAB RFB4-ricin chain A conjugate
MAB LiCO 16-88	MAB RFT5-ricin chain A conjugate
	MAB SC 1

FIG. 28T

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MAb SM-3 -- ICRT	Muc-1 vaccine -- Corixa
MAb SMART 1D10 -- Protein Design Labs	mucosal tolerance -- Aberdeen
MAb SMART ABL 364 -- Novartis	mullerian inhibiting subst
MAb SN6f	muplestim -- Genetics Institute, Novartis,
MAb SN6f-deglycosylated ricin A chain	DSM Anti-Infectives
conjugate --	murine MAb -- KS Biomedix
MAb SN6j	Mutant somatropin -- JCR Pharmaceutical
MAb SN7-ricin chain A conjugate	MV 833 -- Toagosei
MAb T101-Y-90 conjugate -- Hybritech	Mycoplasma pulmonis vaccine
MAb T-88 -- Chiron	Mycoprex -- XOMA
MAb TB94 -- Cancer ImmunoBiology	myeloperoxidase -- Henogen
MAb TEC 11	myostatin -- Genetics Institute
MAb TES-23 -- Chugai	Nacolomab tafenatox -- Pharmacia
MAb TM31 -- Avant	Nagrecor -- Scios
MAb TNT-1 -- Cambridge Antibody Tech.,	nagrestipen -- British Biotech
Peregrine	NAP-5 -- Corvas Intl.
MAb TNT-3	NAPc2 -- Corvas Intl.
MAb TNT-3 -- IL2 fusion protein --	nartograstim -- Kyowa
MAb TP3-A α -211	Natalizumab -- Protein Design Labs
MAb TP3-PAP conjugate --	Nateplase -- NIH, Nihon Schering
MAb UJ13A -- ICRT	nateplase -- Schering AG
MAb UN3	NBI-3001 -- Neurocrine Biosci.
MAb ZME-018-gelonin conjugate	NBI-5788 -- Neurocrine Biosci.
MAb-BC2 -- GlaxoSmithKline	NBI-6024 -- Neurocrine Biosci.
MAb-DM1 conjugate -- ImmunoGen	Nef inhibitors -- BRI
MAb-ricin-chain-A conjugate -- XOMA	Neisseria gonorrhoea vaccine -- Antex
MAb-temoporfin conjugates	Biologics
Monopharm C -- Viventia Biotech	Neomycin B-arginine conjugate
monteplase -- Eisai	Nerelimomab -- Chiron
montirelin hydrate -- Gruenenthal	Nerve growth factor -- Amgen -- Chiron,
moroctocog alfa -- Genetics Institute	Genentech
Moroctocog-alfa -- Pharmacia	Nerve growth factor gene therapy
MP 4	nesiritide citrate -- Scios
MP-121 -- Biopharm	neuregulin-2 -- CeNeS
MP-52 -- Biopharm	neurocan -- NYU
MRA -- Chugai	neuronal delivery system -- CAMR
MS 28168 -- Mitsui Chemicals, Nihon	Neurophil inhibitory Factor -- Corvas
Schering	Neuroprotective vaccine -- University of
MSH fusion toxin -- Ligand	Auckland
MSI-99 -- Genaera	neurotrophic chimaeras -- Regeneron
MT 201 -- Micromet	neurotrophic factor -- NsGene, CereMedix

FIG. 28U

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NeuroVax -- Immune Response	Oncophage -- Antigenics
neurturin -- Genentech	Oncostatin M -- Bristol-Myers Squibb
neutral endopeptidase -- Genentech	OncoVax-CL -- Jenner Biotherapies
NGF enhancers -- NeuroSearch	OncoVax-P -- Jenner Biotherapies
NHL vaccine -- Large Scale Biology	onercept -- Yeda
NIP45 -- Boston Life Sciences	onychomycosis vaccine -- Boehringer
NKI-B20	Ingelheim
NM 01 -- Nissin Food	opebecan -- XOMA
NMI-139 -- NitroMed	opioids -- Arizona
NMMP -- Genetics Institute	Oprelvekin -- Genetics Institute
NN-2211 -- Novo Nordisk	Oregovomab -- AltaRex
Noggin -- Regeneron	Org-33408 b-- Akzo Nobel
Nonacog alfa	Orolip DP -- EpiCept
Norelin -- Biostar	oryzacystatin
Norwalk virus vaccine	OSA peptides -- GenSci Regeneration
NRLU 10 -- NeoRx	osteoblast-cadherin GF -- Pharis
NRLU 10 PE -- NeoRx	Osteocalcin-thymidine kinase gene therapy
NT-3 -- Regeneron	osteogenic protein -- Curis
NT-4/5 -- Genentech	osteopontin -- OraPharma
NU 3056	osteoporosis peptides -- Integra, Telios
NU 3076	osteoprotegerin -- Amgen, SnowBrand
NX 1838 -- Gilead Sciences	otitis media vaccines -- Antex Biologics
NY ESO-1/CAG-3 antigen -- NIH	ovarian cancer -- University of Alabama
NYVAC-7 -- Aventis Pasteur	OX40-IgG fusion protein -- Cantab, Xenova
NZ-1002 -- Novazyme	P 246 -- Diatide
obesity therapy -- Nobex	P 30 -- Alfacell
OC 10426 -- Ontogen	p1025 -- Active Biotech
OC 144093 -- Ontogen	P-113 [^] -- Demegen
OCIF -- Sankyo	P-16 peptide -- Transition Therapeutics
Oct-43 -- Otsuka	p43 -- Ramot
Odulimomab -- Immunotech	P-50 peptide -- Transition Therapeutics
OK PSA - liposomal	p53 + RAS vaccine -- NIH, NCI
OKT3-gamma-1-ala-ala	PACAP(1-27) analogue
OM 991	paediatric vaccines -- Chiron
OM 992	Pafase -- ICOS
Omalizumab -- Genentech	PAGE-4 plasmid DNA -- IDEC
oncoimmunin-L -- NIH	PAI-2 -- Biotech Australia, Human
Oncolysin B -- ImmunoGen	Therapeutics
Oncolysin CD6 -- ImmunoGen	Palifermin (keratinocyte growth factor) --
Oncolysin M -- ImmunoGen	Amgen
Oncolysin S -- ImmunoGen	Palivizumab -- MedImmune

FIG. 28V

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PAM 4 -- Merck	PEG-uricase -- Mountain View
pamiteplase -- Yamanouchi	Pegvisomant -- Genentech
pancreatin, Minitabs -- Eurand	PEGylated proteins, PolyMASC -- Valentis
Pangen -- Fournier	PEGylated recombinant native human leptin
Pantarin -- Selective Genetics	-- Roche
Parainfluenza virus vaccine -- Pharmacia,	Pemtumomab
Pierre Fabre	Penetratin -- Cyclacel
paraoxanase -- Esperion	Pepscan -- Antisoma
parathyroid hormone -- Abiogen, Korea	peptide G -- Peptech, ICRT
Green Cross	peptide vaccine -- NIH, NCI
Parathyroid hormone (1-34) --	Pexelizumab
Chugai/Suntory	pexiganan acetate -- Genaera
Parkinson's disease gene therapy -- Cell	Pharmaprojects No. 3179 -- NYU
Genesys/ Ceregene	Pharmaprojects No. 3390 -- Ernest Orlando
Parvovirus vaccine -- MedImmune	Pharmaprojects No. 3417 -- Sumitomo
PCP-Scan -- Immunomedics	Pharmaprojects No. 3777 -- Acambis
PDGF -- Chiron	Pharmaprojects No. 4209 -- XOMA
PDGF cocktail -- Theratechnologies	Pharmaprojects No. 4349 -- Baxter Intl.
peanut allergy therapy -- Dynavax	Pharmaprojects No. 4651
PEG anti-ICAM MAb -- Boehringer	Pharmaprojects No. 4915 -- Avanir
Ingelheim	Pharmaprojects No. 5156 -- Rhizogenics
PEG asparaginase -- Enzon	Pharmaprojects No. 5200 -- Pfizer
PEG glucocerebrosidase	Pharmaprojects No. 5215 -- Origene
PEG hirudin -- Knoll	Pharmaprojects No. 5216 -- Origene
PEG interferon-alpha-2a -- Roche	Pharmaprojects No. 5218 -- Origene
PEG interferon-alpha-2b + ribavirin --	Pharmaprojects No. 5267 -- ML
Biogen, Enzon, ICN Pharmaceuticals,	Laboratories
Schering-Plough	Pharmaprojects No. 5373 -- MorphoSys
PEG MAb A5B7 --	Pharmaprojects No. 5493 -- Metabolex
Pegacaristim -- Amgen -- Kirin Brewery --	Pharmaprojects No. 5707 -- Genentech
ZymoGenetics	Pharmaprojects No. 5728 -- Autogen
Pegaldesleukin -- Research Corp	Pharmaprojects No. 5733 -- BioMarin
pegaspargase -- Enzon	Pharmaprojects No. 5757 -- NIH
pegfilgrastim -- Amgen	Pharmaprojects No. 5765 -- Gryphon
PEG-interferon Alpha -- Viragen	Pharmaprojects No. 5830 -- AntiCancer
PEG-interferon Alpha 2A -- Hoffman La-	Pharmaprojects No. 5839 -- Dyax
Roche	Pharmaprojects No. 5849 -- Johnson &
PEG-interferon Alpha 2B -- Schering-	Johnson
Plough	Pharmaprojects No. 5860 -- Mitsubishi-
PEG-r-hirudin -- Abbott	Tokyo
PEG-rHuMGDF -- Amgen	

FIG. 28W

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Pharmaprojects No. 5869 -- Oxford GlycoSciences	Plasminogen activators -- Abbott Laboratories, American Home Products, Boehringer Mannheim, Chiron
Pharmaprojects No. 5883 -- Asahi Brewery	Corporation, DuPont Pharmaceuticals, Eli Lilly, Shionogi, Genentech, Genetics Institute, GlaxoSmithKline, Hemispherx Biopharma, Merck & Co, Novartis, Pharmacia Corporation, Wakamoto, Yeda
Pharmaprojects No. 5947 -- StressGen	plasminogen-related peptides -- Bio-Tech. General/MGH
Pharmaprojects No. 5961 -- Theratechnologies	platelet factor 4 -- RepliGen
Pharmaprojects No. 5962 -- NIH	Platelet-derived growth factor -- Amgen -- ZymoGenetics
Pharmaprojects No. 5966 -- NIH	plusonemin-- Hayashibara
Pharmaprojects No. 5994 -- Pharming	PMD-2850 -- Protherics
Pharmaprojects No. 5995 -- Pharming	Pneumococcal vaccine -- Antex Biologics, Aventis Pasteur
Pharmaprojects No. 6023 -- IMMUCON	Pneumococcal vaccine intranasal -- BioChem Vaccines/Biovector
Pharmaprojects No. 6063 -- Cytoclonal	PR1A3
Pharmaprojects No. 6073 -- SIDDCO	PR-39
Pharmaprojects No. 6115 -- Genzyme	pralmorelin -- Kaken
Pharmaprojects No. 6227 -- NIH	Pretarget-Lymphoma -- NeoRx
Pharmaprojects No. 6230 -- NIH	Priliximab -- Centocor
Pharmaprojects No. 6236 -- NIH	PRO 140 -- Progenics
Pharmaprojects No. 6243 -- NIH	PRO 2000 -- Procept
Pharmaprojects No. 6244 -- NIH	PRO 367 -- Progenics
Pharmaprojects No. 6281 -- Senetek	PRO 542 -- Progenics
Pharmaprojects No. 6365 -- NIH	pro-Apo A-I -- Esperion
Pharmaprojects No. 6368 -- NIH	prolactin -- Genzyme
Pharmaprojects No. 6373 -- NIH	Prosaptide TX14(A) -- Bio-Tech. General
Pharmaprojects No. 6408 -- Pan Pacific	prostate cancer antibodies -- Immunex, UroCor
Pharmaprojects No. 6410 -- Athersys	prostate cancer antibody therapy -- Genentech/UroGenesys, Genotherapeutics
Pharmaprojects No. 6421 -- Oxford GlycoSciences	prostate cancer immunotherapeutics -- The PSMA Development Company
Pharmaprojects No. 6522 -- Maxygen	prostate cancer vaccine -- Aventis Pasteur, Zonagen, Corixa, Dendreon, Jenner
Pharmaprojects No. 6523 -- Pharis	Biotherapies, Therion Biologics
Pharmaprojects No. 6538 -- Maxygen	
Pharmaprojects No. 6554 -- APALEXO	
Pharmaprojects No. 6560 -- Ardana	
Pharmaprojects No. 6562 -- Bayer	
Pharmaprojects No. 6569 -- Eos	
Phenoxazine	
Phenylase -- Ibex	
Pigment epithelium derived factor -- plasminogen activator inhibitor-1, recombinant -- DuPont Pharmaceuticals	

FIG. 28X

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prostate-specific antigen -- EntreMed	RD 62198
protein A -- RepliGen	rDnase -- Genentech
protein adhesives -- Enzon	RDP-58 -- SangStat
protein C -- Baxter Intl., PPL Therapeutics,	RecepTox-Fce -- Keryx
ZymoGenetics	RecepTox-GnRH -- Keryx, MTR
protein C activator -- Gilead Sciences	Technologies
protein kinase R antags -- NIH	RecepTox-MBP -- Keryx, MTR
protirelin -- Takeda	Technologies
protocadherin 2 -- Caprion	recFSH -- Akzo Nobel, Organon
Pro-urokinase -- Abbott, Bristol-Myers	REGA 3G12
Squibb, Dainippon, Tosoh -- Welfide	Regavirumab -- Teijin
P-selectin glycoprotein ligand-1 -- Genetics	relaxin -- Connetics Corp
Institute	Renal cancer vaccine -- Macropharm
pseudomonal infections -- InterMune	repifermin -- Human Genome Sciences
Pseudomonas vaccine -- Cytovax	Respiratory syncytial virus PFP-2 vaccine --
PSGL-Ig -- American Home Products	Wyeth-Lederle
PSP-94 -- Procyon	Respiratory syncytial virus vaccine --
PTH 1-34 -- Nobex	GlaxoSmithKline, Pharmacia, Pierre Fabre
Quilimmune-M -- Antigenics	Respiratory syncytial virus vaccine
R 744 -- Roche	inactivated
R 101933	Respiratory syncytial virus-parainfluenza
R 125224 -- Sankyo	virus vaccine -- Aventis Pasteur,
RA therapy -- Cardion	Pharmacia
Rabies vaccine recombinant -- Aventis	Retepase -- Boehringer Mannheim,
Pasteur, BioChem Vaccines, Kaketsuken	Hoffman La-Roche
Pharmaceuticals	Retropep -- Retroscreen
RadioTheraCIM -- YM BioSciences	RFB4 (dsFv) PE38
Ramot project No. 1315 -- Ramot	RFI 641 -- American Home Products
Ramot project No. K-734A -- Ramot	RFTS -- UAB Research Foundation
Ramot project No. K-734B -- Ramot	RG 12986 -- Aventis Pasteur
Ranibizumab (Anti-VEGF fragment) --	RG 83852 -- Aventis Pasteur
Genentech	RG-1059 -- RepliGen
RANK -- Immunex	rGCR -- NIH
ranpirnase -- Alfacell	rGLP-1 -- Restoragen
ranpirnase-anti-CD22 MAb -- Alfacell	rGRF -- Restoragen
RANTES inhibitor -- Milan	rh Insulin -- Eli Lilly
RAPID drug delivery systems -- ARIAD	RHAMM targeting peptides -- Cangene
rasburicase -- Sanofi	rHb1.1 -- Baxter Intl.
rBPI-21, topical -- XOMA	rhCC10 -- Claragen
RC 529 -- Corixa	rhCG -- Serono
rCFTR -- Genzyme Transgenics	Rheumatoid arthritis gene therapy

FIG. 28Y

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Rheumatoid arthritis vaccine -- Veterans Affairs Medical Center	SB RA 31012 --
rhLH -- Serono	SC 56929 -- Pharmacia
Ribozyme gene therapy -- Genset	SCA binding proteins -- Curis, Enzon
Rickettsial vaccine recombinant	scFv(14E1)-ETA Berlex Laboratories, Schering AG
RIGScan CR -- Neoprobe	ScFv(FRP5)-ETA --
RIP-3 -- Rigel	ScFv6C6-PE40 --
Rituximab -- Genentech	SCH 55700 -- Celltech
RK-0202 -- RxKinetix	Schistosomiasis vaccine -- Glaxo Wellcome/Medeva, Brazil
RLT peptide -- Esperion	SCPF -- Advanced Tissue Sciences
rM/NEI -- IVAX	scuPA-suPAR complex -- Hadasit
rmCRP -- Immtech	SD-9427 -- Pharmacia
RN-1001 -- Renovo	SDF-1 -- Ono
RN-3 -- Renovo	SDZ 215918 -- Novartis
RNAse conjugate -- Immunomedics	SDZ 280125 -- Novartis
RO 631908 -- Roche	SDZ 89104 -- Novartis
Rotavirus vaccine -- Merck	SDZ ABL 364 -- Novartis
RP 431 -- DuPont Pharmaceuticals	SDZ MMA 383 -- Novartis
RP-128 -- Resolution	Secretin -- Ferring, Repligen
RPE65 gene therapy --	serine protease inhbs -- Pharis
RPR 110173 -- Aventis Pasteur	sermorelin acetate -- Serono
RPR 115135 -- Aventis Pasteur	SERP-1 -- Viron
RPR 116258A -- Aventis Pasteur	sertenef -- Dainippon
rPSGL-Ig -- American Home Products	serum albumin, Recombinant human -- Aventis Behring
r-SPC surfactant -- Byk Gulden	serum-derived factor -- Hadasit
RSV antibody -- Medimmune	Sevirumab -- Novartis
Ruplizumab -- Biogen	SGN 14 -- Seattle Genetics
rV-HER-2/neu -- Therion Biologics	SGN 15 -- Seattle Genetics
SA 1042 -- Sankyo	SGN 17/19 -- Seattle Genetics
sacrosidase -- Orphan Medical	SGN 30 -- Seattle Genetics
Sant 7	SGN-10 -- Seattle Genetics
Sargramostim -- Immunex	SGN-11 -- Seattle Genetics
saruplase -- Gruenenthal	SH 306 -- DuPont Pharmaceuticals
Satumomab -- Cytogen	Shanvac-B -- Shantha
SB 1 -- COR Therapeutics	Shigella flexneri vaccine -- Avant, Acambis, Novavax
SB 207448 -- GlaxoSmithKline	Shigella sonnei vaccine --
SB 208651 -- GlaxoSmithKline	sICAM-1 -- Boehringer Ingelheim
SB 240683 -- GlaxoSmithKline	Silteplase -- Genzyme
SB 249415 -- GlaxoSmithKline	
SB 249417 -- GlaxoSmithKline	
SB 6 -- COR Therapeutics	

FIG. 28Z

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SIV vaccine -- Endocon, Institut Pasteur	Staphylococcus aureus vaccine conjugate --
SK 896 -- Sanwa Kagaku Kenkyusho	Nabi
SK-827 -- Sanwa Kagaku Kenkyusho	Staphylococcus therapy -- Tripep
Skeletex -- CellFactors	Staphylokinase -- Biovation, Prothera,
SKF 106160 -- GlaxoSmithKline	Thrombogenetics
S-nitroso-AR545C --	Streptococcal A vaccine -- M6
SNTP -- Active Biotech	Pharmaceuticals, North American Vaccine
somatomedin-1 -- GroPep, Mitsubishi-	Streptococcal B vaccine -- Microscience
Tokyo, NIH	Streptococcal B vaccine recombinant --
somatomedin-1 carrier protein -- Insmed	Biochem Vaccines
somatostatin -- Ferring	Streptococcus pyogenes vaccine
Somatotropin/	STRL-33 -- NIH
Human Growth Hormone -- Bio-Tech.	Subalin -- SRC VB VECTOR
General, Eli Lilly	SUIS -- United Biomedical
somatropin -- Bio-Tech. General, Alkermes,	SUIS-LHRH -- United Biomedical
ProLease, Aventis Behring, Biovector,	SUN-E3001 -- Suntory
Cangene, Dong-A, Eli Lilly, Emisphere,	super high affinity monoclonal antibodies --
Enact, Genentech, Genzyme Transgenics,	YM BioSciences
Grandis/InfiMed, CSL, InfiMed, MacroMed,	Superoxide dismutase -- Chiron, Enzon,
Novartis, Novo Nordisk, Pharmacia	Ube Industries, Bio-Tech, Yeda
Serono, TranXenoGen	superoxide dismutase-2 -- OXIS
somatropin derivative -- Schering AG	suppressin -- UAB Research Foundation
somatropin, AIR -- Eli Lilly	SY-161-P5 -- ThromboGenics
Somatropin, inhaled -- Eli Lilly/Alkermes	SY-162 -- ThromboGenics
somatropin, Kabi -- Pharmacia	Systemic lupus erythematosus vaccine --
somatropin, Orasome -- Novo Nordisk	MedClone/VivoRx
Sonermin -- Dainippon Pharmaceutical	T cell receptor peptides -- Xoma
SP(V5.2)C -- Supertek	T cell receptor peptide vaccine
SPf66	T4N5 liposomes -- AGI Dermatics
sphingomyelinase -- Genzyme	TACI, soluble -- ZymoGenetics
SR 29001 -- Sanofi	targeted apoptosis -- Antisoma
SR 41476 -- Sanofi	tasonermin -- Boehringer Ingelheim
SR-29001 -- Sanofi	TASP
SS1(dsFV)-PE38 -- NeoPharm	TASP-V
β 2 microglobulin -- Avidex	Tat peptide analogues -- NIH
β 2-microglobulin fusion proteins -- NIH	TBP I -- Yeda
β -amyloid peptides -- CeNeS	TBP II
β -defensin -- Pharis	TBV25H -- NIH
Staphylococcus aureus infections --	Tc 99m ior cea1 -- Center of Molecular
Inhibitex/ZLB	Immunology
	Tc 99m P 748 -- Diatide

FIG. 28AA

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Tc 99m votumumab -- Intracell	Tissue factor -- Genentech
Tc-99m rh-Annexin V -- Theseus Imaging	Tissue factor pathway inhibitor
teceleukin -- Biogen	TJN-135 -- Tsumura
tenecteplase -- Genentech	TM 27 -- Avant
Teriparatide -- Armour Pharmaceuticals, Asahi Kasei, Eli Lilly	TM 29 -- Avant
terlipressin -- Ferring	TMC-151 -- Tanabe Seiyaku
testisin -- AMRAD	TNF tumour necrosis factor -- Asahi Kasei
Tetrafibricin -- Roche	TNF Alpha -- Cytimmune
TFPI -- EntreMed	TNF antibody -- Johnson & Johnson
tgD-IL-2 -- Takeda	TNF binding protein -- Amgen
TGF-Alpha -- ZymoGenetics	TNF degradation product -- Oncotech
TGF- β -- Kolon	TNF receptor -- Immunex
TGF- β 2 -- Insmad	TNF receptor 1, soluble -- Amgen
TGF- β 3 -- OSI	TNF Tumour necrosis factor-alpha -- Asahi Kasei, Genetech, Mochida
Thalassaemia gene therapy -- Crucell	TNF-Alpha inhibitor -- Tripep
TheraCIM-h-R3 -- Center of Molecular Immunology, YM BioSciences	TNFR:Fc gene therapy -- Targeted Genetics
Theradigm-HBV -- Epimmune	TNF-SAM2
Theradigm-HPV -- Epimmune	Tolerimab -- Innogenetics
Theradigm-malaria -- Epimmune	Toxoplasma gondii vaccine -- GlaxoSmithKline
Theradigm-melanoma -- Epimmune	TP 9201 -- Telios
TheraFab -- Antisoma	TP10 -- Avant
ThGRF 1-29 -- Theratechnologies	TP20 -- Avant
ThGRF 1-44 -- Theratechnologies	tPA -- Centocor
Thrombin receptor activating peptide -- Abbott	trafermin -- Scios
thrombomodulin -- Iowa, Novocastra	TRAIL/Apo2L -- Immunex
Thrombopoietin -- Dragon Pharmaceuticals, Genentech	TRAIL-R1 MAb -- Cambridge Antibody Technologies
thrombopoietin, Pliva -- Recepton	transferrin-binding proteins -- CAMR
Thrombospondin 2 --	Transforming growth factor-beta-1 -- Genentech
thrombostatin -- Thromgen	transport protein -- Genesis
thymalfasin -- SciClone	Trastuzumab -- Genetech
thymocartin -- Gedeon Richter	TRH -- Ferring
thymosin Alpha1 -- NIH	Triabin -- Schering AG
thyroid stimulating hormone -- Genzyme	Triconal
tICAM-1 -- Bayer	Triflavin
Tick anticoagulant peptide -- Merck	troponin I -- Boston Life Sciences
TIF -- Xoma	TRP-2 ^A -- NIH
Tifacogin -- Chiron, NIS, Pharmacia	trypsin inhibitor -- Mochida

FIG. 28BB

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TSP-1 gene therapy --	Vascular endothelial growth factors -- R&D
TT-232	Systems
TTS-CD2 -- Active Biotech	vascular targeting agents -- Peregrine
Tuberculosis vaccine -- Aventis Pasteur,	vasopermeation enhancement agents --
Genesis	Peregrine
Tumor Targeted Superantigens -- Active	vasostatin -- NIH
Biotech -- Pharmacia	VCL -- Bio-Tech. General
tumour vaccines -- PhotoCure	VEGF -- Genentech, Scios
tumour-activated prodrug antibody	VEGF inhibitor -- Chugai
conjugates -- Millennium/ImmunoGen	VEGF-2 -- Human Genome Sciences
tumstatin -- ILEX	VEGF-Trap -- Regeneron
Tuvirumab -- Novartis	viscumin, recombinant -- Madaus
TV-4710 -- Teva	Vitaxin
TWEAK receptor -- Immunex	Vitrage -- ISTA Pharmaceuticals
TXU-PAP	West Nile virus vaccine -- Bavarian Nordic
TY-10721 -- TOA Eiyo	WP 652
Type I diabetes vaccine -- Research Corp	WT1 vaccine -- Corixa
Typhoid vaccine CVD 908	WX-293 -- Willex BioTech.
U 143677 -- Pharmacia	WX-360 -- Willex BioTech.
U 81749 -- Pharmacia	WX-UK1 -- Willex BioTech.
UA 1248 -- Arizona	XMP-500 -- XOMA
UGIF -- Sheffield	XomaZyme-791 -- XOMA
UIC 2	XTL 001 -- XTL Biopharmaceuticals
UK 101	XTL 002 -- XTL Biopharmaceuticals
UK-279276 -- Corvas Intl.	yeast delivery system -- GlobelImmune
urodilatin -- Pharis	Yersinia pestis vaccine
urofollitrophin -- Serono	YIGSR-Stealth -- Johnson & Johnson
Urokinase -- Abbott	Yisum Project No. D-0460 -- Yisum
uteroferin-- Pepgen	YM 207 -- Yamanouchi
V 20 -- GLYCODESIGN	YM 337 -- Protein Design Labs
V2 vasopressin receptor gene therapy	Yttrium-90 labelled biotin
vaccines -- Active Biotech	Yttrium-90-labeled anti-CEA MAb T84.66 --
Varicella zoster glycoprotein vaccine --	ZD 0490 -- AstraZeneca
Research Corporation Technologies	ziconotide -- Elan
Varicella zoster virus vaccine live -- Cantab	ZK 157138 -- Berlex Laboratories
Pharmaceuticals	Zolimomab aritox
Vascular endothelial growth factor --	Zorcell -- Immune Response
Genentech, University of California	ZRXL peptides -- Novartis

FIG. 28CC